

Mapping the human genetic architecture of COVID-19

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COVID-19 Host Genetics Initiative*

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The genetic make-up of an individual contributes to the susceptibility and response to viral infection. Although environmental, clinical and social factors have a role in the chance of exposure to SARS-CoV-2 and the severity of COVID-19^{1,2}, host genetics may also be important. Identifying host-specific genetic factors may reveal biological mechanisms of therapeutic relevance and clarify causal relationships of modifiable environmental risk factors for SARS-CoV-2 infection and outcomes. We formed a global network of researchers to investigate the role of human genetics in SARS-CoV-2 infection and COVID-19 severity. Here we describe the results of three genome-wide association meta-analyses that consist of up to 49,562 patients with COVID-19 from 46 studies across 19 countries. We report 13 genome-wide significant loci that are associated with SARS-CoV-2 infection or severe manifestations of COVID-19. Several of these loci correspond to previously documented associations to lung or autoimmune and inflammatory diseases^{3–7}. They also represent potentially actionable mechanisms in response to infection. Mendelian randomization analyses support a causal role for smoking and body-mass index for severe COVID-19 although not for type II diabetes. The identification of novel host genetic factors associated with COVID-19 was made possible by the community of human genetics researchers coming together to prioritize the sharing of data, results, resources and analytical frameworks. This working model of international collaboration underscores what is possible for future genetic discoveries in emerging pandemics, or indeed for any complex human disease.

The COVID-19 pandemic, caused by infection with SARS-CoV-2, has resulted in an enormous health and economic burden worldwide. One of the most remarkable features of SARS-CoV-2 infection is the variation in consequences, which range from asymptomatic to life-threatening, viral pneumonia and acute respiratory distress syndrome⁸. Although established host factors correlate with disease severity (for example, increasing age, being a man and higher body-mass index¹), these risk factors alone do not explain all of the variability in disease severity observed across individuals.

Genetic factors contributing to COVID-19 susceptibility and severity may provide new biological insights into disease pathogenesis and identify mechanistic targets for therapeutic development or drug repurposing, as treating the disease remains a highly important goal despite the recent development of vaccines. Further supporting this line of inquiry, rare loss-of-function variants in genes involved in the type I interferon response may be involved in severe forms of COVID-19^{9–11}. At the same time, several genome-wide association studies that investigate the contribution of common genetic variation^{12–15} to COVID-19 have provided robust support for the involvement of several genomic loci associated with COVID-19 severity and susceptibility, with the strongest and most robust finding for severity being at the 3p21.31 locus^{12–16}. However, much remains unknown about the genetic basis of susceptibility to SARS-CoV-2 and severity of COVID-19.

The COVID-19 Host Genetics Initiative (COVID-19 HGI) (<https://www.covid19hg.org/>)¹⁷ is an international, open-science collaboration to share scientific methods and resources with research groups across the world with the goal to robustly map the host genetic determinants of SARS-CoV-2 infection and the severity of the resulting COVID-19 disease. Here, we report the latest results of meta-analyses of 46 studies from 19 countries (Fig. 1) for COVID-19 host genetic effects.

Meta-analyses of COVID-19

Overall, the COVID-19 HGI combined genetic data from 49,562 cases and 2 million controls across 46 distinct studies (Fig. 1). The data included studies from populations of different genetic ancestries, including European, admixed American, African, Middle Eastern, South Asian and East Asian individuals (Supplementary Table 1). An overview of the study design is provided in Extended Data Fig. 1. We performed case-control meta-analyses in three main categories of COVID-19 disease according to predefined and partially overlapping phenotypic criteria. These included (1) critically ill cases of COVID-19 defined as those individuals who required respiratory support in hospital or who died due to the disease; (2) cases of moderate or severe COVID-19 defined as those participants who were hospitalized due to symptoms associated with the infection; and (3) all cases with reported SARS-CoV-2 infection

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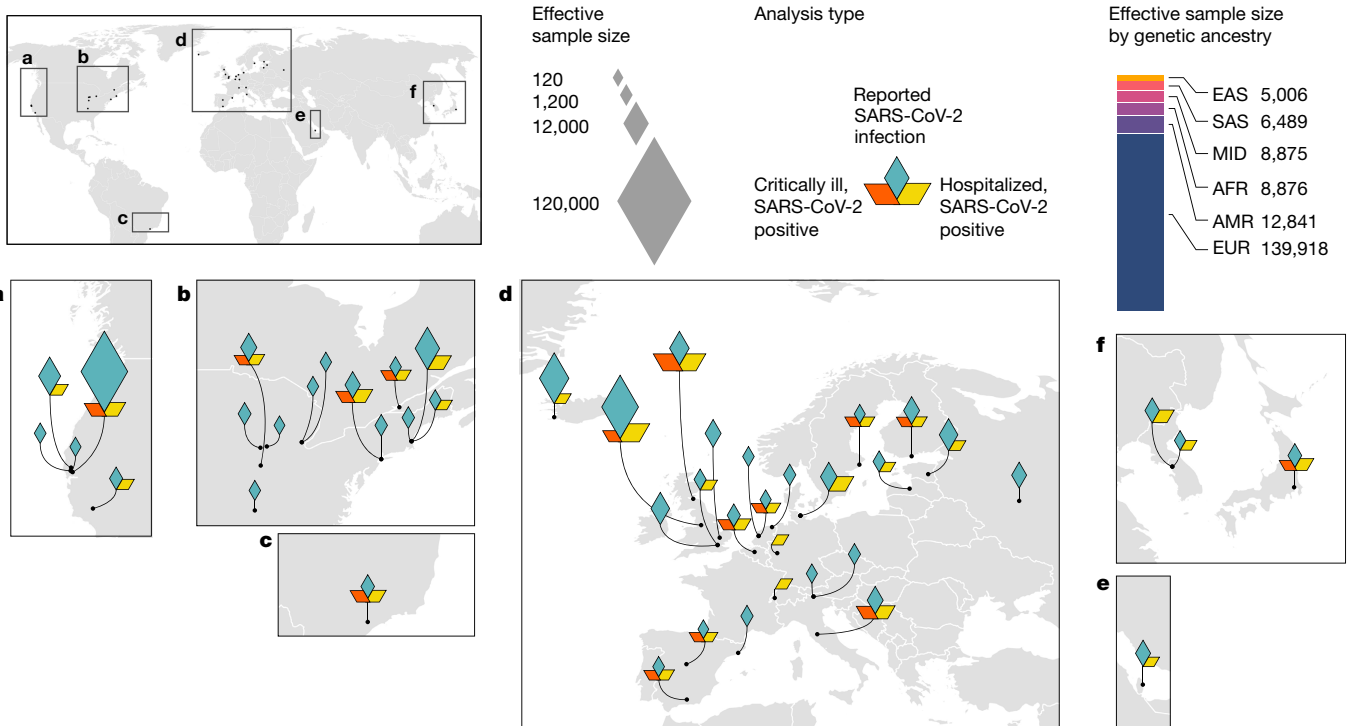


Fig. 1 | Geographical overview of the contributing studies to the COVID-19 HGI and composition by major ancestry groups. Populations are defined as African (AFR), admixed American (AMR), East Asian (EAS), European (EUR), Middle Eastern (MID) and South Asian (SAS).

regardless of symptoms (Methods). Controls for all three analyses were selected as genetically ancestry-matched samples without known SARS-CoV-2 infection, if that information was available (Methods). The average age of the participants with COVID-19 across studies was 55 years (Supplementary Table 1). We report quantile–quantile plots in Supplementary Fig. 1 and ancestry principal component plots for contributing studies in Extended Data Fig. 2.

Across our three analyses, we reported a total of 13 independent genome-wide significant loci associated with COVID-19 (the threshold of $P < 1.67 \times 10^{-8}$ is adjusted for multiple trait testing) (Supplementary Table 2), most of which were shared between two or more COVID-19 phenotypes. Two of these loci are in very close proximity within the 3p21.31 region, which was previously reported as a single locus associated with COVID-19 severity^{12–16} (Extended Data Fig. 3). Overall, we find six genome-wide significant associations for critical illness due to COVID-19, using data from 6,179 cases and 1,483,780 controls from 16 studies (Extended Data Fig. 4). Nine genome-wide significant loci were detected for moderate to severe hospitalized COVID-19 (including five of the six critical illness loci) from an analysis of 13,641 cases of COVID-19 and 2,070,709 controls across 29 studies (Fig. 2a, top). Finally, seven loci reached genome-wide significance in the analysis using data for all available 49,562 reported cases of SARS-CoV-2 infection and 1,770,206 controls, using data from a total of 44 studies (Fig. 2a, bottom). The proportion of cases with non-European genetic ancestry for each of the three analyses was 23%, 29% and 22%. We report the results for the lead variants at the 13 loci in different ancestry-group meta-analyses in Supplementary Table 3. We note that two loci, tagged by lead variants rs1886814 and rs72711165, had higher allele frequencies in southeast Asian (rs1886814; 15%) and East Asian genetic ancestry (rs72711165; 8%) whereas the minor allele frequencies in European populations were less than 3%. This highlights the value of including data from diverse populations for genetic discovery. We discuss the replication of previous findings and the new discoveries from these three analyses in the Supplementary Note.

