



Article **Resolution of a Configurationally Stable Hetero**[4]helicene

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Abstract: We have developed an efficient chemical resolution of racemic hydroxy substituted dithiaaza[4]helicenes (DTA[4]H) **1(OH)** using enantiopure acids as resolving agents. The better diastereomeric separation was achieved on esters prepared with (1*S*)-(–)-camphanic acid. Subsequent simple manipulations produced highly optically pure (\geq 99% enantiomeric excess) (*P*) and (*M*)-**1(OH)** in good yields. The role of the position where the chiral auxiliary is inserted (*cape-* vs. *bay-zone*) and the structure of the enantiopure acid used on successful resolution are discussed.

Keywords: heterohelicene; chirality; resolution; enantiomers; chiroptical; screw-shaped compounds



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1. Introduction

Chirality is one of the most crucial assets of nature and is of paramount importance in several areas of science, technology and medicine. Molecular chirality has been recognized for a long time and has provided guidance in the design of drugs and functional materials. Furthermore, a smart combination of chiral phenomena and supramolecular chemistry resulted in an emerging interdisciplinary field called supramolecular chirality [1].

Helicenes are compounds with a screw-shaped skeleton formed by ortho-condensed (hetero)aromatic rings with a non-planar structure due to the steric superimposition of terminal rings or/and the substituents on these rings [2], which force the molecule to adopt a helical conformation (Figure 1). This important class of axially chiral compounds has a barrier of interconversion between M and P enantiomers increasing with the increase of the ring number forming the helicene backbone.







Circular dichroism (CD) and circularly polarized luminescence (CPL) are just a few of the chiroptical proprieties that make helicenes valuable in potential applications such as

advanced optical information storage, circularly polarized organic light-emitting diodes (CP-OLEDs), circularly polarized light detecting organic field effect transistors (CP-OFETs), chirality-induced spin selectivity (CISS) devices and stereoselective sensing chiroptical probes in biological processes [3–14].

Recently, increased attention has focused on the binding of small molecules to specific DNA structures to inhibit the biological functions in which these structures participate. Indeed, helicenes enantioselectivity offered a way to rationally design Z-DNA-depending inhibitors of biological functions [11].

Over the years, a variety of heterohelicenes and helical-shaped molecules, containing nitrogen, oxygen, sulfur, and other hetero-elements, have been synthesized and their unique properties studied. Great effort has been devoted to the setting of new simple and multi-gram synthetic procedures that allow for the isolation of helicenes in enantiopure form as required for various practical applications, including chirogenesis [7,15,16].

Dithia-aza[4]helicenes (DTA[4]H) **1** (Scheme 1) can be described as bis-phenothiazines with an aryl ring and a nitrogen atom in common forced into a helical shaped structure by the four long carbon–sulfur bonds. These [1,4]benzothiazino[2,3,4-*kl*]phenothiazines represent one of the attractive rare examples of geometrically stable [4]helicenes with racemization barriers higher than those measured for all carbon [5]helicenes.



Scheme 1. Synthetic pathways (A or B) to dithia-aza[4]helicenes (DTA[4]H) 1.

DTA[4]H are obtained [17–21], as racemic mixture, from properly substituted triarylamines (TAA) **2** or *N*-aryl phenothiazines (PTZ) **3** (Scheme 1, pathway A and B respectively), through regioselective sulfenylation(s) with two or one equivalents of phthalimidesulfenyl chloride (PhtNSCl (4), Pht = phthaloyl). Reacting the resulting sulfenylated derivatives **5** or **6** with a Lewis Acid (L.A.), typically BF₃OEt₂ or AlCl₃, causes two or one electrophilic intramolecular cyclization with formation of helicenes **1**.

Along with their peculiar helical-shaped structure, derivatives **1** show a very good one-electron donor ability, being easily, and reversibly, oxidized to the corresponding exceptionally stable crystalline chiral radical cations $1^{\bullet+}$ [18,21] (Scheme 2). We have also demonstrated that the oxidative process is extremely sensitive to the medium, and under acidic conditions, molecular oxygen becomes an efficient single electron transfer (SET) oxidant, giving rise to the formation of $1^{\bullet+}$. Furthermore, radical cations $1^{\bullet+}$ can

AgSbF₆ or (O_2/H^+) or (hv/PhCl)

be generated also via irradiation at 240–400 nm of helicenes in the presence of PhCl [21] (Scheme 2).



DTA[4]H

Na₂S₂O₄ or base

Indeed, we have also prepared, via ring-opening metathesis polymerization, dithiaaza[4]helicene functionalized polynorbornenes showing a pH depending reversible redox behavior as a new class of tunable material reversibly switchable by pH-triggered redox processes [22].

1.

The availability of differently functionalized enantiomerically pure helicenes, avoiding the limitation associated with chiral HPLC resolution [17,23] is mandatory for the development of the appealing applications of these peculiar systems [21,22,24]. Therefore, in recent years we tried to set off regio-, stereo- and enantioselective synthetic approaches for the preparations of **1**.

Actually, the synthetic procedure depicted in Scheme 1, while failing to control the absolute stereochemistry of the process, allowed for the control of the regiochemistry during ring closure as well as the possibility of inserting different substituents in specific positions. Thus, we have studied the chemical resolutions of 1 using the classical temporary insertion of chiral auxiliaries.

The DTA[4]H **1** derivatives requested for the above described applications require the insertion, as an anchoring unit, of a hydroxyl group in different positions of the helical backbone. Thus, we decided to take advantage of these phenolic groups for the introduction of chiral auxiliaries through esterification with enantiopure acids **7** and in order to verify whether the diastereoisomeric mixture of esters **8** obtained can be separated. Herein we report how the helicene topology and chiral auxiliary structure could be matched to allow the resolution of phenolic DTA[4]H **1(OH)** (Scheme 3).



Scheme 3. Esterification with enantiopure acids 7.

2. Results and Discussion

Selected DTA[4]H 1 can be resolved through HPLC in the chiral stationary phase as we previously reported [17,23]; however, this method is unsuitable to obtain enantiopure DTA[4]H in multigram scale. Instead, the diastereomeric process-based resolution has advantages in the the viewpoint of cost, generality and the amount of enantiopure products achieved. Thus, we decided to study the insertion of chiral auxiliaries, for example through esterification reactions, to verify whether the mixture of diastereomers obtained can be separated by flash chromatography allowing isolation of pure M and P DTA[4]H in relevant quantity.

