Molecular and Clinical Characterization of Albinism in a Large Cohort of Italian Patients

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PURPOSE. The purpose of this study was to identify the molecular basis of albinism in a large cohort of Italian patients showing typical ocular landmarks of the disease and to provide a full characterization of the clinical ophthalmic manifestations.

METHODS. DNA samples from 45 patients with ocular manifestations of albinism were analyzed by direct sequencing analysis of five genes responsible for albinism: *TYR*, *P*, *TYRP1*, *SLC45A2* (*MATP*), and *OA1*. All patients studied showed a variable degree of skin and hair hypopigmentation. Eighteen patients with distinct mutations in each gene associated with OCA were evaluated by detailed ophthalmic analysis, optical coherence tomography (OCT), and fundus autofluorescence.

RESULTS. Disease-causing mutations were identified in more than 95% of analyzed patients with OCA (28/45 [62.2%] cases with two or more mutations; 15/45 [33.3%] cases with one mutation). Thirty-five different mutant alleles were identified of which 15 were novel. Mutations in *TYR* were the most frequent (73.3%), whereas mutations in *P* occurred more rarely (13.3%) than previously reported. Novel mutations were also

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identified in rare loci such as *TYRP1* and *MATP*. Mutations in the *OA1* gene were not detected. Clinical assessment revealed that patients with iris and macular pigmentation had significantly higher visual acuity than did severe hypopigmented phenotypes.

CONCLUSIONS. *TYR* gene mutations represent a relevant cause of oculocutaneous albinism in Italy, whereas mutations in *P* present a lower frequency than that found in other populations. Clinical analysis revealed that the severity of the ocular manifestations depends on the degree of retinal pigmentation. (*Invest Ophthalmol Vis Sci.* 2011;52:1281–1289) DOI:10.1167/ iovs.10-6091

lbinism is a rare inherited disorder manifested by the A complete or partial absence of pigment in the skin, hair, or eyes due to a defect in melanin biosynthesis. It can be classified as oculocutaneous albinism (OCA), when it involves the hair, skin, and eyes, or ocular albinism (OA), when the phenotype is mainly restricted to the eyes and the optic system and therefore is associated with specific ocular changes due to a reduced amount of melanin in the developing eye.^{1,2} Classic OCA is usually inherited as an autosomal recessive trait due to mutations in four genes known as TYR,³ P (OCA2),⁴ TYRP1,⁵ and SLC45A2 (MATP),⁶ which are responsible for OCA type 1 (OCA1, MIM 203100), type 2 (OCA2, MIM 203200), type 3 (OCA3, MIM 203290), and type 4 (OCA4, MIM 606574), respectively. Two general forms of OA have been distinguished: the X-linked recessive Nettleship-Falls form (OA1; MIM 300500) and autosomal recessive ocular albinism (AROA). OA1 results from mutations in the OA1 locus.⁷ AROA results from mutations in TYR, P, and possibly TYRP1, thus representing a phenotypically mild variant of OCA.8 OCA is a phenotypic component of syndromic disorders due to different gene mutations, including Hermansky-Pudlak (HPS, MIM 203300), Chediak-Higashi (CHS, MIM 214500), Griscelli (GS, MIM 214450; 607624; 609227), Tietz (TS, MIM 103500), ocular albinism with sensorineural deafness (Waardenburg syndrome type II with ocular albinism; WS2-OA, MIM 103470), Waardenburg (WS, MIM 193500), Cross (oculocerebral syndrome with hypopigmentation, CS, MIM 257800), Prader Willi (PWS, MIM 176270), and Angelman (AS, MIM 105830) syndromes.

The eye and optic system abnormalities are common to all types of albinism and are probably related to the reduction of melanin during embryonic development and early postnatal life.¹ Characteristic changes in the optic system include reduced pigmentation of the iris (iris translucency) and of the retinal pigment epithelium, foveal hypoplasia, decreased visual acuity, misrouting of the optic fibers at the chiasm, nystagmus, strabismus, and refractive errors.¹The degree of skin and hair

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hypopigmentation, when present, varies along a wide clinical spectrum of severe to mild phenotypes. The clinical spectrum of OCA varies both within and among genotypes.

