Hymenoptera Venom Immunotherapy: How to Safely Switch to the Same Venom From a Different Manufacturer

Bilò MB¹, Martini M¹, Berra D², Scarpa A³, Losappio L⁴, Quercia O⁵, Lodi Rizzini F⁶, Bignardi D⁷, Cortellini G⁸, Zisa G⁹, Del Giudice A¹⁰, Manzotti G¹¹, Marcotulli C¹², Murzilli F¹³, Cilia M¹⁴, Pravettoni V¹⁵, Borrelli P¹⁶, Pastorello EA⁴

¹Allergy Unit, Department of Internal Medicine, University Hospital of Ancona - Allergy and Clinical Immunology School, Università Politecnica delle Marche, Ancona, Italy

²Allergology Service, Pneumology Unit, Busto Arsizio Hospital, Varese, Italy

³Allergy Unit - Dermatology, Department of Surgery, Hospital of Mirano, Mirano, Italy

⁴Department of Allergy and Immunology, Asst Grande Ospedale Metropolitano Niguarda, University of Milano, Italy

⁵Allergology High Specialty Unit General Medicine, Faenza Hospital Ravenna, Ravenna, Italy

⁶Allergy Unit, Spedali Civili Brescia, Department of Clinical and Experimental Sciences, University of Brescia, Brescia, Italy ⁷Allergy Unit, Department of Internal Medicine, Ospedale Policlinico San Martino of Genoa, Genoa, Italy

⁸Internal Medicine and Rheumatology Unit, Rimini Hospital, Rimini, Italy

⁹Allergy Unit, Department of Internal Medicine, Azienda Ospedaliero-Universitaria Maggiore della Carità, Novara, Italy ¹⁰Allergy Unit, Department of Internal Medicine, Hospital Martini of Turin, Turin, Italy

¹¹Allergy Unit, Department of Internal Medicine, Treviglio General Hospital, ASST Bergamo Ovest, Treviglio, Italy

¹²Allergy Unit, Department of Pneumology, Asst Garda, Desenzano, Italy

¹³Allergy Unit, Department of Internal Medicine, S.S. Filippo and Nicola Hospital of Avezzano, Azienda Sanitaria Locale n. 1 dell' Abruzzo, Avezzano, Italy

¹⁴Allergy Unit, Health Care Service, Scilla, Italy

¹⁵UOC General Medicine – Immunology and Allergology – IRCCS Foundation Ca'Granda Ospedale Maggiore Policlinico, Milan, Italy ¹⁶Allergy Unit, Department of Internal Medicine, U. Parini Regional Hospital, Aosta, Italy

J Investig Allergol Clin Immunol 2018; Vol. 28(3): 205-208 doi: 10.18176/jiaci.0242

Key words: Extract switching. Hymenoptera. Immunotherapy. Mastocytosis. Safety.

Palabras clave: Cambio de extracto. Himenóptero. Inmunoterapia. Mastocitosis. Seguridad.

Venom immunotherapy (VIT) is a safe and effective treatment for hymenoptera venom allergy (HVA) [1,2]. It is also recommended in patients with underlying clonal mast cell disorders [2,3] and/or stabilized respiratory/cardiovascular diseases [2].

The literature provides no clear approach to VIT-treated patients if the allergenic extract is no longer available [1,2,4]. However, for safety reasons, current clinical practice discourages abrupt switching to the same venom extract from a different manufacturer without an induction phase.

In early 2016, a national shortage of aqueous VIT preparations led clinicians to switch to another extract based on their experience.

We prospectively collected data on VIT switching from 16 Italian centers with broad experience in management of VIT. Whenever discontinuation was not recommended, patients switched to a preparation of the same venom made by another manufacturer. Before VIT, all patients signed an informed consent form and underwent a medical examination. A venous access was also placed. The baseline characteristics of the patients are shown in the Table.

Of the 531 cases recorded in 523 patients (8 patients underwent double VIT), 349 cases (Group A, 66%) switched to a new extract on a single day: their own maintenance dose, which was reached entirely with the new extract, was divided into 3 aliquots in 276 cases and 2 aliquots in 73 cases, with a 30-45 minute interval between injections. In 95 cases (Group B, 18%), the switch was performed on a single day by dividing the maintenance dose into 2 different aliquots injected at a 30-45 minute interval; however, only the old extract was used for the first injection (30% and 50% of the total maintenance dose in 31 and 64 cases, respectively), whereas at the second injection, only the new extract was injected. All the other switches (n=87) involved dose reduction by injecting different amounts of the new extract on the first day (ranging from 10% to 80% of the maintenance dose) and then gradual increases during subsequent visits until 100% of the maintenance