We have demonstrated that *N*-arylphenothiazines PTZ **3** are suitable substrates for the synthesis of unsymmetrically hydroxy substituted helicenes **1(OH)** [20]. We selected helicene **1a(OH)** and **1b(OH)** (Scheme 4) to prepare diastereoisomeric esters using enantiopure acid **7** (Scheme 5).







Scheme 5. Esterification of helicenes 1a(OH) and 1b(OH), panels (A,B), respectively, with enantiopure acids 7.

Firstly, we planned the introduction of chiral auxiliaries in 2-hydroxy-substituted ADT[4]H **1a(OH)** presenting a hydroxyl group in the 2-position (that we indicate as *cape-zone*) of the helicene, which was relatively easy to prepare [20].

Racemic **1a(OH)** was esterified with different enantiopure acids **7a–h** yielding a diastereomeric mixture (D1+D2) of esters **8a(a–h)**. Esterification reactions were carried out in presence of diisopropylcarbodiimide (DIC) and 4-dimethylaminopyridine (DMAP) as catalysts, in dry CH_2Cl_2 at room temperature (Scheme 5A). Regardless, all chiral acids **7a–h** (Scheme 5 panel A and Table 1) allowed the formation of diastereomeric esters **8a(a–h)** (Scheme 5 panel A and Table 1) in good yields, in none of these cases it was possible to separate the diastereoisomeric mixtures by flash chromatography or crystallization.

Table 1. Diastereomeric esters 8 obtained reaction racemic phenols 1a(OH) or 1b(OH) with enantiopure acids 7a–h.

Chiral Auxiliary	Product	Yield	Resolution
$HO = \frac{1}{10}$	8aaD1/8aaD2	60%	No resolution
7b Camphorsulfonyl chloride	8abD1/8abD2	56%	No resolution
^{ال} وب الموالح الموالح (−)-o,o'-Dibenzoyl-L-tartaric acid mono(dimethyl amide)	8acD1/8acD2	56%	No resolution
но	8adD1/8adD2	79%	No resolution
ة من المعنى '''' 7d (1 <i>S</i>)-(+)-Ketopinic acid	8bdD1/8bdD2	55%	No resolution
7e (S)-(+)-2-(6-Methoxy-2- naphthyl)propionic	8aeD1/8aeD2 8beD1/8beD2	94% 58%	No resolution No resolution
acid			
	8afD1/8afD2	54%	No resolution
7f (-)mono-(1R)-Menthylphthalate	8bfD1/8bfD2	45%	No resolution
	8agD1/8agD2	69%	No resolution
^{Ho} من N-Boc pipecolic acid	8bgD1/8bgD2	28%15%	Chromatographic resolution is possible
Sm.PK	8ahD1/8ahD2	57%	No resolution
^{но о} - <u>(</u> 7 h (1 <i>S</i>)-(–)-Camphanic acid	8bhD1/8bhD2	37% 33%	Chromatographic resolution is possible

We thought, as suggested by literature data [25–38], that functionalization of the *cape-zone* position keeps the chiral auxiliaries too far from the superimposition area of the terminal aryl rings vanishing separation. Thus, we moved to helicene **1b(OH)**, which allowed for the insertion of chiral auxiliaries on 1-position, *ortho* to the nitrogen atom, which we indicated as *bay-zone*, i.e., exactly in the area of terminal ring superimposition (Scheme 5B).

Using chiral acids **7d–h** (Scheme 5 panel B and Table 1), the corresponding diastereomeric esters **8b(d–h)** (Scheme 5 panel B and Table 1)were obtained in moderate yields generally lower than those of the corresponding esters prepared using phenol **1a(OH)**, indicating, as expected, a more difficult access to the OH group of **1b(OH)** laying the *bay-zone* (Table 1). For several esters **8b** it was possible to identify the presence of the



two diastereomers (D1 and D2, chromatographic elution order) by ¹H and ¹³C NMR, and, eventually, to reveal a slightly different chromatographic behavior on TLC (Figure 2).

Figure 2. ¹H NMR spectrum of diastereomers (D1 and D2) **8bh**.

Despite the introduction of the chiral auxiliary in the *bay-zone* diastereoisomeric esters D1 and D2 of **8b(d–f)** were not separable by flash chromatography in spite of an accurate selection of eluent mixtures. However, esterification of **1b(OH)** with N-boc pipecolic acid **7g** allowed for the partial separation by flash chromatography on silica gel of diastereomers **8bgD1** and **8bgD2**, which were characterized by ¹H and ¹³C NMR. Optical rotation of **8bgD1** was: $[\alpha]_D^{20} -157$ (*c* 0.1, CH₂Cl₂), while for **8bgD2** was: $[\alpha]_D^{20} +49$ (*c* 0.1, CH₂Cl₂). ¹H NMR spectra show that the product **8bgD1** was isolated as single diastereomers, while the product **8bgD2** was isolated as a roughly 3:1 mixture of the two diastereomers.

Esters **8bgD1** and **8bgD2** were hydrolysed with 3 eq. of NaOH in CH₂Cl₂/MeOH to give enantiomeric phenols (*M*)-**1b(OH)** and (*P*)-**1b(OH)**. Phenols were analysed by HPLC in the chiral stationary phase in order to calculate the enantiomeric ratio. Chromatograms showed that product (*M*)-**1b(OH)** $[\alpha]_D^{20} - 161$ (*c* 0.1, CH₂Cl₂) was obtained as single enantiomer (e.e. \geq 99%), while helicene (*P*)-**1b(OH)** $[\alpha]_D^{20} + 75$ (*c* 0.1, CH₂Cl₂) exhibits the enantiomeric ratio 72:28 (e.e. = 44%).

Esterification of **1b(OH)** with (1*S*)-(–)-camphanic acid **7h** provided, with our great satisfaction, diastereomers **8bhD1** and **8bhD2** that were successfully separated by flash chromatography and characterized by ¹H and ¹³C NMR spectroscopy (Figure 3 and Supplementary Materials).