OCA1 is generally considered a severe form, due to the absence of tyrosinase activity (OCA1A). Patients show a complete lack of melanin production throughout life with light blue to almost pink irises. Those with mutations determining a decreased tyrosinase activity have a milder form (OCA1B) characterized by blue to green/brown irises.³ OCA2, OCA3, and OCA4 show some pigment accumulation over time in both neural crest (skin, iris, and choroids melanocytes) and neuroectodermic (RPE cells) derived cells. OCA2, OCA3, and OCA4 patients typically have higher visual acuity than OCA1.9 OA primarily affects the eye, sparing hair, or skin, as does AROA. OA and AROA patients show the typical ocular landmarks of albinism, with relatively normal skin and hair pigmentation. Albinism can affect all ethnic backgrounds with an overall prevalence of approximately 1 in 20,000 people.¹ Prevalence of the different forms of albinism varies considerably worldwide. Several factors may be involved, including the dissimilar prevalence of different founder gene mutations in different populations.⁹

OCA2 is considered the most common type of OCA worldwide, with the highest prevalence in Africans and African-American OCA patients, which may, in part, result from the existence of a single common deletion throughout many regions of sub-Sahara Africa.^{1,10} Recent findings in a large series of non-Hispanic Caucasian patients have shown, instead, that OCA1 is the most frequent cause of OCA in Caucasian patients.¹¹ OCA3 is virtually nonexistent in Caucasians, whereas it affects frequently several African populations (~1 in 8500).^{9,10} Mutations of *MATP*, responsible for OCA4, are extremely rare in Europeans (1 in 85,000), whereas they are associated with the OCA phenotype in 24% of Japanese OCA patients.^{9,12}

At least 230 different pathologic gene mutations have been reported in TYR, 84 in P, 17 in TYRP1, and 42 in SLC45A2 (cf. Albinism Database; http://albinismdb.med.umn.edu/ developed by William Oetting and provided in the public domain by the University of Minnesota, Minneapolis, MN). Common mutations in different loci have been described in different populations. However, the causative roles of some common DNA variations are still controversial. In this respect, the p.R402Q variation in the TYR locus (common among Caucasians and African Americans), which results in a thermolabile tyrosinase polypeptide with decreased catalytic activity at 37°C (the socalled temperature-sensitive [TS] variant), has been considered for years a nonpathologic polymorphism in different populations.¹³ Instead, different studies have indicated this variant as a possible causative mutation, at least producing an additional negative effect in combination with different heterozygous mutations, even in nonallelic genes.^{14,15} In agreement with this finding, the p.R402Q variation on one allele has recently been identified in most Caucasian patients showing the AROA phenotype in combination with more severe TYR mutations on the other allele.8 Moreover, the elevated frequency of this variant in nondiagnostic OCA1 alleles in a large series of Caucasian patients has suggested that this variant may contribute to the OCA phenotype in some patients.¹¹ In addition, previous studies reported a high frequency of multiple sequence variations within a single individual.¹⁴ As a result of the high clinical and genetic heterogeneity of OCA, it is difficult to clinically distinguish among the different forms of classic OCA.

Indeed, it appears that disease-causing mutations and clinical manifestations correlate poorly. A relationship can be identified only in the case of *TYR* gene mutations. *TYR* null mutations producing inactive or incomplete polypeptides result in the most severe OCA1A clinical phenotype, due to the lack of tyrosinase

enzyme function, which in turn blocks the first step of the melanin biosynthetic pathway and, thus, the synthesis of melanin in pigmented cells. *TYR* mutations producing a partially active or hypomorphic tyrosinase enzymes result in the OCA1B milder phenotype.¹ Furthermore, genotype-phenotype correlations are not clinically valuable in the case of *P*, *TYRP1*, and *MATP* mutations. The lack of functional assays for the P, TYRP1, and MATP proteins and the limited molecular genetic and clinical data of both *TYRP1* and *MATP* hamper the possibility for a diagnostic and prognostic definition of these forms of albinism.¹⁶ Finally, no genotype-phenotype correlations have been identified in the case of mutations of OA1.¹⁷

Therefore, the side-by-side comparison of both molecular and clinical characteristics of patients affected by distinct genes and mutations may provide insights into the albinism disease spectrum and also into a more accurate diagnosis and genetic counseling.

In this study, we describe a comprehensive mutational analysis of all genes associated with nonsyndromic albinism in combination with a full clinical ophthalmic evaluation in a large cohort of 45 Italian patients with a clinical diagnosis of OCA.

MATERIALS AND METHODS

Patient Selection

The diagnostic inclusion criteria of patients were based on the presence of the following ophthalmic characteristic features: photophobia, nystagmus, reduced visual acuity, strabismus, iris translucency, fundus hypopigmentation, and foveal hypoplasia, possibly in combination with various degrees of hypopigmentation of the skin and hair⁸; VEP has not been considered necessary for the routine diagnosis of albinism.^{18,19} Syndromic forms of albinism, such as Hermansky-Pudlak, Chediak-Higashi, Griscelli, Tietz, ocular albinism with sensorineural deafness, Waardenburg, Cross, Prader Willi, or Angelman syndromes were excluded on clinical grounds, based on the absence of additional clinical findings such as deafness; immune deficiency; hematologic abnormalities or bleeding diathesis; heart, lung, genitourinary, gastrointestinal or central nervous system involvement; and the presence of obesity and dysmorphic features, as reported at the time of the clinical diagnosis from the ophthalmologist at the referring center.

All patients analyzed showed variable skin and hair involvement with mild or severe degrees of hypopigmentation, representing the clinical spectrum of OCA phenotypes.