Variant effects on severity and susceptibility

We found no genome-wide significant sex-specific effects at the 13 loci. However, we did identify significant heterogeneous effects ($P < 0.004$) across studies for 3 out of the 13 loci (Methods), which probably reflects the differential ascertainment of cases (Supplementary Table 2). There was a small number of overlapping samples ($n = 8,380$ European ancestry; $n = 745$ East Asian ancestry) between controls from the genOMICC and the UK Biobank studies, but leave-one-out sensitivity analyses did not reveal any bias in the corresponding effect sizes or P values (Extended Data Fig. 5 and Supplementary Information).

We next wanted to better understand whether the 13 significant loci were acting through mechanisms that increased the susceptibility to infection or that affected the progression of symptoms towards more severe disease. For all 13 loci, we compared the lead variant (strongest association P value) odds ratios (ORs) for the risk-increasing allele across our different COVID-19 phenotype definitions.

Focusing on the two better powered analyses: all cases with a reported SARS-CoV-2 infection and all cases hospitalized due to COVID-19, we find that four of the loci have similar odds ratios between these two analyses (Methods and Supplementary Table 2). Such consistency suggests a stronger link to susceptibility to SARS-CoV-2 infection rather than to the development of severe COVID-19. The strongest susceptibility signal was the previously reported *ABO* locus (rs912805253)^{12,13,15,16}. Notably, and in agreement with a previously reported study¹⁵, we also report a locus within the 3p21.31 region that was more strongly associated with susceptibility to SARS-CoV-2 than progression to more severe COVID-19 phenotypes. rs2271616 showed a stronger association with a reported SARS-CoV-2 infection ($P = 1.79 \times 10^{-34}$; OR (95% confidence interval (CI)) = 1.15 (1.13–1.18)) than hospitalization ($P = 1.05 \times 10^{-5}$; OR (95% CI) = 1.12 (1.06–1.19)). For this locus—which contains additional independent signals—the linkage-disequilibrium (LD) pattern is discordant with the P -value expectation (Extended Data Fig. 6 and Supplementary Note), pointing to a key missing causal variant or to a potentially undiscovered multi-allelic or structural variant in this locus.

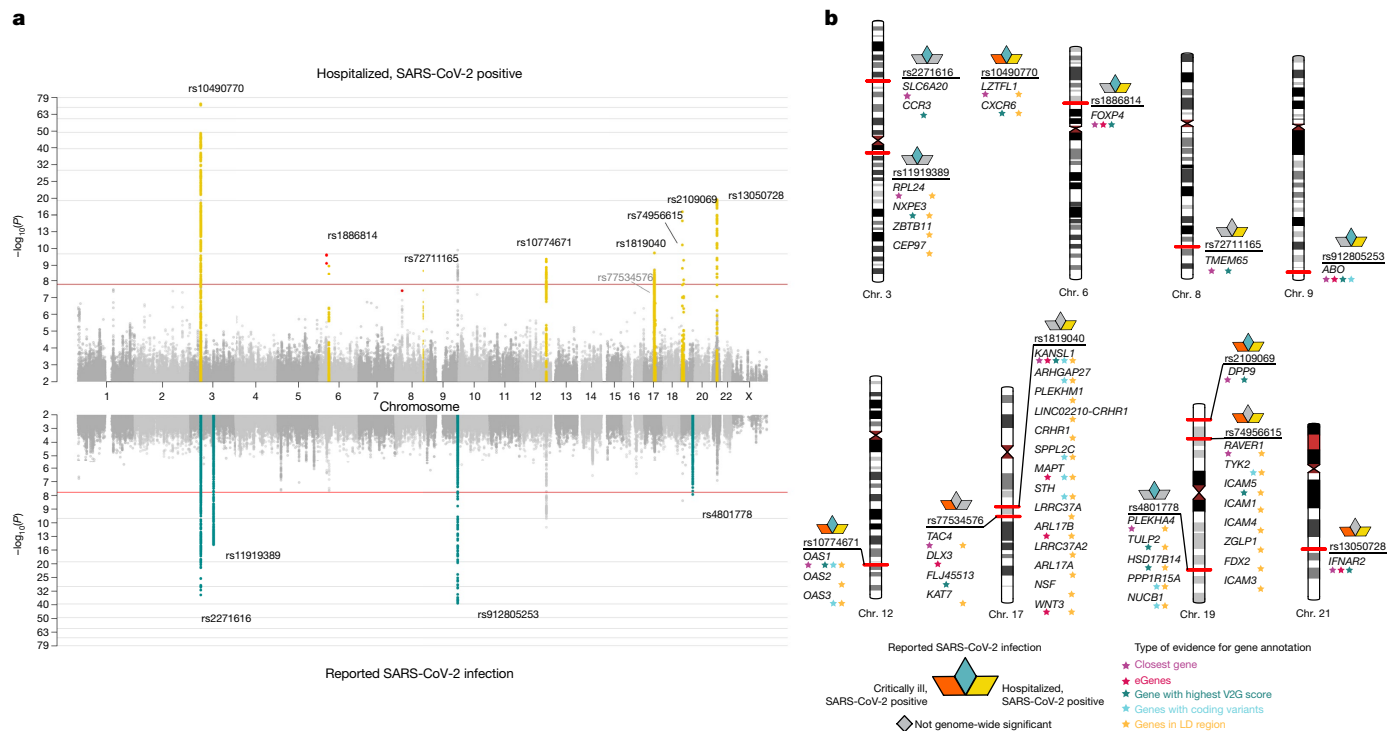


Fig. 2 | Genome-wide association results for COVID-19. **a**, Top, results of a genome-wide association study of hospitalized cases of COVID-19 ($n = 13,641$ cases and $n = 2,070,709$ controls). Bottom, the results of reported SARS-CoV-2 infections ($n = 49,562$ cases and $n = 1,770,206$ controls). Loci highlighted in yellow (top) represent regions associated with the severity of the COVID-19 manifestation—that is, increased odds of more severe COVID-19 phenotypes. Loci highlighted in green (bottom) are regions associated with susceptibility to a SARS-CoV-2 infection—that is, the effect is the same across mild and severe

By contrast, 9 out of the 13 loci were associated with increased risk of severe symptoms with significantly larger odds ratios for hospitalized COVID-19 compared with the mildest phenotype of reported SARS-CoV-2 infection (eight loci were below the threshold of $P < 0.004$ (test for effect size difference) and, in addition, the lead variant rs10774671 had a clear increase in odds ratios despite not passing this threshold) (Supplementary Table 2). We further compared the odds ratios for these nine loci for critical illness due to COVID-19 versus hospitalized due to COVID-19, and found that these loci exhibited a general increase in effect risk for critical illness (Methods, Extended Data Fig. 7a and Supplementary Table 4), but the lower power for association analysis of critically ill COVID-19 means that these results should be considered as suggestive. Overall, these results indicated that these nine loci were more likely to be associated with progression of the disease and worse outcome from SARS-CoV-2 infection compared to being associated with susceptibility to SARS-CoV-2 infection.

For some of these analyses, the controls were simply existing population controls without knowledge of SARS-CoV-2 infection or COVID-19 status, which may bias effect size estimates as some of these individuals may have either become infected with SARS-CoV-2 or developed COVID-19. We perform several sensitivity analyses (Extended Data Fig. 7b, Supplementary Note and Supplementary Table 4) in which we show that using population controls can be a valid and powerful strategy for host genetic discovery of infectious disease, and particularly those that are widespread and with rare severe outcomes.

