

Mutation within *TARDBP* Leads to Frontotemporal Dementia without Motor Neuron Disease

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ABSTRACT: It has been recently demonstrated that the 43-kDa transactive response (TAR)-DNA-binding protein (*TARDBP*) is the neuropathological hallmark of Frontotemporal Dementia (FTD) with ubiquitin-positive and tau-negative inclusions. Large series of FTD patients without motor neuron disease have been previously analysed, but no *TARDBP* mutation was identified. The aim of the present study was to evaluate whether *TARDBP* gene mutations may be associated with FTD. We report that a pathogenetic *TARDBP* mutation is causative of behavioural variant FTD (bvFTD). An aged woman in her seventies initially started to present apathy and depression associated with impairment in executive functions. The diagnosis of bvFTD (apathetic syndrome) was accomplished by three-year follow-up, and structural and functional neuroimaging. By five-years after onset, extensive electrophysiological investigations excluded subclinical motor neuron disease. In this patient, a single base substitution c.800A>G of *TARDBP* gene was identified. This mutation, already described as causative of ALS, predicted the amino acidic change arginine to serine at position 267 (N267S). *In silico* analysis demonstrated that this substitution generates a new phosphorylation site, and western blot analysis on lymphoblastoid cells reported a decrease of protein expression in N267S mutation carrier. Our study suggests that *TARDBP* mutations can be pathogenetic of bvFTD without motor neuron disease. *TARDBP* screening needs to be considered in FTD cases. © 2009 Wiley-Liss, Inc.

KEY WORDS: *TARDBP*; TDP-43; behavioral variant Frontotemporal Dementia; Frontotemporal Lobar Degeneration

INTRODUCTION

The 43-kDa transactive response (TAR)-DNA-binding protein (*TARDBP*; MIM# 605078) was recognized as the major constituent of ubiquitin-positive tau-negative neuronal and glial inclusions in brain autopsy of patients

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affected by Frontotemporal Dementia (FTD) with ubiquitin inclusions and Amyotrophic Lateral Sclerosis (ALS) [Neumann et al., 2006].

TDP-43 immunoreactive histopathology has also been found in a variable proportion of patients with Alzheimer's disease, with hippocampal sclerosis, with Pick's disease and in a subset of patients with Lewy Body related diseases, suggesting the presence of a nosographic entity collectively named as *TDP-43 proteinopathies* [Cairns et al., 2007; Probst et al., 2007; Cook et al., 2008; Dickson, 2008].

TDP-43 is a ubiquitously expressed and evolutionary conserved ribonucleoprotein, containing two RNA recognition motifs and a C-terminal glycine-rich domain, known to promote protein-protein interactions [Buratti and Baralle, 2008]. TDP-43 can bind to the common microsatellite region (GU/GT)_n in RNA and DNA, with a putative role in the regulation of transcriptional activity, of messenger RNA splicing, exon skipping and microRNA biogenesis of several genes. It has been proposed that in neurodegenerative conditions TDP-43 can be redistributed from the nucleus to the cytoplasm, sequestered into inclusions that are mainly composed of abnormally phosphorylated and C-terminally truncated TDP-43 fragments [Buratti and Baralle, 2008; Winton et al., in press].

Additional evidences for the pathogenic role of TDP-43 in neurodegeneration were provided by the identification of mutations within the *TARDBP* gene as causative of both familial and sporadic ALS [Kabashi et al., 2008; Rutherford et al., 2008; Sreedharan et al., 2008; Van Deerlin et al., 2008]. All mutations with just one exception predominantly affect the conserved C-terminal glycine-rich domain of TDP-43, likely impairing the protein interactions occurring in this domain [Kabashi et al., 2008; Rutherford et al., 2008; Sreedharan et al., 2008; Van Deerlin et al., 2008]. So far, mutations in the *TARDBP* gene account for 2-3% of ALS patients.