Mutation Analyses

We analyzed genomic DNA of 45 Italian patients from 40 independent families, referring to ophthalmic institutes in different Italian regions and having a clinical diagnosis of nonsyndromic albinism. The research adhered to the tenets of the Declaration of Helsinki. Informed consent was obtained from each patient, or from the patient's legal representatives in the case of children, after explanation of the nature and possible consequences of the study. To establish a molecular diagnosis, we performed direct DNA sequence analysis of five genes known to be associated with albinism: *TYR*, *P*, *TYRP1*, *MATP*, and *OA1*. Patients were initially screened for *TYRP1*, *MATP*, and *OA1* gene mutations.

A detailed pedigree analysis of the extended families was performed to determine the mode of inheritance, and in 22 independent families, the segregation of mutations was confirmed by gene sequence analysis in all family members.

Genomic DNA was extracted from patients' peripheral whole blood lymphocytes and, when available, from their parents, by using standard techniques (Qiagen Italy, Milan, Italy). The complete coding sequence and the exon-intron boundaries of the OA1, TYR, P, TYRP1, and MATP genes were amplified by standard PCR (Taq Gold DNA polymerase; Roche, Basel, Switzerland). Tyrosinase exon 1; OA1 exons 1, 3, and 8; and *MATP* exon 3, were amplified as a pair of overlapping fragments. The exon 1 of *P* and *TYRP1* genes, which are noncoding, was not analyzed. Human chromosome 11 contains a pseudogene, known as the tyrosinase-like gene (*TYRL*, 11p11.2; MIM 191270), that shares a 98.55% sequence identity with the 3'-region of *TYR* (~68 kb), including exons 4 and 5. Thus, the identification of nucleotide variants in *TYR* by PCR and DNA sequencing is a challenging task and could generate false data due to the co-amplification from both loci. To allow the direct and unequivocal identification of mutations, we used primers for a specific amplification of the *TYR* locus, as described in Chaki et al.²⁰

The PCR was performed in 35 cycles with 50 ng of genomic DNA at 94°C for 1 minute, at the respective primer annealing temperature for 1 minute, and at 72°C for 1 minute. The primers and reaction conditions of the PCR amplification are available on request. Amplicons were screened for mutations by direct sequencing (Prism Big Dye terminator cycle sequencing V2.0 kit; Applied Biosystems, Inc. [ABI], Foster City, CA), and the reactions were analyzed on a genetic analyzer (Prism 3100; ABI). The sequenced exon and intron- exon boundaries were compared against consensus sequences obtained from the National Centre for Biotechnology Information Database (http://www.ncbi.nlm.nih.gov/), using standard software for DNA sequencing analysis (Autoassembler, ver. 2.1; ABI).

Mutation nomenclature conformed to standard convention.²¹ The identified nucleotide variations were researched in either the Albinism Database (http://albinismdb.med.umn.edu/) or the Human Gene Mutation Database (http://www.hgmd.cf.ac.uk/ac/index.php/ provided in the public domain by the Institute of Medical Genetics, Cardiff, Wales, UK) to check whether they are described as causative mutations or polymorphisms.

Nine of 13 newly identified sequence variants leading to amino acid missense substitution were tested in other affected and unaffected family members, to verify the segregation with the albino phenotype. Any novel missense variation was considered a possible causative mutation if it was absent in at least 100 control chromosomes, analyzed by denaturing high performance liquid chromatography (dHPLC). All control products displaying a dHPLC pattern similar to the patient's were sequenced. The newly identified frameshift mutations were not tested in control samples, and they were considered to be causative mutations.

Phenotype Analysis

An additional full ophthalmic evaluation, including visual acuity measurements, evaluation of ocular motility and iris translucency, biomicroscopic examination, fundus examination, fundus autofluorescence (FAF), and retinography, was performed in 18 patients harboring mutations of different genes causing albinism. A 4- and 3-point scale according to Summers et al.²² was used to classify iris translucency and macular transparency, respectively.

Autofluorescence was recorded with a standard confocal scanning laser ophthalmoscope (Heidelberg Retina Angiograph II; Heidelberg Engineering, Heidelberg, Germany). To amplify the autofluorescence signal, we aligned the best five images obtained by using the integrated system software and calculated a mean image.

Cross-sectional retinal reflectivity profiles were obtained with optical coherence tomography (OCT3; Carl Zeiss Meditec, Inc., Oberkochen, Germany). Subjects underwent OCT imaging incorporating 512 A-scans over a 3- μ m transverse scanning length for an optimal sampling rate of 400 A-scans per second centered on the fovea. According to the manufacturer, the longitudinal resolution of the present model is 8 to 10 μ m, and the transverse resolution is approximately 20 μ m. The precise location and orientation of each scan were determined with the OCT simultaneous-view video images.