Gene prioritization and association with other traits

To better understand the potential biological mechanism of each locus, we applied several approaches to prioritize candidate causal genes and

COVID-19 phenotypes. We highlight in red genome-wide significant variants that had high heterogeneity across contributing studies and that were therefore excluded from the list of loci found. **b**, Results of gene prioritization using different evidence measures of gene annotation. Genes in the LD region, genes with coding variants and eGenes (fine-mapped *cis*-eQTL variant $PIP > 0.1$ in GTEx Lung) are annotated if in LD with a COVID-19 lead variant ($r^2 > 0.6$). V2G, highest gene prioritized by the V2G score of Open Target Genetics.

explore additional associations with other diseases and traits. Of the 13 genome-wide significant loci, we found that nine loci implicated biologically plausible genes (Supplementary Tables 2, 5). Protein-altering variants in LD with lead variants implicated genes at six loci, including *TYK2* (chromosome and cytogenetic band (chr.) 19p13.2) and *PPP1R15A* (chr. 19q13.33). The COVID-19 lead variant rs74956615T>A in *TYK2*, which confers risk for critical illness (OR (95% CI) = 1.43 (1.29–1.59), $P = 9.71 \times 10^{-12}$) and hospitalization due to COVID-19 (OR (95% CI) = 1.27 (1.18–1.36), $P = 5.05 \times 10^{-10}$) is correlated with the missense variant rs34536443:G>C (p.Pro1104Ala; $r^2 = 0.82$). This is consistent with the primary immunodeficiency described with complete *TYK2* loss of function³ as this variant is known to reduce function^{18,19}. By contrast, this missense variant was previously reported to be protective against autoimmune diseases (Extended Data Fig. 8 and Supplementary Table 6), including rheumatoid arthritis (OR = 0.74, $P = 3.0 \times 10^{-8}$; UK Biobank SAIGE) and hypothyroidism (OR = 0.84, $P = 1.8 \times 10^{-10}$; UK Biobank). At the 19q13.33 locus, the lead variant rs4801778, which was significantly associated with a reported SARS-CoV-2 infection (OR (95% CI) = 0.95 (0.93–0.96), $P = 2.1 \times 10^{-8}$), is in LD ($r^2 = 0.93$) with a missense variant rs11541192:G>A (p.Gly312Ser) in *PPP1R15A*.

A lung-specific *cis*-expression quantitative trait loci (*cis*-eQTLs) from GTEx v.8²⁰ ($n = 515$) and the Lung eQTL Consortium²¹ ($n = 1,103$) provided further support for a subset of loci (Supplementary Table 7), including *FOXP4* (chr. 6p21.1) and *ABO* (chr. 9q34.2), *OAS1/OAS3/OAS2* (chr. 12q24.13) and *IFNAR2/IL10RB* (21q22.11), where the COVID-19-associated variants modify gene expression in lung. Furthermore, our phenome-wide association study (PheWAS) analysis (Supplementary Table 6) implicated three additional loci related to lung function, with modest lung eQTL evidence—that is, the lead variant was not fine-mapped but significantly associated. An intronic variant

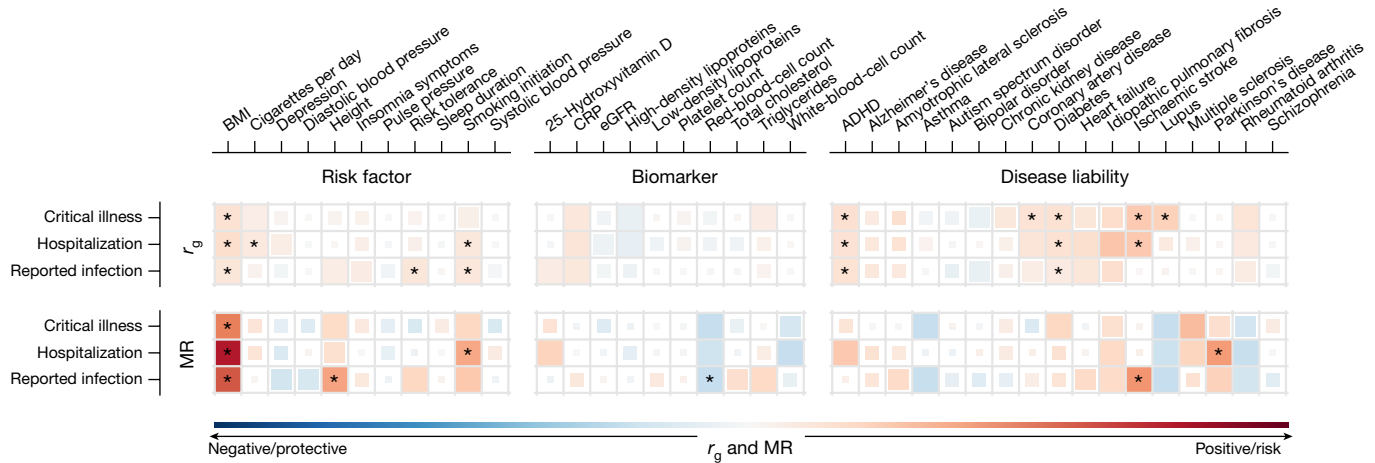


Fig. 3 | Genetic correlations and Mendelian randomization causal estimates between 38 traits and COVID-19 critical illness, hospitalization and reported SARS-CoV-2 infection. Larger squares correspond to P values with higher significance, with genetic correlations (r_g) or Mendelian randomization (MR) causal estimates significantly different from zero. The size of each coloured square indicates the magnitude of the P value, with $P < 0.05$ shown as a full-sized square, $P = 0.05-0.1$ as a large square, $P = 0.1-0.5$

as a medium square and $P > 0.5$ as a small square. Genetic correlations or causal estimates that are significantly different from zero at an FDR of 5% are marked with an asterisk. Two-sided P values were calculated using LDSC for genetic correlations and inverse-variance-weighted analysis for Mendelian randomization. ADHD, attention-deficit hyperactivity disorder; BMI, body mass index; CRP, C-reactive protein; eGFR, estimated glomerular filtration rate.

rs2109069:G>A in *DPP9* (chr. 19p13.3), which is positively associated with critical illness, was previously reported to be risk-increasing for interstitial lung disease (tag lead variant rs12610495:A>G (p.Leu8Pro); OR = 1.29, $P = 2.0 \times 10^{-12}$)⁵. The COVID-19 lead variant rs1886814:A>C in the *FOXP4* locus is correlated ($r^2 = 0.64$) with a lead variant of lung adenocarcinoma (tag variant is rs7741164; OR = 1.2, $P = 6.0 \times 10^{-13}$)^{6,22} and similarly with a lead variant reported for subclinical interstitial lung disease²³. In severe COVID-19, lung cancer and interstitial lung disease, the minor, expression-increasing allele is associated with increased risk. We also found that intronic variants (chr. 1q22) and rs1819040:T>A in *KANSL1* (chr. 17q21.31), associated with protection against hospitalization due to COVID-19, were previously reported for reduced lung function (for example, tag lead variant rs141942982:G>T; OR (95% CI) = 0.96 (0.95–0.97), $P = 1.00 \times 10^{-20}$)⁷. Notably, the 17q21.31 locus is a well-known locus for structural variants containing a megabase inversion polymorphism (H1 and inverted H2 forms) and complex copy-number variations, in which the inverted H2 forms were shown to be positively selected in European individuals^{24,25}.

Lastly, there are two loci in the 3p21.31 region with varying genes prioritized by different methods for different independent signals. For the severity lead variant rs10490770:T>C, we prioritized *CXCR6* with the Variant2Gene (V2G) algorithm²⁶, although *LZTFL1* is the closest gene. The *CXCR6* has a role in chemokine signalling²⁷ and *LZTFL1* has been implicated in lung cancer²⁸. rs2271616:G>T, which is associated with susceptibility, tags a complex region including several independent signals (Supplementary Note) that are all located within the gene body of *SLC6A20*, which encodes a protein that is known to functionally interact with the SARS-CoV-2 receptor ACE2²⁹. However, none of the lead variants in the 3p21.31 region has been previously associated with other traits or diseases in our PheWAS analysis. Although these results provide supporting in silico evidence for candidate causal gene prioritization, further functional characterization is needed. Detailed locus descriptions and LocusZoom plots are provided in Supplementary Fig. 2.

Polygenic architecture of COVID-19

To further investigate the genetic architecture of COVID-19, we used results from meta-analyses including samples from European ancestries

(sample sizes are described in the Methods and Supplementary Table 1) to estimate the heritability explained by common single-nucleotide polymorphisms—that is, the proportion of variation in the two phenotypes that was attributable to common genetic variants—and to determine whether heritability of COVID-19 phenotypes was enriched in genes that were specifically expressed in certain tissues³⁰ from the GTEx dataset³¹. We detected low, but significant, heritability across all three analyses (<1% on observed scale, all P values were $P < 0.0001$) (Supplementary Table 8). The values are low compared to previously published studies¹⁴, but may be explained by differences in the reported estimate scale (observed versus liability), the specific method used, disease-prevalence estimates, phenotypic differences between patient cohorts or ascertainment of controls. Despite the low reported values, we found that heritability of a reported SARS-CoV-2 infection was significantly enriched in genes that were specifically expressed in the lung ($P = 5.0 \times 10^{-4}$) (Supplementary Table 9). These findings, together with the genome-wide significant loci identified in the meta-analyses, suggest that there is a significant polygenic architecture that can be better leveraged with future, larger, sample sizes.