More recently, it has been reported that *TARDBP* mutations might be causative of FTD cases with motor neuron disease (MND) presenting with a behavioral variant FTD and semantic dementia [Benajiba et al., 2009]. Thus, it can be hypothesized that *TARDBP* mutations may contribute to the development of TDP-43 proteinopathies, beyond ALS. Large series of FTD patients without motor neuron disease have been previously analysed, but no *TARDBP* mutation was identified [Rutherford et al., 2008; Gijssels et al., 2009].

In the present work, we report the first case of behavioural variant FTD without MND carrying a pathogenetic mutation within *TARDBP*, namely the N267S (NP_031401.1: p.Asn267Ser).

Our results strongly suggest *TARDBP* mutations may be causative for FTD, and open a new chapter in the study of FTD pathogenesis, and FTD genetic screening.

MATERIALS AND METHODS

Index case.

We examined the index case performing neurological examination, formal standardized neuropsychological assessment at enrolment and periodically for three years. Brain Single Positron Emission Tomography (SPECT) was undertaken at baseline and analysed by Statistical Parametric Mapping (see Supporting Information) as well as a structural Magnetic Resonance Imaging (MRI). Three years after enrolment, a brain MRI, electroneurography (ENG), electromyography (EMG), motor-evoked potential (MEPs), and somatosensory-evoked potential (SSEPs) were assessed. Genealogic investigation was documented by the Family History Questionnaire.

Genetic Analyses

After obtaining informed consent from the patient and her caregiver, DNA was isolated according to standard procedures. Direct sequencing was performed on genomic DNA for both strands of exons of *TARDBP* [Rutherford et al., 2008], *Progranulin*, and *MAPT* genes by ABI 3130xl DNA analyzer (Applied Biosystems, Foster City, CA, USA), and analyzed using SeqScape Software version 2.6 (Applied Biosystems, Foster City, CA, USA).

The pathogenetic *TARDBP* mutation was confirmed by three independent analyses, collecting blood from the proband twice.

The following prediction programs have been used: polyphen (<http://genetics.bwh.harvard.edu/pph/>), SNAP (<http://cubic.bioc.columbia.edu/services/SNAP/>), PMUT (<http://mmb2.pcb.ub.es:8080/PMut/>), NetPhos 2.0 (<http://www.cbs.dtu.dk/services/NetPhos/>), and SIFT (<http://sift.jcvi.org>).

The reported *TARDBP* mutation was further excluded in a sample of 98 FTD patients, by direct sequencing of exon 6, and healthy controls.

Biological analysis.

After immortalization of lymphocytes (see Supporting Information), each cell pellet was re-suspended in 1 ml/3 millions of cells of RPMI 1640 complete medium with FCS 10%, and 1 ml cells aliquot per well into a 24 well plate (3×10^6 cells/well). The cells were maintained at 37° C in a CO₂ 5% incubator. After 1 month in culture, small to large clumps of cells began to emerge and were visible microscopically; cells were then expanded by transferring into T-25 or T-75 tissue culture flasks. The experiments were done in sterile conditions.

Lymphoblastoid preparations were resuspended in PBS plus Complete Protease Inhibitor Cocktail (Roche Applied Science) and briefly sonicated using a Soniprep 150 (Cellai). Samples were loaded on a 10% SDS-PAGE polyacrylamide gel and blotted on a Hybond-C Extra membrane (Amersham). TDP-43 was detected using a rabbit polyclonal antibody raised against a recombinant form of this protein as previously described [Buratti et al., 2001]. The same blot was stained with a mouse polyclonal antibody obtained at ICGEB against tubulin as a control for total protein loading. Protein bands were detected using ECL (Pierce) according to manufacturer's instructions.