Because nearly all patients were affected by nystagmus, which complicates the recording, OCT scans, and autofluorescence were obtained in only 15 patients. Correlation analysis was performed with Spearman's test.

RESULTS

Mutational Analysis

We analyzed 45 Italian individuals showing ophthalmic characteristic features of albinism including photophobia, nystagmus, reduced visual acuity, strabismus, iris translucency, fundus hypopigmentation, and foveal hypoplasia possibly in combination with various degrees of hypopigmentation of the skin and hair (23 males, 22 females, mean age, 17.6 ± 15 years) from 40 independent families. We found causative DNA variation in 43 (95.6%) of 45 patients analyzed. We identified 2 or more causative mutations in 62.2% of all patients, of which 8 (17.7%) were homozygotes and 20 (44.44%) were compound heterozygotes. In 15 (33.3%) cases the second mutation was not detected. We did not find mutations in the genes studied in two (4.4%) patients (Fig. 1). In three patients, more than two causative mutations were present, either in allelic (two cases) or nonallelic (one case) genes (Table 1). Overall, we detected 35 different, and possibly causative, mutations in TYR, P, TYRP1, and MATP, 15 (42.8%) of which were novel (Table 1). Furthermore, we identified 20 DNA variations in TYR, P, TYRP1, MATP, and OA1 that have been considered as polymorphisms (Supplementary Table S1, http://www.iovs.org/ lookup/suppl/doi:10.1167/iovs.10-6091/-/DCSupplemental).

TYR was the most frequently mutated gene in our cohort of patients (33/45; 73.3%), followed by *P* (6/45; 13.3%), *MATP* (3/45; 6.7%), and *TYRP1* (1/45; 2.3%). No causative mutations were detected in the *OA1* gene. A total of 22 missense, 6 nonsense, and 6 frameshift mutations and 1 splicing mutation were identified. Four novel mutations in *TYR* and four in *P* were detected. We also identified mutations in rare loci, including six novel mutations in *MATP* and one in *TYRP1* (Supplementary Table S2, http://www.iovs.org/lookup/suppl/doi: 10.1167/iovs.10-6091/-/DCSupplemental).

OCA1 (TYR)

TYR mutational analysis showed 22 distinct mutations in 33 patients, including missense, frameshift, nonsense, and splicing variations (Table 1). Thirteen of 23 missense mutations identified were located in the exons 1 and 3.

The most frequent mutated allele identified was the c.G1205A (p.R402Q variation), which was found in 17 patients and 21 alleles. Because of the controversial causative role of this mutation, we also analyzed the other OCA and OA genes in four homozygous patients (P23-P26) and did not identify any additional causative mutation in these distinct loci. Ten patients carrying the p.R402Q variation were compound



FIGURE 1. Distribution of mutated genes involved in albinism based on the results of mutational analysis in our cohort of patients.