Genetic correlation and Mendelian randomization

Genetic correlations (r_g) between the three COVID-19 phenotypes was high, although lower correlations were observed between hospitalized COVID-19 and reported SARS-CoV-2 infection (critical illness versus hospitalized: r_g (95% CI) = 1.37 (1.08–1.65), $P = 2.9 \times 10^{-21}$; critical illness versus reported SARS-CoV-2 infection, r_g (95% CI) = 0.96 (0.71–1.20), $P = 1.1 \times 10^{-14}$; hospitalized versus reported SARS-CoV-2 infection: r_g (95% CI) = 0.85 (0.68–1.02), $P = 1.1 \times 10^{-22}$). To better understand which traits are genetically correlated and/or potentially causally associated with COVID-19 hospitalization, critical illness and reported SARS-CoV-2 infection, we chose a set of 38 disease, health and neuropsychiatric phenotypes as potential COVID-19 risk factors based on their clinical correlation with disease susceptibility, severity or mortality (Supplementary Table 10).

We found evidence (false-discovery rate (FDR) < 0.05) of significant genetic correlations between nine traits and hospitalized COVID-19 and reported SARS-CoV-2 infection (Fig. 3, Extended Data Fig. 9 and Supplementary Table 11). Notably, genetic liability to ischaemic

stroke was only significantly positively correlated with critical illness or hospitalization due to COVID-19, but not with a higher likelihood of reported SARS-CoV-2 infection (infection $r_g = 0.019$ versus hospitalization $r_g = 0.41$, $z = 2.7$, $P = 0.006$; infection $r_g = 0.019$ versus critical illness $r_g = 0.40$, $z = 2.49$, $P = 0.013$).

We next used two-sample Mendelian randomization to infer potentially causal relationships between these traits. After correcting for multiple testing (FDR < 0.05), eight exposure–COVID-19 trait pairs showed suggestive evidence of a causal association (Fig. 3, Extended Data Fig. 10, Supplementary Table 12 and Supplementary Fig. 3). Five of these associations were robust to potential violations of the underlying assumptions of Mendelian randomization. Corroborating our genetic correlation results and evidence from epidemiological studies, genetically predicted higher body-mass index (OR (95% CI) = 1.4 (1.3–1.6), $P = 8.5 \times 10^{-11}$) and smoking (OR (95% CI) = 1.9 (1.3–2.8), $P = 0.0012$) were associated with increased risk of COVID-19 hospitalization, with body-mass index also being associated with increased risk of SARS-CoV-2 infection (OR (95% CI) = 1.1 (1.1–1.2), $P = 4.8 \times 10^{-7}$). Genetically predicted increased height (OR (95% CI) = 1.1 (1–1.1), $P = 8.9 \times 10^{-4}$) was associated with an increased risk of reported SARS-CoV-2 infection, whereas a genetically predicted higher red-blood-cell count (OR (95% CI) = 0.93 (0.89–0.96), $P = 5.7 \times 10^{-5}$) was associated with a reduced risk of reported SARS-CoV-2 infection. Despite evidence of a genetic correlation between type II diabetes and COVID-19 outcomes, there was no evidence of a causal association in the Mendelian randomization analyses, which suggests that the observed genetic correlations are due to pleiotropic effects between body-mass index and type 2 diabetes. Further sensitivity analyses relating to sample overlap are discussed in the Supplementary Information.

Discussion

The COVID-19 HGI has brought together investigators from across the world to advance genetic discovery for SARS-CoV-2 infection and severe COVID-19 disease. We report 13 genome-wide significant loci associated with some aspect of SARS-CoV-2 infection or COVID-19. Many of these loci overlap with previously reported associations with lung-related phenotypes or autoimmune or inflammatory diseases, but some loci have no obvious candidate gene.

Four out of the thirteen genome-wide significant loci showed similar effects in the reported SARS-CoV-2 infection analysis (a proxy for disease susceptibility) and all-hospitalized COVID-19 (a proxy for disease severity). Of these, one locus was in close proximity to, yet independent of, the major genetic signal for COVID-19 severity at the 3p21.31 locus. Notably, this locus was associated with COVID-19 susceptibility rather than severity. The locus overlaps *SLC6A20*, which encodes an amino acid transporter that interacts with ACE2. Nonetheless, we caution that more data are needed to resolve the nature of the relationship between genetic variation and COVID-19 at this locus, particularly as the physical proximity, LD structure and patterns of association suggest that untagged genetic variation could drive the association signal in the region. Our findings support the notion that some genetic variants, most notably at the *ABO* and *PPP1R15A* loci, in addition to *SLC6A20*, can indeed affect susceptibility to infection rather than progression to severe COVID-19 once infected.

Several of the loci reported here—as noted in previous publications^{12,14}—intersect with well-known genetic variants that have established genetic associations. Examples of these include variants at *DPP9* and *FOXP4*, which show previous evidence of increasing risk for interstitial lung disease⁵, and missense variants within *TYK2* that show a protective effect on several autoimmune-related diseases^{32–35}. Together with the heritability enrichment observed in genes expressed in lung tissues, these results highlight the involvement of lung-related biological pathways in the development of severe COVID-19. Several other loci show no previously documented genome-wide significant

associations, despite the high significance and attractive candidate genes for COVID-19 (for example, *CXCR6*, *LZTFL1*, *IFNAR2* and *OAS1/OAS2/OAS3* loci). The previously reported associations for the strongest association for COVID-19 severity at the 3p21.31 locus and monocytes count are likely to be due to proximity and not a true co-localization.

Increasing the global representation in genetic studies enhances the ability to detect novel associations. Two of the loci that affect disease severity were only discovered by including the four studies of individuals with East Asian ancestry. One of these loci—close to *FOXP4*—is common particularly in East Asian participants (32%) as well as admixed American participants in the Americas (20%) and Middle Eastern participants (7%), but has a low frequency in most European ancestries (2–3%) in our data. Although we cannot be certain of the mechanism of action, the *FOXP4* association is an attractive biological target, as it is expressed in the proximal and distal airway epithelium³⁶ and has been shown to have a role in controlling epithelial cell fate during lung development³⁷. The COVID-19 HGI continues to pursue expansion of the datasets included in the analyses of the consortium to populations from underrepresented populations in upcoming data releases. We plan to release ancestry-specific results in full once the sample sizes allow for a well-powered meta-analysis.

Care should be taken when interpreting the results from a meta-analysis because of challenges with case and control ascertainment and collider bias (see Supplementary Note for a more detailed discussion on study limitations). Drawing a comprehensive and reproducible map of the host genetics factors associated with COVID-19 severity and SARS-CoV-2 requires a sustained international effort to include diverse ancestries and study designs. To accelerate downstream research and therapeutic discovery, the COVID-19 HGI regularly publishes meta-analysis results from periodic data freezes on the website <https://www.covid19hg.org/> and provides an interactive explorer through which researchers can browse the results and the genomic loci in more detail. Future work will be required to better understand the biological and clinical value of these findings. Continued efforts to collect more samples and detailed phenotypic data should be endorsed globally, allowing for more thorough investigation of variable, heritable symptoms, particularly in light of the newly emerging strains of SARS-CoV-2, which may provoke different host responses that lead to disease.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41586-021-03767-x>.