Table. Characteristics of Patients and VIT Switching

	Total	Group A	Group B	Group C
Patients, No.	523	342 (65.39%)	94 (17.97%)	87 (16.63%)
Male/Female	391/132	258/84	70/24	63/24
Mean age (range), y	55.5 (8-90)	55.5 (11-90)	54.6 (8-85)	48.9 (11-82)
Reaction type ^a				
Grade I	38 (7.27%)	15 (39.47%)	14 (36.84%)	9 (23.68%)
Grade II	70 (13.38%)	40 (57.14%)	13 (18.57%)	17 (24.29%)
Grade III	185 (35.37%)	128 (36.68%)	31 (16.49%)	29 (15.43%)
Grade IV	230 (43.98%)	166 (70.64%)	37 (15.74%)	32 (13.62%)
Mastocytosis and/or high bSTC	62 (11.68%)	44 (70.97%)	11 (17.74%)	7 (11.29%)
Patients with double VIT, No.	8	7	1	0
Type of VIT				
Apis mellifera	58 (10.92%)	36 (62.07%)	12 (20.69%)	10 (17.24%)
Polistes species/P dominula ^b	83 (15.63%)	54 (65.06%)	9 (10.84%)	20 (24.10%)
Vespula species	389 (73.26%)	258 (66.32%)	74 (19.02%)	57 (14.65%)
Vespa crabro	1 (0.19%)	1 (100%)	0 (0.00%)	0 (0.00%)
Mean (SD) administration interval, wk	8 (2.6)			
Mean (SD) VIT period, y	7 (5.6)			
Switches, No.	531	349 (65.73%)	95 (17.89%)	87 (16.38%)
Switch protocol	/	Full dose 1 day	Full dose 1 day	Reduced dose 1 day ^c
Extract	/	Ν	$\mathbf{N} + \mathbf{O}^{d}$	Ν
Type of new extract				
Aqueous ^e	357 (67.23%)	227 (63.59%)	64 (17.93%)	66 (18.49%)
Depot ^f	174 (32.77%)	122 (70.11%)	31 (17.82%)	21 (12.07%)
Systemic reaction	1 (0.19%)	0	0	1

Abbreviations: N, new extract; O, old extract; bSTC, serum baseline tryptase concentration; VIT, venom immunotherapy.

^aMueller grading system [6].

^bForty-four patients (46%) were treated with mixed American *Polistes* extract (*Polistes annularis, fuscatus*, and *exclamans* species), and all the others were treated with *Polistes dominula* extracts.

°100% of new extract was used during the following visits.

^dThe dose was gradually increased to the full dose during the following visits.

^ePurified, nonpurified, or capillary extracted extracts.

^fPurified aluminium hydroxide adsorbed or tyrosine adsorbed extracts.

dose was reached (Group C, 16%). The extracts used were both aqueous (purified aqueous, 10%; nonpurified aqueous, 51%; capillary extracted aqueous, 6%) and depot (purified aluminium hydroxide adsorbed, 16%; tyrosine adsorbed, 17%). All the *Polistes*-allergic patients were switched to the new *Polistes dominula* extracts. The switch protocols and the extracts used are reported in the Table. Subsequent administration of maintenance VIT doses was scheduled with a timeframe similar to that followed before the switch (8 [2.6] weeks).

The switch was well tolerated in all but 1 patient. One grade I systemic reaction (SR) [5] was reported in an 82-year-old man who had been receiving 100 μ g of *Vespula* VIT for 3.8 years (Group C). The patient had mastocytosis and ischemic heart disease treated with a β -blocker and a sartan. He experienced generalized pruritus a few hours after the injection (60 μ g) and recovered spontaneously in 4 days. His nonpurified *Vespula* aqueous extract was switched to a purified aluminium hydroxide adsorbed preparation. He tolerated subsequent injections at the same total dose after 1 week and at a dose of 100 μ g after a further week, with no SRs. Local reactions were not taken into consideration, as they are very common.

No SRs were reported in the other patients with mastocytosis and/or elevated serum baseline tryptase concentration (n=61), even though 44 (71%) followed the Group A protocol. Similarly, SRs were not observed in patients with cardiovascular comorbidity (19% of patients) or respiratory comorbidity (3%) or in patients treated with β -blockers (4%), ACE-inhibitors (5%), or sartans (9%). In addition, adverse effects were not reported in any bee venom–allergic patients (n=58) or in patients with a long interval between injections (more than 8 weeks) (35%) before switching. Moreover, patients previously treated with a mix of American *Polistes* venom tolerated the switch to *P dominula* extracts. None of the patients reported any subsequent SRs during a mean follow-up period of 18 (4.9) months.