TABLE 1. 1	Positive Cases in th	he Mutat	ional Scr	eening									
					Mutation 1				Mutation 2			Other Mutations	
Gene/ Patient	Independent Families	Age	Sex	Nucleotide Change	AA Change	Parent	Ref	Nucleotide Change	AA Change	Parent	Ref		Ref
TYR													
P1	1		F	c.98A>C	p.K33T	Μ		c.286_287insA	p.F95Fr	F	23		
P2	2	21	F	c.255T>A	p.Y85X	Μ	24	c.61C>T	p.P21S		16		
P3	ŝ	14	Μ	c.255T>A	p.Y85X	Μ	24	c.346C>T	p.R116X	Ц	25	<i>TYR</i> : c.1467_68insT	26
P4	4	12	Μ	c.255T>A	p.Y85X	M	24	NF	4				
P5	v	32	ц	c.255T>A	p.Y85X	Μ	24	NF					
P6	6	47	ц	c.619G>A	p.A206T		27	c.823G>T	p.V275F		11		
$\mathbf{P7}$	7		W	c.606T>G	p.H202Q			NF	-				
P8	80	63	Ľ.	c.823G>T	p.V275F		11	c.823G>T	p.V275F		11		
P9	6	37	ц	c.883G>A	p.G295R		28	NF	4				
P10	10		Μ	c.1037-7T>A	4		80	c.1037-7T>A			8		
P11	11	8	Μ	c.1037-7T>A		Μ	œ	c.1177delG	p.V393Fr	F			
P12	12	13	Μ	c.1037-7T>A		Μ	œ	NF					
P13		13	Μ	c.1037-7T>A		Μ	œ	NF					
P14	13		ц	c.1118C>A	p.T373K	Н	œ	c.1351A>G	p.Y451C	Μ			
P15	14	16	ц	c.1204C>T	p.R402X	Μ	80	c.137G>A	p.C46Y	Н			
P16		24	W	c.1204C>T	p.R402X	M	œ	c.137G>A	p.C46Y	F			
P17	15	6	Μ	c.1205G>A	p.R402Q	Μ	œ	c.74_75 insT	p.V25Fr				
P18	16	13	W	c.1205G>A	p.R4020	M	œ	c.137G>A	p.C46Y				
P19	17	8	W	c.1205G>A	p.R4020	M	œ	c.573delA	p.Y191Fr	F			
					-							TYR:	
P20	18	к	F	c.1205G>A	p.R402Q		œ	c.832C>T	p.R278X			c.346C>T (p.R116X)	25
P21	19		н	c.1205G>A	p.R402Q	ц	œ	c.1177delG	p.V393Fr	M			
P22	20	15	н	c.1205G>A	p.R402Q	ц	œ	c.1177delG	p.V393Fr	M			
P23	21	40	W	c.1205G>A	p.R402Q		x o	c.1205G>A	p.R402Q		×		
P24	22	0	ц	c.1205G>A	p.R402Q		x a	c.1205G>A	p.R402Q		8		
P25	23	24	ц	c.1205G>A	p.R402Q		0 0	c.1205G>A	p.R402Q		×		
P26	24	12	Z ;	c.1205G>A	p.R402Q	1	c 9	c.1205G>A	p.R402Q	;	×		
P27	57	10	E i	c.1205G>A	p.K402Q	ц,	, 0	c.1217C>1	p.P406L	E i	×		
P28	26	17	щ;	c.1205G>A	p.R402Q	Μ;	c a	c.1217C>T	p.P406L	ц,	x S		
P29	/7	• ۷	Ξı	c.12056>A	p.K402Q	Μ) oc	C.140/_08 ms1	p.A490Hr	ц	07		
P.50	70 70		ı X	c.1205C>A	p.K402Q		00	NF					
1.71 D3.7	64 6 0	11	Ц	C.1205G>A	D204A.c		œ	NF					
174 D22	00 15	v	4 1	C.1203G/A	7204N.q		80	nr 610\T	5 D316	Ν	91		
CC1 d	10	0	4	C.12000/A	D20FN.q			C.010/1	017J.J.J	M	01		
P34	32	33	W	c.1025A>G	p.G432C			NF					
P35	33	L-	Μ	c.1327G>A	p.V4431	ц	29	c.2360C>T	p.A787V	Μ	25		
P36	34	4	н	c.2060C>T	p.A687V			NF	4				
P37	35	41	ц	c.2216T>C	p.I739T	н		c.2216T>C	p.1739T	Μ			
P38		24	M	c.2216T>C	p.1739T	ц		c.2216T>C	p.1739T	M			
P39	36	13	Щ	c.2329T>C	p.C777R	Ч		NF				(cont	(unes)
													(~~~~)

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					Mutation 1				Mutation 2			Other Mutations	
ne/ ient	Independent Families	Age	Sex	Nucleotide Change	AA Change	Parent	Ref	Nucleotide Change	AA Change	Parent	Ref		Ref
RPI													
940 17P	37	16	н	c.869G>A	p.C290Y	н		c.869G>A	p.C290Y	W			
				c.161 171insG								TYR:	
o41	38		н	GTGGAGGCAG				c.1532C>A	p.A511E			c.1205G>A (p.R402Q)	x
942	39		М	c.G126T	p.M421			c.T1280C	p.L427P			,	
943	40	4	Μ	c.375T>A	p.V126D	Ц		c.1108G>C	p.G370R	Μ			
Novel	mutations are sha	ded. NF,	, not fou	ind; M, mother; F, fath	er.								

TABLE 1 (continued). Positive Cases in the Mutational Screening

heterozygous in combination with either described or novel causative mutant alleles. The p.R402Q variant was also found in a patient bearing two causative mutations in *MATP*. In three further patients (P30-P32) carrying the p.R402Q variation, a second causative mutation was not detected.

We found three novel missense mutations: p.Y451C (c.A1351G), p.C46Y (c.137C>A), and p.H202Q (c.606T>G), all affecting amino acid residues highly conserved across evolution (from *Xenopus laevis* to *Homo sapiens*). A fourth novel mutation found in three compound heterozygous patients leads to a frameshift (c.1177delG). In two patients (P3, P19) we found three causative mutations in *TYR*.

OCA2 (P), OCA3 (TYRP1), and OCA4 (MATP)

Mutations of the *P* gene were found in six patients. All identified mutations (including four novel ones; Table 1) were missense variations. Two sibling patients (P37, P38) were homozygous for the novel mutation c.2216T>C (p.I739T) and one patient (P35) was compound heterozygous for two known mutations (c.1327G>A, p.V443I and c.2360C>T, p.A787V). In the remaining three individuals, a second mutation was not found (P34, P36, and P39).

The coding sequence of the *TYRP1* gene was mutated in only one patient (P40), showing a novel homozygous missense mutation in exon 4 that results in a cysteine-to-tyrosine amino acid change (p.C290Y, c.G869A). The absence of this variation in 220 healthy control chromosomes with the absence of any apparent mutations in the other OCA and OA genes, and also the conservation of the amino acid residue across evolution, suggests that this variation may cause the disease phenotype.