Optical rotation was measured and gave $[\alpha]_D^{20} - 129$ (*c* 0.1, CH₂Cl₂) for **8bhD1** and $[\alpha]_D^{20} + 126$, (*c* 0.1, CH₂Cl₂) for **8bhD2**. Hydrolysis of diastereometric esters provided helicenes (*M*)-**1b(OH)** and (*P*)-**1b(OH)**, respectively. HPLC analysis with a chiral stationary phase showed that the first eluted product, (*P*)-**1b(OH)** $[\alpha]_D^{20} + 166$ (*c* 0.1, CH₂Cl₂), exhibits an enantiometric ratio = 98:2 (e.e. = 96%), while the second eluted product, (*M*)-**1b**(OH) $[\alpha]_D^{20} - 167$ (*c* 0.1, CH₂Cl₂), was obtained as a single enantiometric (e.e. \geq 99%), (Scheme 6).



Figure 3. ¹H-NMR spectra of diastereomers 8bhD1 and 8bhD2.



Scheme 6. Chemical resolution of helicene (rac)-1b(OH).

The assignment of the absolute configuration of DTA[4]H **1** derivatives has been established as P-(+) and M-(-), which is typical for helicene systems; opposite assignment is quite unusual, as we have established in ref [17,19]. In this work the absolute configuration of **1b(OH)** was validated by comparison of the electronic circular dichroism (ECD) spectra of the two optical enantiomers-(+)-**1b(OH)** and (-)-**1b(OH)**, assigned to (P)-**1b(OH)** and (M)-**1b(OH)**, with the calculated spectrum of the M structure.

In order to assign the configuration, DFT and TD-DFT calculations have been conducted with the Gaussian16 package [39]. Two orientations are possible for the hydroxylgroup; the two optimized structures in the *M* configuration are reported in Figure 4 with their Boltzmann populations. Two functionals have been considered; the two differing in the amount of the exact exchange included M06 with 27% HF exchange and M06-2X with 54% HF exchange [40]. The solvent has been treated at the iefpcm level. Structural results are similar for the two functionals.



Figure 4. Optimized 3D-structures for the two possible conformers of (*M*)-**1b(OH)**. Percent population factors calculated at M06/cc-pvtz, iefpcm level; in parenthesis population factors calculated at M062X/cc-pvtz, iefpcm level.

CD and absorption spectra have been calculated at the same level of theory, a constant Gaussian 0.2 eV bandwidth was applied to each transition. The experimental CD and absorption spectra have been recorded for the two enantiomers in 4.2 mM dichloromethane solution in a 0.1 mm quartz cuvette using a JASCO-815SE instrument.

The comparison of experimental data with calculations are presented in Figure 5. In order to compare with data, +4 nm shift has been applied to the results obtained with M06, +26 nm with M06-2X; calculation of similarity index between experimental and calculated spectra suggested the best shift for the best correspondence, as reported

in the Supplementary Materials paragraph. It is also shown that the two conformers give very similar spectra, while in Figure 5 the Boltzmann weighed average is presented. Both functionals permit confirmation of the configuration as M-(-) (correspondingly P-(+)). This conclusion agrees with what was obtained for the parent molecule triarylamine hetero[4]helicene examined in reference [19].



Figure 5. CD (top) and absorption (bottom) experimental and calculated spectra with two choices for the DFT functional (see text); calculation performed on *M*-**1b(OH)**. The calculated spectra are Boltzmann weighed averages of the two conformers A and B.

Overall, our results confirm that, on chemical resolution of helicenes, the position where the chiral auxiliaries are inserted is extremely important, being the *bay-zone* that allows for the higher effect on enantiomeric discrimination. At the same time we have confirmed previous studies reporting chromatographic resolutions of [7]carbo- and [7]hetero-helicenes by means of tetra- and monocamphanate esters [25–31]. In each of these cases, the (1*S*)-camphanate of the (*P*)-helicenol moves more slowly upon chromatography on silica gel than the (1*S*)-camphanate of the (*M*)-helicenol [27].

3. Materials and Methods

¹H and ¹³C NMR spectra were recorded with Varian Mercury Plus 400, Varian Inova 400, using CDCl₃ as solvent. Residual CHCl₃ at δ = 7.26 ppm and central line of CDCl₃ at δ = 77.16 ppm were used as the reference of ¹H-NMR spectra and ¹³C NMR spectra,

respectively. FT-IR spectra were recorded with a Spectrum Two FT-IR Spectrometer. ESI-MS spectra were recorded with a JEOL MStation JMS700. Melting points were measured with a Stuart SMP50 Automatic Melting Point Apparatus. Optical rotation measurements were performed on a JASCO DIP-370 polarimeter (JASCO, Easton, MD, USA) and the specific rotation of compounds was reported [41].

All the reactions were monitored by TLC on commercially available precoated plates (silica gel 60 F 254) and the products were visualized with acidic vanillin solution. Silica gel 60 (230–400 mesh) was used for column chromatography. Dry solvents were obtained by The PureSolv Micro Solvent Purification System. Chloroform was washed with water several times and stored over calcium chloride. Pyridine and TEA were freshly distilled from KOH. Phthalimide sulfenyl chloride was prepared from the corresponding disulfide (purchased from Chemper snc) as reported elsewhere. Helicenes **1a** and **1b** were described elsewhere [17].

General Procedure for the synthesis of diastereomeric esters from 1 by Steglich esterification: To a solution of 1 in dry CH_2Cl_2 (roughly 0.03–0.04 M), the enantiopure acid 7 (1.2 eq), DMAP (0.1 eq) and DIC (1–1.2 eq) were added at 0 °C. The solution was stirred at room temperature under a nitrogen atmosphere for 2–29 h, then was diluted with CH_2Cl_2 (60 mL), washed with a saturated solution of NH_4Cl (2 × 40 mL), with a saturated solution of $NaHCO_3$ (3 × 40) then with NH_4Cl (3 × 40 mL). The organic layer was dried over Na_2SO_4 , filtered and evaporated under reduced pressure. The crude was purified by flash chromatography on silica gel.