RESULTS

Patient description

At age of 74 years, a right-handed woman with 5 years of education started to complain apathy and depression. She had no history for substance abuse and other forms of psychiatric or neurological illnesses. She had a history of diabetes and hypertension under treatment. On the basis of interviews with her relatives, no family history for dementia, MND or other neurodegenerative disorder was reported (see Supp. Figure S1 for pedigree).

Two years later, she was admitted at our clinic because of the worsening in mood disturbances, and personality change along with progressive withdrawn. The neurological examination was unremarkable, but she was severely apathetic and mildly depressed. Executive functions and comprehension of complex syntactic structures were impaired. She had no insight of the deficits (see Table 1, baseline).

Metabolic or infective causes of dementia were excluded by laboratory tests. Apolipoprotein E genotype was $\epsilon 3/3$.

Cognitive deficits at disease presentation were consistent with regional brain dysfunction as shown by SPECT scan (see Figure 1, panel A). Namely, there was a selective hypoperfusion in the dorsolateral frontal cortex, bilaterally ($x,y,z=-42,12,8$; $T=8.57$, cluster size=2375; $-6,56,2$, $T=4.53$, cluster size=1719; $38,26,-4$, $T=4.35$, cluster size=1685), and in the medial frontal cortex ($5,56,20$; $T=3.95$, cluster size=1719) (SPM2, $P<0.005$). Visual inspection of brain MRI showed bilateral frontotemporal atrophy (Figure 1, panel B).

Therefore, the diagnosis of behavioural variant FTD was made (apathetic syndrome) [Neary et al., 1998; McKhann et al., 2001].

Neuropsychological assessment at 3 point follow-up is reported in Table 1. By three years after first evaluation, the instrumental activities of daily living (IADL) progressively worsened, and at present she is completely dependent on IADL. Behavioural disturbances have persisted unchanged. Neuropsychological assessment showed progressive decline in language comprehension and in tests tapping executive functions.

Brain MRI performed at 3-year follow-up showed greater frontotemporal atrophy compared to baseline (Figure 1, panel C). At three-year follow-up, no clinical and electrophysiological signs of MND have been developed. The EMG recordings made from the extensor digitorum brevis muscle, brachial biceps muscle, deltoid muscle, anterior tibial muscle, femoral quadriceps muscle, bilaterally, showed no spontaneous activity (fibrillations, fasciculations, and serial discharges), and motor unit potentials were within normal range. ENG and SSEPs studies were unremarkable as well as MEPs (Supp. Table S1).

TABLE 1. Neuropsychological assessment of the proband at baseline and at follow-up.

Test	Baseline	1-year follow-up	2-year follow-up	3-year follow-up	Cut-off*
MMSE	20	20	19	16	24
IADL (lost)	5	5	7	8	0
BADL (lost)	2	2	3	3	0
NPI	30	22	24	22	0
FBI-AB	24	16	18	17	0
UPDRS-III	0	0	5	5	0
Story Recall test	9.5 (1)	10.0 (1)	9.5 (1)	11.5 (1)	7.5
Rey Figure, copy	4.7 (0)	5.2 (0)	4.5 (0)	0 (0)	28.75
Rey Figure, recall	n.a. (0)	n.a. (0)	n.a. (0)	n.a. (0)	9.46
Raven Colored Matrices	13 (0)	13 (0)	11 (0)	10 (0)	17.5
Digit Span	4.7 (3)	4.7 (3)	4.7 (3)	4.7 (3)	3.75
Token test	26.7 (1)	26.7 (1)	28.7 (1)	24.7 (0)	26.5
Fluency, letter	28 (3)	31 (3)	26 (2)	24 (2)	16
Fluency, category	24 (0)	26 (1)	26 (1)	25 (1)	24
Trail Making test, A	n.a. (0)	n.a. (0)	n.a. (0)	n.a. (0)	<93
Trail Making test, B	n.a. (0)	n.a. (0)	n.a. (0)	n.a. (0)	<282

MMSE: Mini-Mental State Examination; IADL: Instrumental Activities of Daily Living; BADL: Basic Activities of Daily Living; NPI: Neuropsychiatry Inventory; FBI-AB: Frontal Behavioural Inventory, Parts A and B; UPDRS: Unified Parkinson Disease Rating Scale.