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General Hospital, Edinburgh, UK. ⁶⁰¹Mid and South Essex NHS Foundation Trust, Basildon, UK. ⁶⁰²Hinchingbrooke Hospital, North West Anglia NHS Foundation Trust, Peterborough, UK. ⁶⁰³Royal Preston Hospital, Lancashire Teaching Hospitals NHS Foundation Trust, Preston, UK. ⁶⁰⁴University Hospital (Coventry), University Hospitals Coventry and Warwickshire, Coventry, UK. ⁶⁰⁵The Walton Centre, The Walton Centre, Liverpool, UK. ⁶⁰⁶Hull Royal Infirmary, Hull University Teaching Hospital Trust, Hull, UK. ⁶⁰⁷Darlington Memorial Hospital, County Durham and Darlington Foundation Trust, Darlington, UK. ⁶⁰⁸Queen Elizabeth Hospital (Gateshead), Gateshead NHS Foundation Trust, Newcastle, UK. ⁶⁰⁹Warrington Hospital, Warrington & Halton Hospitals NHS Foundation Trust, Warrington, UK. ⁶¹⁰University Hospitals Bristol and Weston NHS Foundation Trust, Bristol, UK. ⁶¹¹St Mary's Hospital (Isle of Wight), Isle of Wight NHS Trust, Isle of Wight, UK. ⁶¹²The Maidstone Hospital, Maidstone & Tunbridge Wells NHS Trust, Maidstone, UK. ⁶¹³Huddersfield Royal, Calderdale and Huddersfield NHS Foundation Trust, Huddersfield, UK. ⁶¹⁴Royal Surrey County Hospital, Guildford, UK. ⁶¹⁵Countess of Chester Hospital, Countess of Chester Hospital NHS Foundation Trust, Chester, UK. ⁶¹⁶Frimley Park Hospital, Frimley Health Foundation Trust, Frimley, UK. ⁶¹⁷Leeds General Infirmary, Leeds Teaching Hospitals, Leeds, UK. ⁶¹⁸North Middlesex Hospital, North Middlesex University Hospital NHS Trust, London, UK. ⁶¹⁹Arrowe Park Hospital, Wirral University Teaching Hospital NHS Foundation Trust, Wirral, UK. ⁶²⁰Great Ormond Street Hospital, Great Ormond Street Hospital for Children NHS Foundation Trust, London, UK. ⁶²¹Royal Shrewsbury Hospital, Shrewsbury and Telford Hospital NHS Trust, Shrewsbury, UK. ⁶²²East Surrey Hospital (Redhill), Surrey & Sussex Healthcare, Redhill, UK. ⁶²³Burton Hospital, University Hospitals of Derby & Burton NHS Foundation Trust, Burton-on-Trent, UK. ⁶²⁴Kent and Canterbury Hospital, East Kent Hospitals NHS Foundation Trust, Canterbury, UK. ⁶²⁵Weston Area General Trust, University Hospitals Bristol and Weston NHS Foundation Trust, Bristol, UK. ⁶²⁶Luton and Dunstable University Hospital, Luton, UK. ⁶²⁷Glasgow Royal Infirmary, Greater Glasgow and Clyde, Glasgow, UK. ⁶²⁸Derbyshire Healthcare, Derbyshire Healthcare NHS Foundation Trust, Derby, UK. ⁶²⁹Macclesfield General Hospital, East Cheshire NHS Foundation Trust, Macclesfield, UK. ⁶³⁰Chelsea and Westminster Hospital, Chelsea and Westminster NHS Trust, London, UK. ⁶³¹Institute of Microbiology and Infection, University of Birmingham, Birmingham, UK. ⁶³²Prince Philip Hospital, Hywel Dda University Health Board, Llanelli, UK. ⁶³³George Eliot Hospital - Acute Services, George Eliot Hospital, Nuneaton, UK. ⁶³⁴Kettering General Hospital, Kettering General Hospital NHS Foundation Trust, Kettering, UK. ⁶³⁵Birmingham Heartlands Hospital, Birmingham, UK. ⁶³⁶Russells Hall Hospital, The Dudley Group NHS Foundation Trust, Dudley, UK. ⁶³⁷Harefield Hospital, Royal Brompton & Harefield Trust, London, UK. ⁶³⁸Lister Hospital, East and North Hertfordshire NHS Trust, Stevenage, UK. ⁶³⁹Musgrove Park Hospital (Taunton & Somerset), Somerset NHS Foundation Trust, Taunton, UK. ⁶⁴⁰Queen's Hospital, Havering (Romford), Barking, Havering and Redbridge University Hospitals NHS Trust, London, UK. ⁶⁴¹Southport & Formby District General Hospital, Southport and Ormskirk Hospital NHS Trust, Southport, UK. ⁶⁴²New Cross Hospital, The Royal Wolverhampton NHS Trust, Wolverhampton, UK. ⁶⁴³King's College Hospital, London, UK. ⁶⁴⁴The Royal Victoria Infirmary, Newcastle Hospitals NHS Trust, Newcastle, UK. ⁶⁴⁵The Great Western Hospital,