Diastereoisomers **8aaD1** and **8aaD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-*kl*]pheno thiazine-2-yl (*S*)-perillate. Following the general *Steglich esterification* procedure from **1a(OH)** (60 mg, 0.18 mmol) and (*S*)-(–)-perillic acid **7a** (36 mg, 0.22 mmol), kept for 22 h at rt. The crude was purified by flash chromatography on silica gel (petroleum ether/ CH₂Cl₂ 1:3, *Rf* 0.86) to afford the mixture of the two diastereomeric compounds **8aaD1** and **8aaD2** (52 mg, 60% yield) as a white solid (mp 105–115 °C). ¹H NMR (400 MHz, CDCl₃)* δ : 1.45-1.55 (m, 2H), 1.76 (s, 6H), 1.89–1.95 (m, 2H), 2.12 (s, 6H), 2.16–2.41 (m, 8H), 2.51–2.59 (m, 2H), 4.74 (bs, 2H), 4.78 (bs, 2H), 6.89 (bs, 2H) 6.92–7.05 (m, 8H), 7.06 (bs, 2H), 7.12–7.25 (m, 8H) ppm. ¹³C NMR (100 MHz, CDCl₃)* δ : 15.8, 20.9, 24.76, 24.80, 27.1, 31.4, 40.1, 108.63, 108, 64, 110.0, 110.6, 114.7, 120.6, 124.1, 124.8, 125.0, 125.7, 125.75, 125.83, 126.0, 127.0, 127.3, 127.7, 128.0, 129.3, 129.5, 139.5, 141.2, 141.8, 142.5, 148.7, 149.2, 165.2, 165.3 ppm (34 signals for 58 different carbons). Elem. Anal. for C₂₉H₂₅NO₂S₂: Calcd. C 72.02, H 5.21, N 2.90; found C 71.80; H 5.21, N 2.89. *Et₃N was added to neutralize CHCl₃ acidity.

Diastereoisomers **8abD1** and **8abD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-kl]pheno thiazine-2-yl (1S)-10-camphorsulfonate. To a solution of 1a(OH) (60 mg; 0.18 mmol) and TEA (22 mg, 0.22 mmol) in 4 mL of dry CH_2Cl_2 , (1S)-(+)-10-camphorsulfonyl chloride 7b (51 mg, 0.20 mmol) is added at 0 °C. After 10 min the solution was allowed to warm at room temperature and was stirred for 18 h under a nitrogen atmosphere. The mixture was diluted with AcOEt (25 mL) and washed with water (3 \times 20 mL). The organic layer was dried over Na₂SO₄, filtered, and then evaporated under reduced pressure. The crude was purified by flash chromatography on silica gel (petroleum ether/AcOEt: 5/1, Rf 0.38) to afford the mixture of the two diastereomeric compounds 8abD1 and 8abD2 (55 mg, 56% yield) as a white solid (mp 190–195 °C). ¹H NMR (400 MHz, CDCl₃) δ: 0.87 (s, 3H), 0.88 (s, 3H), 1.11 (s, 6H), 1.40–1.47 (m, 2H), 1.64–1.72 (m, 2H), 1.92–1.98 (m, 2H), 2.00–2.13 (m, 4H), 2.31 (s, 3H), 2.32 (s, 3H), 2.36–2.57 (m, 4H), 3.13–3.18 (m, 2H), 3.73–3.78 (m, 2H), 6.93–7.25 (m, 18H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ: 11.6, 16.4, 16.5, 19.8, 20.02, 20.03, 25.3, 26.9, 27.0, 42.5, 43.0, 43.1, 46.3, 48.01, 48.03, 48.4, 48.6, 58.19, 58.24, 114.7, 114.8, 120.45, 120.50, 125.1, 125.2, 125.4, 125.5, 125.7, 125.8, 125.9, 126.0, 126.1, 126.2, 127.1, 127.8, 127.9, 128.11, 128.13, 128.2, 128.4, 130.0, 130.1, 139.2, 141.29, 141.32, 142.26, 142.29, 147.0, 147.1, 213.8, 213.9 ppm. Elem. Anal. for C₂₉H₂₇NO₄S₃: Calcd. C 63.36, H 4.95, N 2.55; found C 63.38, H 4.95, N 2. 54.

Diastereoisomers **8acD1** and **8acD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-kl]pheno thiazine-2-yl o,o'-dibenzoyl-L-tartrate. Following the general procedure from **1a(OH)**

(85 mg, 0.25 mmol) and (-)-o,o'-dibenzoyl-L-tartaric acid mono(dimethyl amide) **7c** (117 mg, 0.30 mmol), kept for 2 h at room temperature. The crude was purified by flash chromatography on silica gel (AcOEt/CH₂Cl₂ 1/20, *Rf* 0.65) to afford the mixture of the two diastereomeric compounds **8acD1** and **8acD2** (99 mg, 56% yield) as a white solid (mp 102–106 °C). ¹H NMR (400 MHz, CDCl₃) δ : 2.09 (s, 3H), 2.10 (s, 3H), 2.92 (s, 3H), 2.93 (s, 3H), 3.17 (s, 3H), 3.30 (s, 3H), 6.16 (d, 1H, *J* = 4.8 Hz), 6.20 (d, 1H, *J* = 5.2 Hz), 6.31 (d, 1H, *J* = 4.8 Hz), 6.33 (d, 1H, *J* = 5.2 Hz), 6.83–7.20 (m, 18H), 7.44–7.54 (m, 8H), 7.54–7.58 (m, 4H), 7.99–8.08 (m, 8H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ : 15.61, 15.63, 36.27, 36.31, 37.2, 69.5, 69.6, 70.9, 113.8, 114.0, 120.35, 120.41, 124.8, 124.9, 125.0, 125.1, 125.5, 125.6, 125.72, 125.74, 125.86, 125.92, 126.75, 126.84, 126.9, 127.6, 127.7, 127.9, 128.0, 128.35, 128.43, 128.52, 128.53, 128.58, 128.59, 128.60, 129.58, 129.59, 130.0, 130.09, 130.12, 130.15 133.82, 133.86, 139.12, 139.13, 141.1, 141.2, 142.1, 142.3, 148.09, 148.10, 165.0, 165.1, 165.3, 165.43, 165.44, 165.45, 165.50 ppm (59 signals for 78 carbons). IR (ATR solid) n: 3063, 2929, 1763, 1724, 1664, 1477, 1432, 1240 cm⁻¹. Elem. Anal. for C₃₉H₃₀N₂O₇S₂: Calcd. C 66.65, H 4.30, N 3.99; found C 66.61, H 4.28, N 4.00.