Finally, six different novel mutations were detected in *MATP* gene in three patients. Patient 42 and 43 were compound heterozygous for two novel missense *MATP* mutations. Patient 41 showed two novel mutations affecting the first and the seventh exons of *MATP* and an additional nonallelic *TYR* gene variation (p.R402Q).

Identification of Polymorphisms

Direct sequencing analysis revealed 20 different polymorphisms in exons or into nearby intronic sequences of the screened genes (Supplementary Table S1, http://www.iovs. org/lookup/suppl/doi:10.1167/iovs.10-6091/-/DCSupplemental). We found overall eight novel polymorphisms in *P*, *TYRP1*, and *MATP* and none in the *TYR* gene.

Thirty percent of the exonic polymorphisms resulted in amino acid changes, whereas the remaining ones were silent.

The *P* gene had a significantly high frequency of missense variations with no apparent pathogenic significance (Supplementary Table S1, http://www.iovs.org/lookup/suppl/doi:10. 1167/iovs.10-6091/-/DCSupplemental). Among them, we identified two novel nucleotide variations: c.1355T>A (p.452V/E) and c.1113C>T (no change D371).

We found two new intronic polymorphisms of the *TYRP1* gene with high frequency. IVS7–19InsA was identified in 36.9% and IVS6+20C/T in 32.6% of the patients.

Patient 43 showed a c.1122G>C and a c.G814A substitution affecting exon 5 and 3, respectively, of the *MATP* gene that lead to the nonpathogenic protein variations p.374F/L and p.E272K.

Two polymorphic intronic variations were found at the OA1 gene: IVS6+10G/C and IVS8+12T/C, both with a frequency of 4.3%.

Phenotype Analysis

A detailed ophthalmic evaluation, including visual acuity measurements, evaluation of ocular motility and iris translucency, fundus examination, fundus autofluorescence (FAF), and opti-

TABLE 2.	Ophthalmic	Features of	of 18	Patients	Analyzed	Phenotypically
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Pt.	Gene	Age	Iris Pigmentation	Iris Translucens	Macular Transparency	Foveal Pit	Autofluorescence Macular Pigment	CVA RE	CVA LE	Nystagmus	Strabismus
3	TYR	14	LB	3	1	No	Absent	20/200	20/200	Yes	No
4	TYR	12	LB	3	1	No	Absent	20/100	20/100	Yes	_
9	TYR	37	LB	2	2	No	ND	20/200	20/200	Yes	Yes
11	TYR	8	LB	2	2	_	Absent	20/200	20/200	Yes	Yes
15	TYR	16	LB	3	2	No	Absent	20/600	20/600	Yes	No
16	TYR	24	LB	3	2	No	Absent	20/600	20/600	Yes	No
17	TYR	9	LB	1	3	Yes	ND	20/100	20/50	No	Yes
18	TYR	13	GB	1	3	No	Present	20/25	20/25	No	No
22	TYR	15	LB	3	2	No	Absent	20/100	20/100	Yes	Yes
23	TYR	40	LB	3	1	No	Absent	20/400	20/200	Yes	Yes
26	TYR	12	LB	3	1	ND	ND	20/400	20/400	Yes	Yes
33	TYR	5	LB	2	3	No	Present	20/100	20/100	No	Yes
35	Р	8	GB	2	2	Yes	Absent	20/200	20/200	Yes	Yes
37	Р	41	GG	1	3	No	Present	20/60	20/60	Yes	Yes
38	Р	34	GB	1	3	Yes	Present	20/60	20/40	Yes	Yes
40	TYRP1	18	LB	1	ND	Yes	Present	20/100	20/100	Yes	Yes
41	MATP	51	GB	3	1	No	Absent	20/400	20/200	Yes	Yes
43	MATP	4	ND	ND	ND	Yes	Present	20/100	20/200	Yes	Yes

cal coherence tomography (OCT), was performed in 18 OCA patients (mean age, 19 ± 14 years) from 16 independent families with mutations in each of the four OCA genes studied (Table 2). Clinical analysis revealed that nystagmus and strabismus were present in most patients independently from the mutated gene. Autofluorescence showed absence of macular pigment in 9 of 15 patients examined, even in those showing an intermediate degree (grade 2) of fundus pigmentation (macular transparency evaluation).

The patients carrying *P*, *TYRP1*, and *MATP* mutations showed a milder ocular phenotype than most patients carrying *TYR* mutations. Those carrying *TYR* mutations showed high prevalence of light blue (LB) iris pigmentation (11/12 cases) and high degree of iris translucency (7/12 patients with grade 3 iris translucency). Fundus examination and autofluorescence analysis revealed a high or intermediate degree of fundus hypopigmentation (9/12 patients with grade 1 or 2 macular transparency), whereas OCT analysis demonstrated the absence of the foveal pit in 9 of 10 cases. Most patients (7/12) displayed low visual acuity, ranging between 20/600 and 20/200.