Great Western Hospitals NHS Foundation Trust, Swindon, UK. ⁶⁴⁶Ninewells Hospital, Tayside, Dundee, UK. ⁶⁴⁷Poole Hospital NHS Trust, Poole, UK. ⁶⁴⁸Burton Hospital, University Hospitals of Derby & Burton NHS Foundation Trust, Derby, UK. ⁶⁴⁹William Harvey Hospital, Ashford, East Kent Hospitals NHS Foundation Trust, Willesborough, UK. ⁶⁵⁰King's Mill Hospital, Sherwood Forest Hospitals NHS Foundation Trust, Sutton-in-Ashfield, UK. ⁶⁵¹Liverpool Women's NHS Foundation Trust, Liverpool, UK. ⁶⁵²Dewsbury Hospital, Mid Yorkshire Hospitals NHS Trust, Dewsbury, UK. ⁶⁵³Northern Devon District Hospital, Northern Devon Healthcare NHS Trust, Barnstaple, UK. ⁶⁵⁴Tameside General Hospital, Tameside and Glossop Integrated Care NHS Foundation Trust, Manchester, UK. ⁶⁵⁵Sandwell General Hospital, Sandwell and West Birmingham Hospitals NHS Trust, Birmingham, UK. ⁶⁵⁶Broomfield Hospital, Mid and South Essex University Hospitals Group, Broomfield, UK. ⁶⁵⁷Wycombe Hospital, Buckingham Healthcare NHS Trust, Wycombe, UK. ⁶⁵⁸University Hospital of North Tees, North Tees and Hartlepool NHS Trust, Stockton-on-Tees, UK. ⁶⁵⁹Royal Manchester Children's Hospital, Manchester University Hospitals NHS Foundation Trust, Manchester, UK. ⁶⁶⁰Bedford Hospital, Bedford, UK. ⁶⁶¹Colchester General Hospital, East Suffolk and North Essex Foundation Trust, Colchester, UK. ⁶⁶²Queen Elizabeth Hospital (Birmingham) and Heartlands, University Hospital Birmingham NHS Foundation Trust, Birmingham, UK. ⁶⁶³Chesterfield Royal Hospital, Chesterfield Royal Hospital NHS Foundation Trust, Chesterfield, UK. ⁶⁶⁴Princess Alexandra Hospital, The Princess Alexandra Hospital NHS Trust, Harlow, UK. ⁶⁶⁵Watford General Hospital, West Hertfordshire Hospitals NHS Trust, Watford, UK. ⁶⁶⁶Milton Keynes Hospital, Milton Keynes University Hospital NHS Foundation Trust, Milton Keynes, UK. ⁶⁶⁷Royal Bolton General Hospital, Bolton Foundation Trust, Bolton, UK. ⁶⁶⁸Royal Gwent (Newport), Aneurin Bevan University Health Board, Newport, UK. ⁶⁶⁹The Royal Marsden Hospital (London), The Royal Marsden NHS Foundation Trust, London, UK. ⁶⁷⁰Queen Victoria Hospital (East Grinstead), Queen Victoria Hospital NHS Foundation Trust, East Grinstead, UK. ⁶⁷¹County Hospital (Stafford), University Hospitals of North Midlands NHS Trust, Stafford, UK. ⁶⁷²Whiston Hospital, St Helen's & Knowsley Hospitals NHS Trust, Prescot, UK. ⁶⁷³Croydon University Hospital, London, UK. ⁶⁷⁴Gloucester Royal, Gloucestershire Hospitals NHS Foundation Trust, Gloucester, UK. ⁶⁷⁵Medway Maritime Hospital, Medway Maritime NHS Trust, Gillingham, UK. ⁶⁷⁶Royal Papworth Hospital Everard, Royal Papworth Hospital NHS Foundation Trust, Cambridge, UK. ⁶⁷⁷Derriford (Plymouth), University Hospital Plymouth NHS Trust, Plymouth, UK. ⁶⁷⁸St Helier Hospital, Epsom and St Helier University Hospital NHS Trust, London, UK. ⁶⁷⁹Royal Berkshire Hospital, Royal Berkshire Foundation Trust, London, UK. ⁶⁸⁰Bradford Royal Infirmary, Bradford Teaching Hospitals NHS Foundation Trust, Bradford, UK. 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Livingston, UK. ⁷⁴⁸Aberdeen Royal Infirmary, Aberdeen, UK. ⁷⁴⁹Addenbrooke's Hospital, Cambridge, UK. ⁷⁵⁰Aintree University Hospital, Liverpool, UK. ⁷⁵²Arrow Park Hospital, Wirral, UK. ⁷⁵³Ashford and St Peter's Hospital, Lyne, UK. ⁷⁵⁴Basingstoke and North Hampshire Hospital, Basingstoke, UK. ⁷⁵⁵Borders General Hospital, Melrose, UK. ⁷⁵⁶Chesterfield Royal Hospital Foundation Trust, Chesterfield, UK. ⁷⁵⁷Eastbourne District General Hospital, East Sussex, UK and Conquest Hospital, Eastbourne, UK. ⁷⁵⁸Barnsley Hospital, Barnsley, UK. ⁷⁵⁹Blackpool Victoria Hospital, Blackpool, UK. ⁷⁶⁰East Surrey Hospital, Redhill, UK. ⁷⁶¹Good Hope Hospital, Birmingham, UK. ⁷⁶²Hereford County Hospital, Hereford, UK. ⁷⁶³Hull Royal Infirmary, Hull, UK. ⁷⁶⁵Kent & Canterbury Hospital, Canterbury, UK. ⁷⁶⁶Manchester Royal Infirmary, Manchester, UK. ⁷⁶⁷Nottingham University Hospital, Nottingham, UK. ⁷⁶⁸Pilgrim Hospital, Lincoln, UK. ⁷⁶⁹Queen Elizabeth Hospital, Birmingham, UK. ⁷⁷⁰Salford Royal Hospital, Manchester, UK. ⁷⁷¹Tameside General Hospital, Ashton-under-Lyne, UK. ⁷⁷²The Tunbridge Wells Hospital and Maidstone Hospital, Maidstone, UK. ⁷⁷³The Royal Oldham Hospital, Manchester, UK. ⁷⁷⁴The Royal Papworth Hospital, Cambridge, UK. ⁷⁷⁵University College Hospital, London, UK. ⁷⁷⁶Withybush General Hospital, Haverfordwest, UK. ⁷⁷⁷Wythenshawe Hospital, Manchester, UK. ⁷⁷⁸Yeovil Hospital, Yeovil, UK. ⁷⁷⁹Cumberland Infirmary, Carlisle, UK. ⁷⁸⁰Darent Valley Hospital, Dartford, UK. ⁷⁸¹Dumfries and Galloway Royal Infirmary, Dumfries, UK. ⁷⁸²Ealing Hospital, London, UK. ⁷⁸³Fairfield General Hospital, Bury, UK. ⁷⁸⁴George Eliot Hospital NHS Trust, Nuneaton, UK. ⁷⁸⁵Clan Clwyd Hospital, Bodelwyddan, UK. ⁷⁸⁶Glangwili General Hospital, Camarthen, UK. ⁷⁸⁷The Great Western Hospital, Swindon, UK. ⁷⁸⁸Guys and St Thomas' Hospital, London, UK. ⁷⁸⁹Harefield Hospital, London, UK. ⁷⁹⁰Harrogate and District NHS Foundation Trust, Harrogate, UK. ⁷⁹²James Paget University Hospital NHS Trust, Great Yarmouth, UK. ⁷⁹⁴King's Mill Hospital, Nottingham, UK. ⁷⁹⁵Kingston Hospital, Kingston, UK. ⁷⁹⁶Lincoln County Hospital, Lincoln, UK. ⁷⁹⁷Liverpool Heart and Chest Hospital, Liverpool, UK. ⁷⁹⁸Macclesfield District General Hospital, Macclesfield, UK. ⁷⁹⁹Medway Maritime Hospital, Gillingham, UK. ⁸⁰⁰Milton Keynes University Hospital, Milton Keynes, UK. ⁸⁰¹Morrison Hospital, Swansea, UK. ⁸⁰²National Hospital for Neurology and Neurosurgery, London, UK. ⁸⁰³Norfolk and Norwich University Hospital (NNUH), Norwich, UK. ⁸⁰⁴North Middlesex University Hospital NHS Trust, London, UK. ⁸⁰⁶Northumbria Healthcare NHS Foundation Trust, North Shields, UK. ⁸⁰⁷Peterborough City Hospital, Peterborough, UK. ⁸⁰⁸Prince Charles Hospital, Merthyr Tydfil, UK. ⁸⁰⁹Royal Sussex County Hospital, Brighton, UK. ⁸¹⁰Princess Royal Hospital, Haywards Heath, UK. ⁸¹¹Princess of Wales Hospital, Llantrisant, UK. ⁸¹²Queen Alexandra Hospital, Portsmouth, UK. ⁸¹³Queen Elizabeth Hospital, London, UK. ⁸¹⁵Queen Victoria Hospital, East Grinstead, UK. ⁸¹⁶Queen's Hospital Burton, Burton-On-Trent, UK. ⁸¹⁷Raigmore Hospital, Inverness, UK. ⁸¹⁸Rotherham General Hospital, Rotherham, UK. ⁸¹⁹Royal Blackburn Teaching Hospital, Blackburn, UK. ⁸²⁰Royal Preston Hospital, Preston, UK. ⁸²¹Royal Surrey County Hospital, Guildford, UK. ⁸²²Royal Albert Edward Infirmary, Wigan, UK. ⁸²³The Royal Alexandra Children's Hospital, Brighton, UK. ⁸²⁴Royal Alexandra Hospital, Paisley, UK. ⁸²⁵Royal Bolton Hospital, Bolton, UK. ⁸²⁶University Hospitals Dorset NHS Foundation Trust, Dorchester, UK. ⁸²⁷Royal Brompton Hospital, London, UK. ⁸²⁸Imperial College London, London, UK. ⁸²⁹Royal Cornwall Hospital, Truro, UK. ⁸³⁰Royal Free Hospital, London, UK. ⁸³¹Royal Glamorgan Hospital, Pontyclun, UK. ⁸³²Royal Gwent Hospital, Newport, UK. ⁸³³Northern General Hospital, Sheffield, UK. ⁸³⁴Royal Hampshire County Hospital, Winchester, UK. ⁸³⁵Royal Manchester Children's Hospital, Manchester, UK. ⁸³⁶Royal Stoke University Hospital, Stoke-on-Trent, UK. ⁸³⁷Salisbury District Hospital, Salisbury, UK. ⁸³⁸Sandwell General Hospital, Birmingham, UK. ⁸³⁹Scarborough General Hospital, Scarborough, UK. ⁸⁴⁰Scunthorpe General Hospital, Scunthorpe, UK. ⁸⁴¹Southmead Hospital, Bristol, UK. ⁸⁴²St George's Hospital, London, UK. ⁸⁴³St Mary's Hospital, Newport, UK. ⁸⁴⁴Stoke Mandeville Hospital, Aylesbury, UK. ⁸⁴⁵Sunderland Royal Hospital, Sunderland, UK. ⁸⁴⁶Alexandra Hospital, Redditch and Worcester Royal Hospital, Worcester, UK. ⁸⁴⁷The Christie NHS Foundation Trust, Manchester, UK. ⁸⁴⁸The Queen Elizabeth Hospital, King's Lynn, UK. ⁸⁴⁹The Royal Liverpool University Hospital, Liverpool, UK. ⁸⁵⁰The Royal Marsden NHS Foundation Trust, London, UK. ⁸⁵¹Torbay Hospital, Torquay, UK. ⁸⁵²University Hospital Monklands, Airdrie, UK. ⁸⁵³University Hospital Lewisham, London, UK. ⁸⁵⁴University Hospital North Durham, Darlington, UK. ⁸⁵⁵University Hospital of North Tees, Stockton-on-Tees, UK. ⁸⁵⁶University Hospital of Wales, Cardiff, UK. ⁸⁵⁷University Hospital Wishaw, Wishaw, UK. ⁸⁵⁸Victoria Hospital, Kirkcaldy, UK. ⁸⁵⁹Warrington General Hospital, Warrington, UK. ⁸⁶⁰West Cumberland Hospital, Whitehaven, UK. ⁸⁶¹Western Sussex Hospitals, Chichester, UK. ⁸⁶²Whiston Hospital, Prescot, UK. ⁸⁶³York Hospital, York, UK. ⁸⁶⁴Ysbyty Gwynedd, Bangor, UK. ⁸⁶⁵Countess of Chester Hospital, Chester, UK. ⁸⁶⁶Croydon University Hospital, Croydon, UK. ⁸⁶⁷Diana Princess of Wales Hospital, Grimsby, UK. ⁸⁶⁸Dorset County Hospital, Dorchester, UK. ⁸⁶⁹Forth Valley Royal Hospital, Falkirk, UK. ⁸⁷⁰Furness General Hospital, Barrow-in-Furness, UK. ⁸⁷¹Alder Hey Children's Hospital, Liverpool, UK. ⁸⁷²Derriford Hospital, Plymouth, UK. ⁸⁷³Glasgow Royal Infirmary, Glasgow, UK. ⁸⁷⁴Glenfield Hospital, Leicester, UK. ⁸⁷⁵Gloucestershire Royal Hospital, Gloucester, UK. ⁸⁷⁶Golden Jubilee National Hospital, Clydebank, UK. ⁸⁷⁷Great Ormond St Hospital and UCL Great Ormond St Institute of Child Health NIHR Biomedical Research Centre, London, UK. ⁸⁷⁸Homerton University Hospital Foundation NHS Trust, London, UK. ⁸⁷⁹James Cook University Hospital, Middlesbrough, UK. ⁸⁸⁰John Radcliffe Hospital, Oxford, UK. ⁸⁸¹Leicester Royal Infirmary, Leicester, UK. ⁸⁸²Lister Hospital, Stevenage, UK. ⁸⁸³New Cross Hospital, Wolverhampton, UK. ⁸⁸⁴Royal Victoria Infirmary, Newcastle Upon Tyne, UK. ⁸⁸⁵Ninewells Hospital, Dundee, UK. ⁸⁸⁶North Devon District Hospital, Barnstaple, UK. ⁸⁸⁷North Manchester General Hospital, Manchester, UK. ⁸⁸⁸Northwick Park Hospital, London, UK. ⁸⁸⁹Prince Philip Hospital, Lianelli, UK. ⁸⁹⁰Pinderfields General Hospital, Wakefield, UK. ⁸⁹¹Poole Hospital, Poole, UK. ⁸⁹²Royal Shrewsbury Hospital, Shrewsbury, UK. ⁸⁹³Princess Royal Hospital, Telford, UK. ⁸⁹⁹Queen Elizabeth Hospital Gateshead, Gateshead, UK. ⁹⁰⁰Queen Elizabeth University Hospital, Glasgow, UK. ⁹⁰¹Royal Berkshire NHS Foundation Trust, Reading, UK. ⁹⁰²Royal Derby Hospital, Derby, UK. ⁹⁰³Royal Devon and Exeter Hospital, Exeter, UK. ⁹⁰⁴Royal Infirmary of Edinburgh, Edinburgh, UK. ⁹⁰⁵Royal Lancaster Infirmary, Lancaster, UK. ⁹⁰⁶Royal United Hospital, Bath, UK. ⁹⁰⁷Russells Hall Hospital, Dudley, UK. ⁹⁰⁸Sheffield Children's Hospital, Sheffield, UK. ⁹⁰⁹Southampton General Hospital, Southampton, UK. ⁹¹⁰Southend University Hospital, Westcliff-on-Sea, UK. ⁹¹¹Southport and Forby District General Hospital, Ormskirk, UK. ⁹¹²St James's University Hospital and Leeds General Infirmary, Leeds, UK. ⁹¹³Bristol Royal Infirmary, Bristol, UK. ⁹¹⁴Stepping Hill Hospital, Stockport, UK. ⁹¹⁵The Princess Alexandra Hospital, Harlow, UK. ⁹¹⁶University Hospital Crosshouse, Kilmarnock, UK. ⁹¹⁷University Hospital Hairmyres, East Kilbride, UK. ⁹¹⁸Craigavon Area Hospital, Craigavon, UK. ⁹¹⁹Warwick Hospital, Warwick, UK. ⁹²⁰West Middlesex Hospital, Isleworth, UK. ⁹²²Whittington Hospital, London, UK. ⁹²³William Harvey Hospital, Ashford, UK. ⁹³²Section of Molecular Virology, Imperial College London,