Diastereoisomers **8adD1** and **8adD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-*k*]pheno thiazine-2-yl (1*S*)-ketopinate. Following the general *Steglich esterification* procedure from **1a(OH)** (60 mg, 0.18 mmol) and (1*S*)-(+)-ketopinic acid **7d** (56 mg, 0.31 mmol), kept for 29 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/CH₂Cl₂ 1/3, *Rf* 0.45) to afford the mixture of the two diastereomeric compounds **8adD1** and **8adD2** (71 mg, 79% yield) as a white solid (mp 130–133 °C). ¹H NMR (400 MHz, CDCl₃)* δ : 1.14 (s, 6H), 1.21 (s, 3H), 1.22 (s, 3H), 1.42–1.48 (m, 2H), 1.88–1.97 (m, 3H), 2.01–2.10 (m, 3H), 2.13–2.15 (m, 2H), 2.18 (s, 3H), 2.19 (s, 3H), 2.40–2.48 (m, 2H), 2.54–2.60 (m, 2H), 6.85 (s, 1H), 6.87 (s, 1H), 6.94–7.06 (m, 10H), 7.12–7.23 (m, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃)* δ : 16.2, 20.0, 21.50, 21.52, 26.49, 26.52, 26.7, 26.8, 44.02, 44.58, 44.61, 49.51, 49.56, 68.20, 68.24, 110.6, 114.70, 114.72, 120.41, 120.44, 124.6, 124.8, 124.9, 125.0, 125.70, 125.72, 125.8, 126.1, 126.8, 127.2, 127.3, 127.75, 127.82, 128.1, 129.59, 129.61, 139.4, 140.9, 141.0, 142.66, 142.71, 148.8, 168.1, 210.3 ppm (44 signals for 58 different carbons). Elem. Anal. for C₂₉H₂₅NO₃S₂: Calcd. C 69.71, H 5.04, N 2.80; found: C 69.73, H 5.01, N 2.77. *Et₃N was added to neutralize CHCl₃ acidity.

Diastereoisomers **8aeD1** and **8aeD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-*k*]pheno thiazine-2-yl (*S*)-2-(6-methoxy-2-naphthyl) propionate. Following the general *Steglich esterification* procedure from **1a(OH)** (60 mg, 0.18 mmol) and (*S*)-(+)-2-(6-methoxy-2-naphthyl) propionic acid **7e** (50 mg, 0.22 mmol), kept for 20 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/CH₂Cl₂ 1/3, *Rf* 0.80) to afford the mixture of the two diastereomeric compounds **8aeD1** and **8aeD2** (94 mg, 94% yield) as an orange solid (mp 132–136 °C). ¹H NMR (400 MHz, CDCl₃)* δ : 1.39 (d, 6H, *J* = 6.5 Hz), 1.77 (s, 3H), 1.79 (s, 3H), 3.39 (s, 6H), 3.84 (q, 1H, *J* = 6.6 Hz), 4.12 (q, 1H, *J* = 6.7 Hz), 6.81 (bs, 2H), 6.91–7.05 (m, 10H), 7.10–7.20 (m, 10H), 7.44–7.47 (m, 2H), 7.67–7.73 (m, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃)* δ : 15.46, 15.48, 18.5, 18.6, 20.6, 21.2, 22.7, 23.6, 42.3, 45.56, 45.59, 55.4, 105.7, 108.6, 110.6, 114.4, 114.5, 119.3, 120.5, 124.3, 124.8, 125.0, 125.7, 125.8, 125.96, 126.01, 126.3, 126.4, 126.9, 127.1, 127.2, 127.4, 127.7, 127.8, 127.98, 128.01, 129.0, 129.4, 129.5, 133.9, 134.95, 134.99, 139.4, 141.0, 142.50, 142.54, 148.9, 157.9, 172.7 ppm (49 signals for 66 different carbons). Elem. Anal. for C₃₃H₂₅NO₃S₂: Calcd. C 72.37, H 4.60, N 2.56; found C 72.29; H 4.58, N 2.57. *Et₃N was added to neutralize CHCl₃ acidity.

Diastereoisomers **8afD1** and **8afD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-*kl*]pheno thiazine-2-yl mono-(1*R*)-menthylphthalate. Following the general procedure from **1a(OH)** (70 mg, 0.21 mmol) and (-)mono(1*R*)-menthylphthalate **7f** (76 mg, 0.25 mmol), kept for 24 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/ CH₂Cl₂ 2/1, *Rf* 0.78)) to afford the mixture of the two diastereomeric compounds **8afD1** and **8afD2** (70 mg, 54% yield) as a white solid (mp 140.8–146.8 °C). ¹HNMR (400 MHz, CDCl₃) δ : 0.73–0.69 (m, 6H), 0.83–0.89 (m, 14H), 0.97–1.16 (m, 4H), 1.36–1.51 (m, 4H), 1.64–1.71 (m, 4H), 1.85–1.98 (m, 2H), 2.05–2.11 (m, 2H), 2.21 (s, 3H), 2.22 (s, 3H), 4.82–4.89 (m, 2H), 6.93–7.00 (m, 6H), 7.03 (dd, 2H, *J* = 1.2 Hz, *J* = 7.5 Hz), 7.08 (bs,

1H), 7.09 (bs, 1H), 7.11–7.16 (m, 2H), 7.18 (d, 2H, J = 7.7 Hz), 7.24–7.29 (m, 2H), 7.54–7.60 (m, 4H), 7.68–7.73 (m, 2H), 7.85–7.90 (m, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ : 15.19, 16.0, 16.3, 16.5, 20.89, 20.93, 22.15, 22.18, 23.5, 23.6, 26.3, 26.4, 31.55, 31.57, 34.37, 34.41, 40.6, 40.7, 47.23, 47.25, 75.95, 75.02, 114.47, 114.49, 120.60, 120.64, 124.5, 124.6, 124.9, 125.0, 125.71, 125.75, 125.77, 125.79, 126.07, 126.08, 126.95, 126.99, 127.33, 127.34, 127.71, 127.75, 127.98, 128.00, 128.8, 129.0, 129.51, 129.52, 129.56, 129.60, 130.6, 130.89, 130.94, 131.0, 131.8, 131.9, 133.5, 133.7, 139.5, 141.2, 142.50, 142.52, 148.97, 148.99, 165.3, 165.4, 166.8, 166.9 ppm (68 signals for 74 different carbons). IR (ATR solid) n: 2953, 2923, 2867, 1749, 1715, 1477, 1431, 1276, 1236 cm⁻¹. Elem. Anal. for C₃₇H₃₅NO₄S₂: Calcd. C 71.47, H 5.67, N 2.25; found C 71.36, H 5.65, N 2.26.