Results are expressed as mean values, and equivalent scores between brackets; n.a.: patient was unable to perform the test. * cut-off scores according to Italian normative data.

Genetic analysis

In DNA from the patient, the single base substitution c.800A>G (NM_007375.3) of *TARDBP* gene predicted the amino acidic change arginine to serine at position 267 (N267S) (see Figure 2).

This mutation was already known to be pathogenic for ALS in an Italian patient [Corrado et al., 2009]. We further analysed 98 FTD (59.2% female, mean age=66.7±7.2; 2 patients with FTD-MND) and 106 healthy controls for this mutation, with negative results.

In silico analysis indicated that the substitution from arginine to serine generates a new phosphorylation site (NetPhos 2.0).

Furthermore, in the index case, no additional mutations were found within the *MAPT* and *Progranulin* gene. Progranulin dosage in the serum was within normal range (187 ng/mL), compared to a sample of FTLN patients carrying *Progranulin* mutations (data not shown).

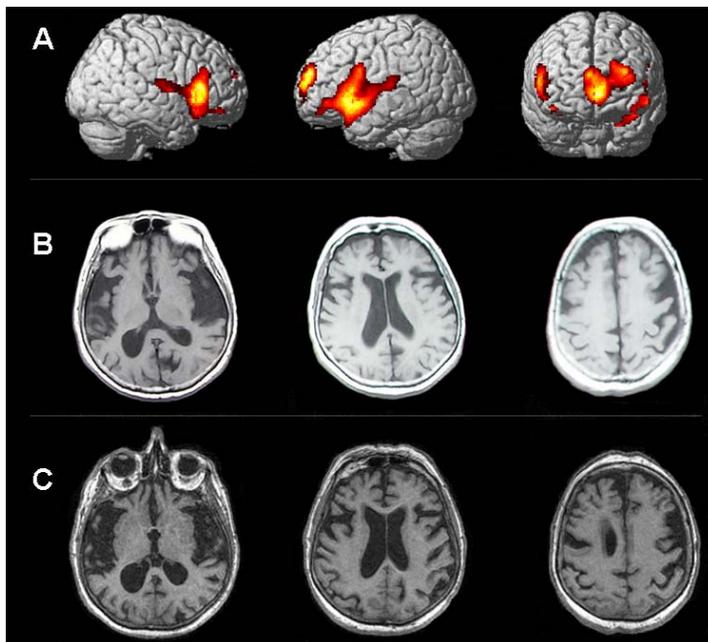


Figure 1. Brain functional and structural neuroimaging data of the proband at diagnosis, and at 3-year follow-up. **Panel A.** SPECT-Statistical Parametric Mapping (SPM) analysis performed at diagnosis, superimposed on 3D template. Index case showed a significant reduction of blood flow in the dorsolateral frontal cortex, bilaterally, and in the medial frontal cortex. **Panel B.** Axial MRI scans assessed at diagnosis. Frontotemporal atrophy was reported. **Panel C.** Axial MRI scans assessed at 3-year follow-up. Greater frontotemporal atrophy compared to baseline was reported.