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Methods

Contributing studies

All of the participants were recruited following protocols approved by local Institutional Review Boards; this information is collected in Supplementary Table 1 for all 46 studies. All protocols followed local ethics recommendations and informed consent was obtained when required. Information about sample numbers, sex and age from for each contributing study is given in Supplementary Table 1. In total, 16 studies contributed data to the analysis of critical illness due to COVID-19, 29 studies contributed data to hospitalized COVID-19 analysis and 44 studies contributed to the analysis of all cases of COVID-19. Each individual study that contributed data to a particular analysis met a minimum threshold of 50 cases, as defined by the phenotypic criteria, for statistical robustness. The effective sample sizes for each ancestry group shown in Fig. 1 were calculated for display using the formula: $((4 \times N_{\text{case}} \times N_{\text{control}}) / (N_{\text{case}} + N_{\text{control}}))$. Details of contributing research groups are provided in Supplementary Table 1.

Phenotype definitions

COVID-19 disease status (critical illness and hospitalization status) was assessed following the Diagnosis and Treatment Protocol for Novel Coronavirus Pneumonia³⁸. The critically ill COVID-19 group included patients who were hospitalized owing to symptoms associated with laboratory-confirmed SARS-CoV-2 infection and who required respiratory support or whose cause of death was associated with COVID-19. The hospitalized COVID-19 group included patients who were hospitalized owing to symptoms associated with laboratory-confirmed SARS-CoV-2 infection.

The reported SARS-CoV-2 infection group included individuals with laboratory-confirmed SARS-CoV-2 infection or electronic health record, ICD coding or clinically confirmed COVID-19, or self-reported COVID-19 (for example, by questionnaire), with or without symptoms of any severity. Genetic-ancestry-matched control individuals for the three case definitions were sourced from population-based cohorts, including individuals whose exposure status to SARS-CoV-2 was either unknown or infection-negative for questionnaire/electronic-health-record-based cohorts. Additional information regarding individual studies contributing to the consortium are described in Supplementary Table 1.

Genome-wide association studies and meta-analyses

Each contributing study genotyped the samples and performed quality controls, data imputation and analysis independently, but following the consortium recommendations (information is available at <https://www.covid19hg.org/>). We recommended that genome-wide association study (GWAS) analyses were run using Scalable and Accurate Implementation of GEneralized mixed model (SAIGE)³⁹ on chromosomes 1–22 and X. The recommended analysis tool was SAIGE, but studies also used other software such as PLINK⁴⁰. The suggested covariates were age, age², sex, age × sex and the 20 first principal components. Any other study-specific covariates to account for known technical artefacts could be added. SAIGE automatically accounts for sample relatedness and case–control imbalances. Quality-control and analysis approaches for individual studies are reported in Supplementary Table 1.

Study-specific summary statistics were then processed for meta-analysis. Potential false positives, inflation and deflation were examined for each submitted GWAS. Allele frequency plots against gnomAD 3.0 genomes were manually inspected for each study. Standard error values as a function of the effective sample size were used to find studies that deviated from the expected trend. Summary statistics passing this manual quality control were included in the meta-analysis. Variants with an allele frequency of >0.1% and an imputation INFO score of >0.6 were carried forward from each study. Variants and alleles were lifted over to genome build GRCh38, if needed, and harmonized to

gnomAD 3.0 genomes⁴¹ by finding matching variants by strand flipping or switching the ordering of alleles. If multiple matching variants were included, the best match was chosen according to the minimum fold change in absolute allele frequency. Meta-analysis was performed using the inverse-variance-weighted (IVW) method on variants that were present in at least two-thirds of the studies contributing to the phenotype analysis. The method summarizes effect sizes across the multiple studies by computing the mean of the effect sizes weighted by the inverse variance in each individual study.

We report 13 meta-analysis variants that pass the genome-wide significance threshold after adjusting the threshold for multiple traits tested ($P < 5 \times 10^{-8}/3$). We report the unadjusted *P* values for each variant. We tested for heterogeneity between estimates from contributing studies using Cochran's *Q*-test^{42,43}. This is calculated for each variant as the weighted sum of squared differences between the effect sizes and their meta-analysis effect, the weights being the inverse variance of the effect size. *Q* is distributed as a χ^2 statistic with *k* (number of studies) minus one degrees of freedom. Two loci reached genome-wide significance but were excluded from the significant results in Supplementary Table 2 due to heterogeneity between estimates from contributing studies and missingness between studies at chr. 6: 31057940–31380334 and chr. 7: 54671568–54759789; however, these regions are not excluded from the corresponding summary statistics in data release 5 (COVID-19 HGI (<https://www.covid19hg.org/results/r5/>) and GWAS Catalog (study code GCST011074)). For each of the lead variants reported in Supplementary Table 2, we aimed to find loci specific to susceptibility or severity by testing whether there was heterogeneity between the effect sizes associated with hospitalized COVID-19 (progression to severe disease) and reported SARS-CoV-2 infection. We used the Cochran's *Q* measure^{42,43}, calculated for each variant as the weighted sum of squared differences between the two analysis effect sizes and their meta-analysis effect with the weights being the inverse variance of the effect size. A significant *P* value of $P < 0.004$ ($(0.05/13)$ loci) for multiple tests) indicates that the effect sizes for a particular variant are significantly different in the two analyses (Supplementary Table 2). For the nine loci, in which the lead variant effect size was significantly higher for hospitalized COVID-19, we carried out the same test again but comparing effect sizes from hospitalized COVID-19 with critically ill COVID-19 (Supplementary Table 4). Furthermore, we carried out the same test comparing meta-analysed hospitalized COVID-19 (population as controls) and hospitalized COVID-19 (SARS-CoV-2-positive but non-hospitalized as controls) (Supplementary Table 4). For these pairs of phenotype comparisons, we generated new meta-analysis summary statistics to use; including only those studies that could contribute data to both phenotypes that were under comparison.

Principal component projection

To project every GWAS participant into the same principal component (PC) space, we used pre-computed PC loadings and reference allele frequencies. For reference, we used unrelated samples from the 1000 Genomes Project and the Human Genome Diversity Project and computed PC loadings and allele frequencies for the 117,221 single-nucleotide polymorphisms (SNPs) that (1) are available in every cohort; (2) have a minor allele frequency of >0.1% in the reference; and (3) are LD-pruned ($r^2 < 0.8$; 500-kb window). We then asked each cohort to project their samples using our automated script provided at <https://github.com/covid19-hg/>. It internally uses the PLINK2⁴⁴ --score function with the variance-standardize option and reference allele frequencies (--read-freq); so that each cohort-specific genotype/dosage matrix is mean-centred and variance-standardized with respect to reference allele frequencies, but not cohort-specific allele frequencies. We further normalized the projected PC scores by dividing the values by a square root of the number of variants used for projection to account for a subtle difference due to missing variants.