Diastereoisomers **8agD1** and **8agD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-*kl*]pheno thiazine-2-yl (S)-N-Boc pipecolinate. Following the general procedure from **1a(OH)** (61 mg, 0.18 mmol) and (S)-N-Boc pipecolic acid **7g** (50 mg, 0.22 mmol) kept for 22 h at room temperature. The crude was purified by flash chromatography (CH₂Cl₂, *Rf* 0.51) on silica gel to afford the mixture of the two diastereomeric compounds **8agD1** and **8agD2** (70 mg, 69%yield) as a white solid (mp 106–117 °C). ¹H NMR (400 MHz, CDCl₃)* δ : 1.37–1.44 (m, 22H), 1.65–1.80 (m, 8H), 2.13 (s, 6H), 2.29–2.33 (m, 2H), 2.93–3.09 (m, 2H), 3.89–3.95 (m, 1H), 4.04–4.09 (m, 2H), 4.92 (bs, 1H), 5.07 (bs, 1H), 7.21–6.77 (m, 18H) ppm. Elem. Anal. For C₃₀H₃₀N₂O₄S₂: Calcd. C 65.91, H 5.53, N 5.12; found: C 65.35, H 5.31, N 4.98. *Et₃N was added to neutralize CHCl₃ acidity.

Diastereoisomers **8ahD1** and **8ahD2**. (*M*/*P*)-3-methyl[1,4]benzothiazino[2,3,4-*kl*]pheno thiazine-2-yl (1*S*)-camphanate. Following a *Steglich esterification* procedure from **1a(OH)** (60 mg, 0.18 mmol) and (1*S*)-(–)-camphanic acid **7h** (43 mg, 0.22 mmol), kept for 17 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/AcOEt/diethyl ether: 10:1:3, *Rf* 0.43)) to afford the mixture of the two diastereomeric compounds **8ahD1** and **8ahD2** (52 mg, 57% yield) as a white solid (mp 170 °C dec). ¹H NMR (400 MHz, CDCl₃)* δ : 1.04 (s, 3H), 1.06 (s, 3H), 1.11 (s, 6H), 1.13 (s, 3H), 1.14 (s, 3H), 1.69–1.77 (m, 2H), 1.92–1.99 (m, 2H), 2.11–2.19 (m, 8H), 2.45–2.54 (m, 2H), 6.84 (s, 1H), 6.86 (s, 1H), 6.93–7.00 (m, 6H), 7.02–7.08 (m, 4H), 7.12–7.21 (m, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃)* δ : 9.78, 9.81, 16.0, 16.2, 16.94, 16.96, 17.01, 17.04, 29.0, 31.2, 31.3. 53.6, 54.6, 54.97, 55.00, 90.89, 90.94, 114.16, 114.21, 120.4, 120.5, 125.0, 125.1, 125.24, 125.27, 125.6, 125.8, 125.9, 126.0, 126.1, 126.50, 126.52, 126.9, 127.0, 127.8, 128.1, 129.8, 139.3, 141.2, 141.3, 142.4, 142.5, 148.1, 148.2, 165.8, 165.9, 177.8 ppm (47 signals for 58 different carbons). Elem. Anal. for C₂₉H₂₅NO₄S₂: Calcd. C 67.55, H 4.89, N 2.72; found C 67.49, H 4.88, N 2.72. *Et₃N was added to neutralize CHCl₃ acidity.

Diastereoisomers **8bdD1**, **8bdD2**. (*M*/*P*)-1,3,7-trimethyl[1,4]benzothiazino[2,3,4-*kl*] phenothiazine-2-yl (1*S*)-ketopinate. Following procedure from **1b(OH)** (40 mg, 0.11 mmol) and (1*S*)-(+)-ketopinic acid **7d** (20 mg, 0.11 mmol), kept for 18 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum eter/CH₂Cl₂ 1/2, *Rf* 0.38)) to afford the mixture of the two diastereomeric compounds **8bdD1** and **8bdD2** (32 mg, 55% yield) as a white solid (mp 190–199 °C). ¹H NMR (400 MHz, CDCl₃) δ : 0.76 (s,3H), 1.01 (s, 3H), 1.06 (s, 3H), 1.11 (s, 3H), 1.18–1.36 (m, 5H), 1.63–1.70 (m, 1H), 1.73–1.90 (m, 4H), 1.94–1.99 (m, 2H), 2.21–2.31 (m, 18H), 2.42–2.49 (m, 2H), 6.77–6.84 (m, 7H), 6.88–7.01 (m, 7H) ppm. ¹³C NMR (100 MHz, CDCl3) δ : 19.61, 19.64, 20.51, 20.54, 20.61, 20.7, 20.78, 20.82, 20.9, 24.9, 25.3, 26.16, 26.24, 43.7, 43.8, 43.9, 44.0, 48.2, 48.5, 67.5, 67.6, 118.1, 118.6, 123.4, 123.8, 125.45, 125.55, 125.6, 125.78, 125.83, 125.9, 126.0, 126.3, 126.4, 127.0, 127.2, 127.5, 127.8, 128.3, 128.5, 129.4, 130.0, 130.7, 130.9, 133.6, 133.7, 134.8, 135.70, 135.74, 138.0, 138.1, 142.1, 142.4, 142.5, 167.2, 167.8, 210.54, 210.55 ppm (58 signals for 62 carbons). IR (ATR solid) n: 2961, 2920, 2888, 1762, 1737, 1480, 1448, 1313, 1270 cm⁻¹. Elem. Anal. for C₃₁H₂₉NO₃S₂: Calcd. C 70.56, H 5.54, N 2.65; found C 70.48, H 5.55, N 2.64.