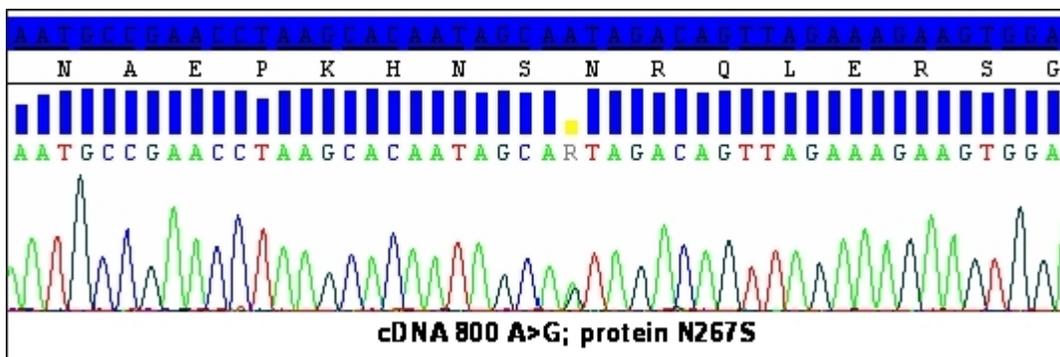


Figure 2. Chromatogram indicating proband's missense mutation. Missense mutation identified within *TARDBP* gene in the case index. The observed single base substitution c.800A>G, and predicted amino acid change arginine to serine at position 267 (N267S) are indicated in the chromatogram. cDNA numbering is according to the largest *TARDBP* transcript (NM_007375.3), and starting at the translation initiation codon. Protein numbering is relative to the largest TDP-43 isoform (NP_031401.1).

Biological analyses

In previous analyses, both Kabashi et al. [2008] and Rutherford et al. [2009] demonstrated the presence of increased TDP-43 degradation fragments with respect to control samples in cell lines derived from ALS patients carrying TDP-43 mutations: G348C, R361S, N390D/S, M337V, N345K, and I383V. A similar result was also obtained by direct analysis of frozen lymphocyte preparations from ALS patients carrying mutations S393L and A382T [Corrado et al., 2009]. Based on these observations, we performed a similar study on lymphoblastoid cells derived from this patient and three appropriate controls, including the healthy son. Compared to the previously published examples we did not detect an increased degradation pattern from the patient cell lines with respect to controls (see Figure 3). Instead, we detected a substantial drop in TDP-43 expression levels in the patient sample. A western blot directed specifically against a C-terminal fragment of TDP-43 did not also detect an increased amount of degraded TDP-43 in the MB patient sample (data not shown). Taken together, these data suggest that the N267S mutation has a direct and aberrant effect on TDP-43 cellular levels in these patient cells with respect to control samples.

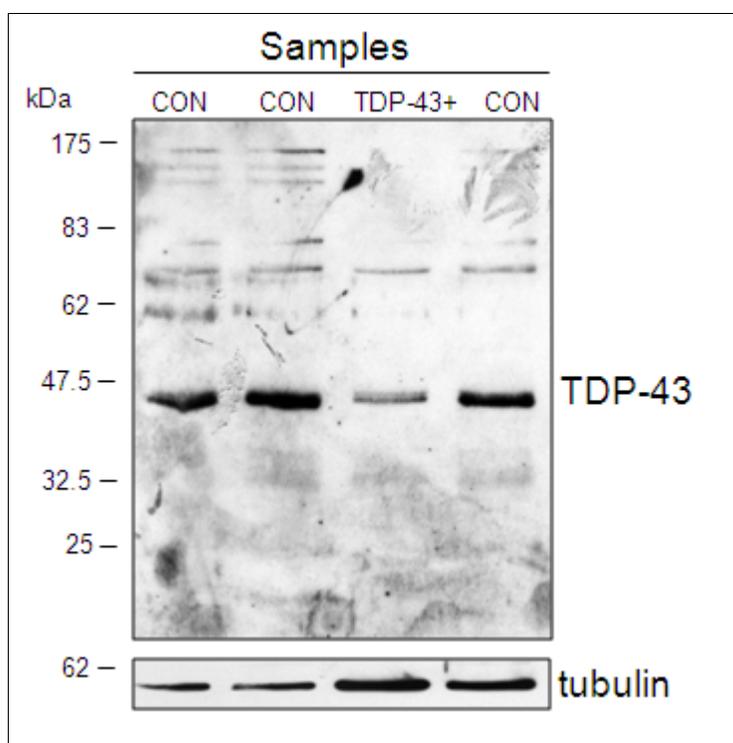


Figure 3. Western blot analysis of N267S lymphoblastoid cell line from patient and controls. Western blot analysis of TDP-43 protein from lymphoblastoid lysates of controls (CON) and the FTD patient carrying N267S mutation (lane TDP-43+). Samples were standardized using tubulin as an internal control. Similar results were obtained in two independent western blot experiments (data not shown).