Three patients carrying P mutations showed gray (GG) or gray-blue (GB) iris pigmentation with a variable amount of iris pigment and punctate transillumination, a high degree of fundus pigmentation (grade 3 in two of three patients), and the presence of the foveal pit in two of three patients. In two patients, visual acuity was preserved (from 20/60 to 20/40) despite the presence of nystagmus and strabismus.

The only patient carrying mutations of *TYRP1* showed a mild ocular phenotype, LB iris, minimal punctuate transillumination, and presence of the foveal pit based on OCT evaluation and a visual acuity of 20/100 in both eyes.

Two patients bearing *MATP* mutations presented heterogeneous clinical findings, with a more severe ocular involvement and a worse visual acuity in the patient carrying the additional p.R402Q variation (c.G1205A) of the *TYR* gene.

Correlation analysis revealed that, independent of the gene mutation, reduced iris translucency and macular transparency (as markers of pigmentation) were significantly associated with (1) better visual acuity (P = 0.0007); (2) the presence of macular pigment assessed by autofluorescence analysis (P < 0.0001); (3) the presence of the foveal pit by OCT evaluation (P = 0.04); and (4) the absence of nystagmus (P = 0.05). Importantly, preserved visual acuity correlated with statistical significance (P = 0.0007) with presence of macular pigment,

as assessed by autofluorescence analysis. No correlation was found between better visual acuity and the presence of the foveal pit (P = 0.3) or the absence of nystagmus (P = 0.06).

Examples of clinical findings including fundus photography, FAF, and OCT examinations in four patients with mutations in four different OCA genes are reported in Figure 2. As shown in the figure, a mild phenotype was found in two patients carrying *P* and *TYRP1* mutations, which showed presence of the foveal pit, no visible choroidal macula vessels, and the presence of macular pigment evaluated by autofluorescence analysis. Two patients carrying *TYR* and *MATP* mutations, respectively, showed absence of the foveal pit, visible choroidal macula vessels, and absence of macular pigment by autofluorescence analysis.

DISCUSSION

In this study, we provided the first comprehensive mutational analysis of all genes associated with nonsyndromic albinism, performed in combination with a detailed clinical ophthalmic evaluation of a large series of Italian patients with albinism.

In agreement with previous results in Caucasian patients¹¹ we found that the most frequent mutations are associated with *TYR* in the Italian population rather than with *P*. We found 73.3% OCA1, 13.3% OCA2, 2.3% OCA3, and 6.7% OCA4 in 45 Italian patients. In 4.4% of the cases, we were not able to identify any pathologic mutations. In 62.2% of patients we found homozygous or compound heterozygous mutations, whereas in 33.3% of cases we identified mutations in only one allele.

We found a wide spectrum of causative mutations in *TYR*, *P*, *TYRP1*, and *MATP* and additional DNA polymorphisms in five genes associated with different forms of albinism. Moreover, we identified 15 novel causative mutations in four different loci, expanding the database of OCA-causing mutations.

A missense mutation cluster was identified in putative functional domains of the TYR enzyme.^{27,30} Consistently, our data showed that many of the missense mutations identified are located in/or alongside the copper-binding sites corresponding to exons 1 and 3. Mutations in these regions are considered to act either by affecting copper binding or by disrupting the substratebinding site.³¹ Our data confirm that different mutations within the tyrosinase-coding region underlie OCA1 disorders of varying severity. In one unusual subset of OCA1 patients, OCA1-TS, the



FIGURE 2. Fundus photography, FAF, and OCT analysis in four patients with mutations in different OCA genes: P3 (A, E, I), carrying TYR mutation; P36 (B, F, J) bearing a P mutation; P40 (C, G, K) with a TYRP1 mutation; and P41 (D, H, L) showing an MATP mutation (see Table 1). (A-D) Fundus photographs: choroidal vessels are visible in the macula in P3 (A) and P41 (D), but they are not visible in P36 (B) and P40 (C); (E-H) FAF: please note the absence of macular pigment in P3 (E) and P41 (H) and the presence of macular pigment in P36 (F) and P40 (G). (I-L) OCT of the posterior pole crossing the fovea, showing absence of the foveal pit in P3 (I) and P41 (L) and presence of the foveal pit in patients P36 (J) and P40 (K).

mutation generates a temperature-sensitive tyrosinase protein.32 Consequently, melanin synthesis occurs only in the cooler areas of the body, such as the arms and legs. Several TS variants of tyrosinase have been sequenced. One of these (p.R402Q) represented the most common TYR mutation identified in our cohort of Italian families (37% of patients), who showed an evident OCA phenotype with various degrees of severity in ocular phenotype. This finding is in agreement with recent studies.^{8,14,15} The p.R402Q variation (c.G1205A) was identified in our cohort of patients, either in compound heterozygotes (in combination with known or novel pathogenic mutant alleles) or in patients who did not show a second causative mutation. Interestingly, we also found this variation to be the only identified mutation with a possible causative role in four homozygous patients with an oculocutaneous phenotype, two of whom showed severe visual loss. These data suggest a causative role of the p.R402Q variation of TYR, which has been considered for years a nonpathologic polymorphism.¹³ Furthermore, we identified this variation as an additional nonallelic mutation in 1 patient (P41) bearing two novel mutations in MATP. This patient showed a more severe ocular involvement in comparison with patients carrying only MATP mutations, thus suggesting an additional negative effect of the p.R402Q mutation in combination with allelic and nonallelic mutations.