Diastereoisomers **8beD1** and **8beD2**. (M/P)-1,3,7-trimethyl[1,4]benzothiazino[2,3,4*kl*]phenothiazine-2-yl (*S*)-2-(6-methoxy-2-naphthyl) propionate. Following the general procedure from **1b(OH)** (48 mg, 0.13 mmol) and (*S*)-(+)-2-(6-methoxy-2-naphthyl) propionic acid **7e** (28 mg, 0.13 mmol), kept for 12 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/CH₂Cl₂ 1/2, *Rf* 0.74)) to afford the mixture of the two diastereomeric compounds **8beD1** and **8beD2** (43 mg, 58% yield) as a white solid (mp 250 °C dec). ¹H NMR (400 MHz, CDCl₃) δ : 1.23 (d, 3H, *J* = 7.2 Hz), 1.39 (d, 3H, *J* = 7.2 Hz) 2.208 (s, 3H), 2.214 (s, 3H), 2.22 (s, 3H), 2.23 (s, 3H), 2.29 (s, 3H), 2.33 (s, 3H), 3.10 (q, 1H, *J* = 7.3 Hz), 3.20 (q, 1H, *J* = 7.2 Hz), 3.91 (s, 3H), 3.92 (s, 3H), 6.52 (bs, 1H), 6.65 (bs, 1H), 6.75–6.94 (m, 10H), 6.99 (bs, 1H), 7.04 (bs, 1H), 7.09–7.17 (m, 5H), 7.27–7.30 (m, 1H), 7.39 (bs, 1H), 7.53 (bs, 1H), 7.64–7.71 (m, 4H) ppm. Elem Anal. for C₃₅H₂₉NO₃S₂: Calcd. C 73.02, H 5.08, N 2.43; found C 73.03, H 5.09, N 2.43.

Diastereoisomers 8bfD1 and 8bfD2. (*M/P*)-1,3,7-trimethyl[1,4]benzothiazino[2,3,4kl]phenothiazine-2-yl mono(1R)-menthylphthalate. Following the general procedure from **1b(OH)** (40 mg, 0.11 mmol) and (-)mono(1*R*)-menthylphthalate **7f** (33 mg, 0.11 mmol), kept for 18 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/ CH_2Cl_2 1/1, Rf 0.63)) to afford the mixture of the two diastereometric compounds 8bfD1 and 8bfD2 (32 mg, 45% yield) as a white solid (mp 180–190 °C). ¹HNMR (400 MHz, CDCl₃) δ: 0.66 (d, 3H, J = 6.9 Hz), 0.78–0.90 (m, 17H), 0.98–1.11 (m, 4H), 1.40–1.53 (m, 4H), 1.65–1.71 (m, 4H), 1.90–1.99 (m, 2H), 2.05–2.26 (m, 14H), 2.53 (s, 6H), 4.85–4.93 (m, 2H), 6.70–6.85 (m, 10H), 6.95–7.00 (m, 4H), 7.07 (bs, 2H), 7.26–7.30 (m, 2H), 7.44–7.54 (m, 4H) ppm. ¹³CNMR (100 MHz, CDCl₃) δ: 16.1, 16.4, 20.5, 20.6, 20.90, 20.92, 21.02, 22.1, 22.2, 23.4, 23.5, 26.2, 26.3, 31.5, 31.6, 34.4, 34.5, 40.5, 40.7, 47.3, 47.4, 75.7, 118.45, 118.50, 123.35, 123.40, 125.7, 125.76, 125.77, 125.79, 125.9, 126.26, 126.34, 126.4, 127.0, 127.46, 127.48, 128.05, 128.12. 128.16, 128.3, 129.50, 129.53, 129.6, 129.75, 129.77, 130.15, 130.18, 130.7, 130.8, 131.4, 131.5, 133.6, 133.9, 134.1, 134.7, 134.8, 135.36, 135.40, 138.1, 138.2, 141.45, 141.52, 141.98, 142.01, 163.82, 163.84, 167.28, 167.33 ppm (56 signals for 78 different carbons). Elem. Anal. for C₃₉H₃₉NO₄S₂: Calcd. C 72.08, H 6.05, N 2.16; found C 71.99, H 6.06, N 2.15.

Diastereoisomers **8bgD1** and **8bgD2**. (*M/P*)-1,3,7-trimethyl[1,4]benzothiazino[2,3,4kl]phenothiazine-2-yl (S)-N-Boc pipecolate. Following the general procedure from 1b(OH) (40 mg, 0.11 mmol) and (S)-N-Boc pipecolic acid 7g (25 mg, 0.11 mmol), kept for 3 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/CH₂Cl₂ 1/3, D1 Rf 0.27, D2 Rf 0.20) to afford the product 8bgD1 (17 mg, 28% yield) as a white solid (mp 79–82 $^{\circ}$ C) and the product **8bgD2** (9 mg, 15% yield) as a white solid (mp 121–125 °C). **8bgD1**: ¹HNMR (400Mz, CDCl₃) δ: 1.10–1.21 (m, 1H), 1.26–1.53 (m, 14H), 2.21 (s, 3H), 2.28 (s, 3H), 2.29 (s, 3H), 2.29 (s, 3H), 2.80–2.95 (m, 1H), 3.79–3.96 (m, 1H), 4.40-4.47 (m, 1H), 6.78-6.91 (m, 6H), 7.00 (bs, 1H) ppm. ¹³C NMR (100 MHz, CDCl3) δ: 20.6, 20.7, 21.1, 21.4, 24.7, 25.0, 26.0, 28.5, 29.8, 41.2, 42.2, 54.3, 55.3, 77.2, 80.0, 80.1, 118.2, 122.6, 123.2, 125.5, 125.8, 126.3, 127.2, 127.7, 127.9, 128.1, 128.5, 129.5, 130.9, 131.3, 133.8, 134.2, 134.8, 134.9, 135.5, 138.1, 141.9, 142.4, 155.4, 155.7, 169.7, ppm. IR (ATR solid) n: 2973, 2924, 2860, 1764, 1689, 1480, 1448, 1364, 1252 cm⁻¹. $[\alpha]_D^{20}$ –157, (c 0.1, CH₂Cl₂). **8bgD2**: ¹HNMR (400Mz, CDCl₃) δ: 0.79-0.99 (m, 1H), 1.26-1.45 (m, 14H), 2.14-2.29 (m, 9H), 2.46-2.94 (m, 1H), 3.65–3.92 (m, 1H), 4.39–4.63 (m, 1H), 6.78–6.91 (m, 6H), 7.00–7.02 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl3) δ: 20.1, 20.2, 20.5, 20.6, 20.70, 20.74, 20.8, 21.0, 24.7, 24.9, 26.0, 26.1, 28.4, 28.6, 41.0, 42.0, 54.4, 55.3, 80.0, 80.3, 115.2, 118.1, 118.2, 122.7, 122.9, 125.5, 125.8, 126.4, 127.5, 128.1, 128.3, 128.9, 129.5, 131.5, 133.8, 134.9, 135.6, 138.1, 142.5, 155.7, 169.7, 169.9, ppm. $[\alpha]_D^{20}$ +49 (*c* 0.1, CH₂Cl₂).