DISCUSSION

The search for *TARDBP* mutations in affected FTD with ubiquitin inclusions and ALS patients has been quite intensive in order to establish a clear molecular connection with disease, and thus rule out the possibility that TDP-43 might be just a pathological curiosity [Rothstein, 2007].

Initial studies in FTD cohorts failed to identify a genetic association for the disease when analyzing common *TARDBP* variations, especially with regards to single-nucleotide-polymorphisms [Rollinson et al., 2007; Schumacher et al., 2009]. Nonetheless, despite the evidence of causative role of *TARDBP* mutations in approximately 2-3% of sporadic and familial forms of ALS [Banks et al., 2008; Corrado et al., 2009], monogenic forms of FTD has not been described yet. Up to now, two cases with FTD but with MND have been described [Benajiba et al., 2009].

In the present study, we report the first robust clinical evidence of a pathogenic mutation, namely N267S, in the *TARDBP* gene as causative of behavioural variant of FTD (apathetic syndrome) without MND. The same mutation was previously found as causative of a spinal form of ALS, and excluded in 771 neurologically healthy controls (1.542 chromosomes) [Corrado et al., 2009]. *In silico* analyses using a variety of prediction programs indicated that the substitution from arginine into serine may generate a new phosphorylation site; the novel creation of this site may alter the properties of TDP-43 or the interaction with other molecular partners. From a functional point of view, analysis of the patient lymphoblasts highlighted a substantial reduction on TDP-43 expression levels in patient's cells with respect to controls, suggesting that only the wild-type TDP-43 protein produced by the non-mutated allele can be observed in our analysis. Further work is currently in progress to better characterize the effects of the N267S mutation on impairing the cellular levels of TDP-43 inside the cells.

Up to now, the described *TARDBP* pathogenetic mutations cause MND with or without dementing illness [Kabashi et al., 2008; Rutherford et al., 2008; Sreedharan et al., 2008; Van Deerlin et al., 2008; Benajiba et al., 2009].

In our case, the pathogenic heterozygous N267S variation was found in an aged woman, who has displayed since her late seventies progressive behavioural disturbances along with cognitive deficits in executive functions. Functional brain imaging study was consistent with dorsolateral and frontal medial hypoperfusion, in accordance with previous work on apathetic presentation of bvFTD [Franceschi et al., 2005].

Extensive clinical investigations, three-year follow-up assessment, and electrophysiological studies excluded an evolution towards a form associated with MND. We cannot definitively exclude that the patient will develop MND over disease progression, but it is however true that she has had no preclinical signs or symptoms of MND by five years of estimated disease onset. The two patients reported by Benajiba and colleagues both presented with isolated FTD at onset, but they developed MND two years after [Benajiba et al., 2009].

Autopsy analyses will be needed to definitely clarify the neuropathology of our patient, but these data strikingly highlight the effect of a new genetic determinant in FTD.

There is evidence that neuropathologically TDP-43 disorders include ALS, FTD, and ALS with FTD [McCluskey et al., 2009]; accordingly, our study argues for a further extension of the pathogenetic role of *TARDBP* gene mutations in these neurodegenerative processes. This suggests that *TARDBP* mutation screening should be considered even in FTD cases without signs or symptoms of MND.