OCA2 is the most common type of albinism, especially in black African OCA patients, occurring in approximately 50% of OCA patients worldwide.³³ We found mutations of *P* only in 14% of Italian patient, carrying either known or novel mutations. Unlike from the *TYR* gene analysis, *P* gene mutations did

not appear to cluster in specific regions but they were found between or in the proximity of transmembrane domain borders in the central region of the protein. Presently, few mutations causing the OCA3 phenotype worldwide have been identified in the TYRP1 gene.^{5,34-36} TYRP1 mutations causing OCA have been considered virtually absent in Caucasians,11 although a single German patient has been described.35 We identified one patient carrying a novel homozygous missense mutation of TYRP1 (p.C290Y), who showed a mild ocular phenotype. This novel mutation affects an amino acid residue that is highly conserved across evolution. Despite the low frequency of mutations in MATP (OCA4) worldwide¹² with the exception of Japan, we found six novel MATP mutations in three Italian patients with a variable OCA phenotype. As expected, based on the selection of patients who showed at least some degree of skin and hair hypopigmentation, we did not find any mutation of the OA1 gene.

We were unable to identify mutations in 4.4% of patients screened, and we found mutations in only one allele in 33.3% (15/45) of cases. In these patients, mutations may be present in intronic or regulatory regions, which were not analyzed in our molecular analysis. Also, large genomic rearrangements cannot be identified with the strategy used in the present study. Finally, in patients with unidentified mutations, the disease may be due to mutations in OCA genes that are still to be identified as OCA-causing genes.

In a previous study,¹⁴ we identified more than two causative mutations in three patients (P3, P20, P41), either in allelic or nonallelic genes. Digenic mutation types (mutations in nonallelic genes) have been described in some cases of ocular albinism and Waardenburg syndrome.^{15,37} Our findings support the hypothesis that heterozygous mutations in nonallelic genes may result in a more severe phenotype.

Our comprehensive analysis of clinical and molecular data confirms that a phenotypic variability is present among patients and that it is generally independent from the mutated gene and the specific mutation. The lack of a sufficient number of patients with each gene mutation did not allow us to draw genotype-phenotype correlations within and among genotypes.

Most of the TYR patients showed a more severe ocular phenotype than did patients bearing mutations of P, TYRP1, or MATP, as previously reported in a genotype-phenotype correlation study.²⁸ However, independent of the gene involved, the severity of the clinical phenotype inversely correlated with the degree of retinal pigmentation, confirming previous findings.38,39 In this respect, we found that visual acuity was more preserved in patients with low iris translucency and macular transparency assessed by autofluorescence analysis. In addition, autofluorescence analysis showed absence of macular pigment (and a poor visual acuity) also in those patients in whom fundus pigmentation assessments revealed an intermediate degree of macular transparency. These data support the use of autofluorescence analysis as a more sensitive analysis to predict the visual phenotype severity compared with fundus macular transparency in albino patients. We also found that low visual acuity was not associated with the presence of the foveal pit, in accordance with the recent report of Marmor et al.⁴⁰

In conclusion, our clinical evaluation suggests that the degree of ocular pigmentation correlates with preservation of visual function. Our results confirm the wide clinical and genetic heterogeneity of OCA. They also suggest that *TYR* mutations cause a more severe ocular phenotype in most patients in comparison with *P*, *TYRP1*, or *MATP* mutations. Data obtained from the detailed ophthalmic evaluation also suggest that the severity of ocular phenotype depends on the degree of retinal pigmentation and support the use of autofluorescence analysis of macular pigment as a sensitive prognostic tool to predict the visual function in albino patients.

Finally, as the different OCA forms are indistinguishable on clinical grounds, and also considering both the high frequency of multiple DNA changes in the same patient and the lack of clear cut correlations between the clinical phenotype and molecular genotypes, we concluded that a comprehensive mutational analysis of at least four genes, such as *TYR*, *P*, *MATP*, and *TYRP1* is necessary for the differential diagnosis and genetic counseling of patients who have oculocutaneous albinism. We believe that the wide range of patients analyzed in the present study provides valuable information for a better prognostic evaluation of patients who could benefit from the future experimental development of therapies.^{41,42}

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