Diastereoisomers **8bhD1** and **8bhD2**. (*M*/*P*)-1,3,7-trimethyl[1,4]benzothiazino[2,3,4*kl*]phenothiazine-2-yl (1*S*)-camphanate. Following the general procedure from **1b(OH)** (259 mg, 0.71 mmol) and (1*S*)-(–)-camphanic acid **7h** (170 mg, 0.86 mmol), kept for 22 h at room temperature. The crude was purified by flash chromatography on silica gel (petroleum ether/CH₂Cl₂ 1/3, D1 *Rf* 0.37, D2 *Rf* 0.26) to afford product **8bhD1** (143 mg, 37% yield) as a white solid (mp 70–72 °C) and product **8bhD2** (126 mg, 33% yield) as a white solid (mp 86–88 °C). **8bhD1**: ¹HNMR (400 MHz, CDCl₃) δ : 0.97 (s, 3H), 0.98 (s, 3H), 1.06 (s, 3H), 1.23–1.30 (m, 1H), 1.50–1.55 (m, 1H), 1.77–1.84 (m, 1H), 1.98–2.05 (m, 1H), 2.22 (s, 3H), 2.26 (s, 3H), 2.32 (s, 3H), 6.73 (bs, 1H), 6.79–6.81 (m, 2H), 6.85 (bs, 1H), 6.91–6.96 (m, 2H), 7.00 (bs, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ : 9.8, 16.9, 17.0, 20.6, 20.7, 20.8, 29.0, 29.7, 54.5, 54.8, 90.8, 118.2, 122.7, 125.46, 125.49, 126.0, 126.2, 126.5, 127.1, 127.9, 128.7, 129.8, 131.6, 134.5, 135.0, 135.7, 137.9, 141.3, 141.6, 164.5, 177.9. IR (ATR solid) n: 2970, 2922, 2867, 1787, 1776, 1481, 1449, 1309, 1250 cm⁻¹. $[\alpha]_D^{20}$ –129 (*c* 0.1, CH₂Cl₂). **8bhD2** ¹HNMR (400 MHz, CDCl₃) δ : 0.89 (s, 3H), 0.93 (s, 3H), 1.06 (s, 3H), 1.51–1.56 (m, 3H), 1.65–1.71 (m, 1H), 2.21 (s, 3H), 2.25 (s, 3H), 2.31 (s, 3H), 6.68 (bs, 1H), 6.77–6.79 (m, 2H), 6.84–6.89 (m, 2H), 6.96 (bs, 1H), 7.02 (bs, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ : 9.8, 16.9, 17.0, 20.6, 20.7, 20.8, 28.9 29.6, 54.3, 54.8, 90.8, 118.1, 122.7, 125.2, 125.9, 126.3, 126.6 (2C), 127.5, 128.0, 128.2, 129.9, 132.0, 133.9, 135.0, 135.7, 137.8, 141.6, 142.1, 164.5, 177.7 ppm. IR (ATR solid) n: 2967, 2922, 2865, 1776, 1482, 1449, 1309, 1250 cm⁻¹. $[\alpha]_D^{2D}$ + 126 (*c* 0.1, CH₂Cl₂).

General Procedure for the hydrolysis: To a solution of ester **8** in CH₂Cl₂/MeOH: 10/1 (roughly 0.05 M) 3 eq. of NaOH was added, and the solution was stirred for 4–6 h at room temperature. The solution was diluted with water and HCl (1M) was added until the pH was neutral, then the mixture was extracted with CH₂Cl₂ (3 × 5 mL). The organic layer was dried over Na₂SO₄, filtered, and then evaporated under reduced pressure. The crude was purified by flash chromatography on silica gel (Petroleum Ether/CH₂Cl₂ 1/2) to afford the products (*M*)-**1b(OH)** or (*P*)-**1b(OH)** as a white solid in quantitative yield. (*M*)-**1b(OH)** [α]_D²⁰ –161 (*c* 0.1, CH₂Cl₂) and (*P*)-**1b(OH)** [α]_D²⁰ +166 (*c* 0.1, CH₂Cl₂).

Experimental HPLC Analytical ($250 \times 4.6 \text{ mm}$) column packed with Chiralpak IA chiral stationary phase was purchased from Chiral Technologies Europe. The HPLC resolution of products was performed on a HPLC Waters Alliance 2695 equipped with a 200 µL loop injector and a spectrophotometer UV Waters PDA 2996. The mobile phase, delivered at a flow rate of 1.2 mL/min, was hexane/CH₂Cl₂ 70/30 v/v + 1% MeOH.

4. Conclusions

In this paper we have reported that the fine matching of the structures of the chiral auxiliaries used and, above all, the topology of their insertion on the helical skeleton, *bay-zone* vs *cape-zone*, allow for the chemical resolution of DTA[4]H) **1**. Helicene **1b(OH)** allowed for the insertion of the chiral auxiliary on the 1-position, the area of terminal ring superimposition that we indicated as the *bay-zone*. Esterification of **1b(OH)** with (1*S*)-(–)-camphanic acid **7h** provided diastereomers **8bhD1** and **8bhD2** which were successfully separated by flash chromatography and hydrolyzed providing enantiopure helicenes (*M*)-**1b(OH)** and (*P*)-**1b(OH)**, respectively.

Supplementary Materials: The following supporting information is available online: HPLC Analysis, NMR spectra, DFT calculations of compound **1b(OH)**, Optimized structures' coordinates.

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Sample Availability: Samples of the compounds are available from the authors.

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