This is the first real-life multicenter study showing the safety of switching VIT to the same venom from another manufacturer in a large number of patients. No severe SRs were reported. No significant differences were observed between 1-day protocols (Group A and B) and the dose reduction protocols (Group C), suggesting that there is no need for an induction phase using lower doses of venom. We propose 3 explanations for our encouraging safety results. First, even though 79% of patients developed a class III-IV SR [6] at baseline and 27% had a comorbid condition, the venom was switched mainly after a maintenance phase of several years without adverse effects. Second, patients were managed by clinicians with specific expertise in VIT. Third, the comorbidities and treatments analyzed do not seem to be risk factors for severe adverse effects, individually. Interestingly, the only SR reported occurred in a mastocytosis patient with ischemic heart disease treated with a ß-blocker and a sartan. Even though the SR was mild and affected only the skin, we cannot rule out the possibility that the combination of mastocytosis with cardiovascular disease could have played a role in the case we report. In clinical practice, special attention should be paid in patients with comorbidities, especially when mastocytosis occurs with other severe comorbidities.

Although bee venom is considered a risk factor for adverse effects during VIT and different allergen compositions have been demonstrated for bee venom extracts [7], our results showed that bee venom allergy does not seem to be related to a higher risk of SRs during switching. Similarly, switching from American *Polistes* species to *P dominula* venom did not cause any SRs, despite the incomplete cross-reactivity between American and European *Polistes* venom [8,9].

Notwithstanding the pitfalls normally associated with multicenter real-life studies, such as the different protocols and extracts used in this study, no differences in safety were observed between aqueous extracts and depot preparations or between the purified and nonpurified aqueous formulations.

Before switching, no patients in our study experienced severe SRs to the old extract, although we believe that, in this case, VIT should be restarted with a rush/ultrarush protocol in centers with broad experience in HVA and VIT [10], or with a conventional protocol in less experienced centers.

The unexpected and accidental shortage of some extracts was the only reason that made the VIT switch necessary in our study.

In conclusion, our results showed that switching VIT, if strictly necessary, is a safe option in patients who previously tolerated VIT, even without reducing the maintenance dose already reached by the patient and when performed in an appropriate environment by experienced staff.

Funding

The authors declare that no funding was received for the present study.

Conflicts of Interest

MBB has received speaker's honoraria and/or travel support from ALK-Abelló and Thermo Fisher Scientific. She has also received consultancy fees from ALK-Abelló for participation in an advisory board.

AS has received reimbursement for symposia from ALK Abelló and Stallergenes.

The other authors declare that they have no conflict of interests.

References

- Bonifazi F, Jutel M, Bilo BM, Birnbaum J, Muller U, EAACI Interest Group on Insect Venom Hypersensitivity. Prevention and treatment of hymenoptera venom allergy: guidelines for clinical practice. Allergy. 2005;60:1459-70.
- Sturm GJ, Varga E-M, Roberts G, Mosbech H, Bilò MB, Akdis CA, et al. EAACI Guidelines on Allergen Immunotherapy: Hymenoptera venom allergy. Allergy. 2018;73:744-64.
- Bonadonna P, Gonzalez-de-Olano D, Zanotti R, Riccio A, De Ferrari L, Lombardo C et al. Venom immunotherapy in patients with clonal mast cell disorders: efficacy, safety, and practical considerations. J Allergy Clin Immunol Pract. 2013;1:474-8.
- Wolf BL, Hamilton RG. Near-fatal anaphylaxis after Hymenoptera venom immunotherapy. J Allergy Clin Immunol. 1998;102:527-8.

- Cox L, Larenas-Linnemann D, Lockey RF, Passalacqua G. Speaking the same language: The World Allergy Organization Subcutaneous Immunotherapy Systemic Reaction Grading System. J Allergy Clin Immunol. 2010;125:569-74.
- Mueller HL. Diagnosis and treatment of insect sensitivity. J Asthma Res. 1966;3:331-3
- Blank S, Seismann H, Michel Y, McIntyre M, Cifuentes L, Braren I, et al. Api m 10, a genuine A. mellifera venom allergen, is clinically relevant but underrepresented in therapeutic extracts. Allergy. 2011;66:1322-9.
- Severino MG, Campi P, Macchia D, Manfredi M, Turillazzi S, Spadolini I, et al. European Polistes venom allergy. Allergy. 2006 Jul;61(7):860-3.
- Schiener M, Eberlein B, Moreno-Aguilar C, Pietsch G, Serrano P, McIntyre M, et al. Application of recombinant antigen 5 allergens from seven allergy-relevant Hymenoptera species in diagnostics. Allergy. 2017;72:98-108.
- Birnbaum J, Ramadour M, Magnan A, Vervloet D. Hymenoptera ultra-rush venom immunotherapy (210 min): a safety study and risk factors. Clin Exp Allergy. 2003;33:58-64.

Manuscript received December 12, 2017; accepted for publication February 16, 2018.

Maria Beatrice Bilò

Allergy Unit, Department of Internal Medicine University Hospital of Ancona Via Conca 71, 60126 Ancona, Italy E-mail: MariaBeatrice.Bilo@ospedaliriuniti.marche.it