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SUPPORTING INFORMATION

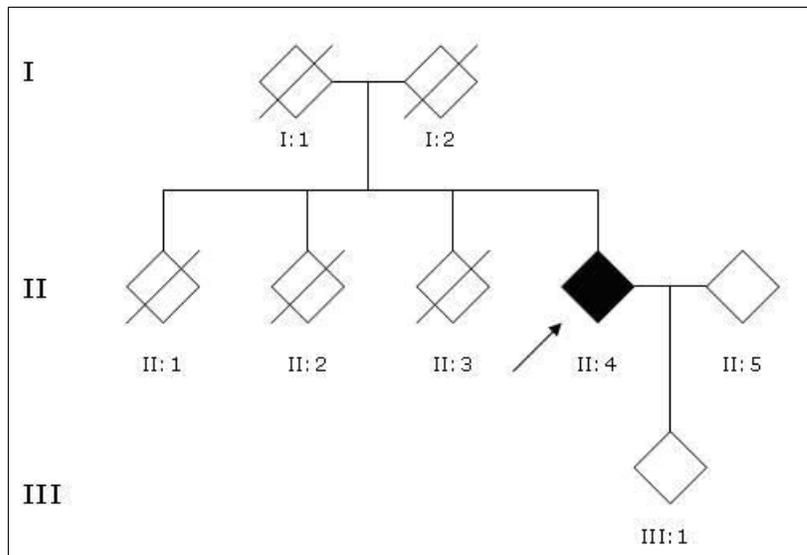
Methods

Regional Cerebral Blood Flow (rCBF) measurement.

SPECT imaging of rCBF for the index case and for 14 control subjects (mean age \pm SD=66.3 \pm 15.4) was performed using a maximum dose of 1110 MBq ^{99m}Tc -ethyl cysteinate dimer (ECD), injected intravenously, as previously published. Statistical Parametric Mapping (SPM2, Wellcome Department of Cognitive Neurology, University College, London), and Matlab 6.1 (Mathworks Inc., Sherborn, MA) were used for image pre-processing. Images were spatially normalized to a reference stereotactic template (Montreal Neurological Institute, MNI), and smoothed by a Gaussian kernel of 8x8x8mm FWHM.

Immortalization of lymphocytes.

The patients carrying *TARDBP* N267S mutation and three controls were considered. Blood drawing occurred while fasting at 9.00-10.00 am. A blood sample (10 ml) was taken, and collected into EDTA and NaCl. Blood underlayered with 5 ml Histopaque 1077 was centrifuged at 2000 rpm for 30 min at room temperature. The opaque lymphocyte layer along with the Histopaque, was collected. The cells were washed twice by repeated centrifugations in NaCl. The cells were re-suspended in 1 ml/3 millions of cells of EBV supernatant. After an incubation of 3 hours at 37° C with CO₂ 5%, a centrifugation at 1200 rpm for 10 min at room temperature was carried out.



Supp. Figure S1. Pedigree of the case index carrying TARDBP N267S mutation.

Solid symbol indicated the proband (II:4). To protect confidentiality, sex of individuals is disguised. The proband's parents (I:1 and I:2) died in their seventies for unknown causes; the three brothers died for cancer at 76 (II:1), 70 (II:2), and 53 (II:3) year-old. The proband's son (III:1) is in his fifties, and he does not complain of cognitive and behavioural disturbances.

Supp. Table S1. Motor evoked potential at 3 year follow-up (5 years from estimated onset of symptoms) showed values within normal range

Limb	Stimulation	Latency (msec)	Amplitude (mV)	CMCT (msec)	Cut-off values (msec)
Upper, right	Cervical	13.5	4.60	8.15	< 10
	Cortical	21.65			
Upper, left	Cervical	13.50	2.70	8.10	<10
	Cortical	21.60			
Lower, right	Cervical	14.80	3.80	14.00	<17
	Cortical	28.80			
Lower, left	Cervical	14.50	3.20	14.20	<17
	Cortical	28.70			

CMCT=Central Motor Conduction Time