Gene prioritization

To prioritize candidate causal genes reported in full in Supplementary Table 2, we used various gene prioritization approaches using both locus-based and similarity-based methods. Because we only describe the in silico gene prioritization results without characterizing the actual functional activity in vitro or in vivo, we aimed to provide a systematic approach to nominate potential causal genes in a locus using the following criteria.

(1) The closest gene: a gene that is closest to a lead variant by distance to the gene body.

(2) Genes in the LD region: genes that overlap with a genomic range containing any variants in LD ($r^2 > 0.6$) with a lead variant. For LD computation, we retrieved LD matrices provided by gnomAD v.2.1.1⁴¹ for each population analysed in this study (except for admixed American, Middle Eastern and South Asian genetic ancestry populations, for whom data are not available). We then constructed a weighted-average LD matrix by per-population sample sizes in each meta-analysis, which we used as a LD reference.

(3) Genes with coding variants: genes with at least one loss-of-function or missense variant (annotated by VEP⁴⁵ v.95 with GENCODE v.29) that is in LD with a lead variant ($r^2 > 0.6$).

(4) eGenes: genes with at least one fine-mapped *cis*-eQTL variant (PIP > 0.1) that is in LD with a lead variant ($r^2 > 0.6$) (Supplementary Table 5). We retrieved fine-mapped variants from the GTEx v.8²⁰ (<https://www.finucanelab.org/>) and eQTL catalogue⁴⁶. In addition, we looked up significant associations in the Lung eQTL Consortium²¹ ($n = 1,103$) to further support our findings in lung with a larger sample size (Supplementary Table 7). We note that, in contrast to the GTEx or eQTL catalogue, we only looked at associations and did not fine-map our data to the Lung eQTL Consortium data.

(5) V2G: a gene with the highest overall V2G score based on Open Targets Genetics (OTG)²⁶. For each variant, the overall V2G score aggregates differentially weighted evidence of variant–gene associations from several data sources, including molecular *cis*-QTL data (for example, *cis*-protein QTLs from ref.⁴⁷, *cis*-eQTLs from GTEx v.7 and so on), interaction-based datasets (for example, promoter capture Hi-C), genomic distance and variant effect predictions (VEP) from Ensembl. A detailed description of the evidence sources and weights used is provided in the OTG documentation (<https://genetics-docs.opentargets.org/our-approach/data-pipeline>)²⁶.

Phenome-wide association study

To investigate the evidence of shared effects of 15 index variants for COVID-19 and previously reported phenotypes, we performed a phenome-wide association study. We considered phenotypes in OTG obtained from the GWAS catalogue (this included studies with and without full summary statistics, $n = 300$ and 14,013, respectively)⁴⁸ and from the UK Biobank. Summary statistics for UK Biobank traits were extracted from SAIGE³⁹ for binary outcomes ($n = 1,283$ traits) and Neale v.2 ($n = 2,139$ traits) for both binary and quantitative traits (<http://www.nealelab.is/uk-biobank/>) and FinnGen Freeze 4 cohort (https://www.finnngen.fi/en/access_results). We report PheWAS results for phenotypes for which the lead variants were in high LD ($r^2 > 0.8$) with the 13 genome-wide significant lead variants from our main COVID-19 meta-analysis (Supplementary Table 6). This conservative approach allowed spurious signals primarily driven by proximity rather than actual colocalization to be removed (see Methods).

To remove plausible spurious associations, we retrieved phenotypes for GWAS lead variants that were in LD ($r^2 > 0.8$) with COVID-19 index variants.

Heritability

LD score regression v.1.0.1⁴⁹ was used to estimate the SNP heritability of the phenotypes from the meta-analysis summary statistic files. As this

method depends on matching the LD structure of the analysis sample to a reference panel, the summary statistics of European ancestry only were used. Sample sizes were $n = 5,101$ critically ill cases of COVID-19 and $n = 1,383,241$ control participants, $n = 9,986$ hospitalized cases of COVID-19 and $n = 1,877,672$ control participants, and $n = 38,984$ cases and $n = 1,644,784$ control participants for the analysis of all cases—all including the 23andMe cohort. Pre-calculated LD scores from the 1000 Genomes European reference population were obtained online (<https://data.broadinstitute.org/alkesgroup/LDSCORE/>). Analyses were conducted using the standard program settings for variant filtering (removal of non-HapMap3 SNPs, the HLA region on chromosome 6, non-autosomal, $\chi^2 > 30$, minor allele frequency of <1%, or allele mismatch with reference). We additionally report SNP heritability estimates for the all-ancestries meta-analyses, calculated using European panel LD scores, in Supplementary Table 8.

Partitioned heritability

We used partitioned LD score regression⁵⁰ to partition COVID-19 SNP heritability in cell types in our summary statistics for European ancestry only. We ran the analysis using the baseline model LD scores calculated for European populations and regression weights that are available online (<https://github.com/bulik/ldsc>). We used the COVID-19 summary statistics for European ancestry only for the analysis.

Genome-wide association summary statistics

We obtained genome-wide association summary statistics for 43 complex-disease, neuropsychiatric, behavioural or biomarker phenotypes (Supplementary Table 10). These phenotypes were selected based on their putative relevance to COVID-19 susceptibility, severity or mortality, with 19 selected based on the Centers for Disease Control list of underlying medical conditions associated with COVID-19 severity⁵¹ or traits reported to be associated with increased risk of COVID-19 mortality by OpenSafely⁵². Summary statistics generated from GWAS using individuals of European ancestry were preferentially selected if available. These summary statistics were used in subsequent genetic correlation and Mendelian randomization analyses.

Genetic correlation

LD score regression⁵⁰ was also used to estimate the genetic correlations between our COVID-19 meta-analysis phenotypes reported using samples of only European ancestry, and between these and the curated set of 38 summary statistics. Genetic correlations were estimated using the same LD score regression settings as for heritability calculations. Differences between the observed genetic correlations of SARS-CoV-2 infection and COVID-19 severity were compared using a z-score method⁵³.

Mendelian randomization

Two-sample Mendelian randomization was used to evaluate the potential for causal association of the 38 traits on COVID-19 hospitalization, on COVID-19 severity and reported SARS-CoV-2 infection using samples of only European ancestry. Independent genome-wide significant SNPs robustly associated with the exposures of interest ($P < 5 \times 10^{-8}$) were selected as genetic instruments by performing LD clumping using PLINK⁴⁰. We used a strict r^2 threshold of 0.001, a 10-Mb clumping window, and the European reference panel from the 1000 Genomes Project⁵⁴ to discard SNPs in LD with another variant with a smaller P -value association. For genetic variants that were not present in the hospitalized COVID-19 analysis, PLINK was used to identify proxy variants that were in LD ($r^2 > 0.8$). Next, the exposure and outcome datasets were harmonized using the R package TwoSampleMR⁵⁵. Namely, we ensured that the effect of a variant on the exposure and outcome corresponded to the same allele, we inferred positive-strand alleles and dropped palindromes with ambiguous allele frequencies, as well as incompatible alleles. Supplementary Table 10 includes the harmonized datasets used in the analyses.

The global test from Mendelian randomization pleiotropy residual sum and outlier (MR-PRESSO)⁵⁶ software was used to investigate overall horizontal pleiotropy. In brief, the standard IVW meta-analytic framework was used to calculate the average causal effect by excluding each genetic variant used to instrument the analysis. A global statistic was calculated by summing the observed residual sum of squares, that is, the difference between the effect predicted by the IVW slope excluding the SNP, and the observed effect of the SNP on the outcome. Overall horizontal pleiotropy was subsequently analysed by comparing the observed residual sum of squares, with the residual sum of squares expected under the null hypothesis of no pleiotropy. The MR-PRESSO global test was shown to perform well when the outcome and exposure GWASs are not disjoint (although the power to detect horizontal pleiotropy is slightly reduced by complete sample overlap). We also used the regression intercept in MR-Egger⁵⁷ to evaluate potential bias due to directional pleiotropic effects. This additional check was used in Mendelian randomization analyses with an I_{GX}^2 index surpassing the recommended threshold ($I_{GX}^2 > 90\%$)⁵⁸. Contingent on the MR-PRESSO global test results we analysed the causal effect of each exposure on COVID-19 hospitalization by using a fixed-effect IVW meta-analysis as the primary analysis, or, if pleiotropy was present, the MR-PRESSO outlier-corrected test. The IVW approach estimates the causal effect by aggregating the single-SNP causal effects (obtained using the ratio of coefficients method—that is, the ratio of the effect of the SNP on the outcome over the effect of the SNP on the exposure) in a fixed-effects meta-analysis. The SNPs were assigned weights based on their inverse variance. The IVW method confers the greatest statistical power for estimating causal associations⁵⁹, but assumes that all variants are valid instruments and can produce biased estimates if the average pleiotropic effect differs from zero. Alternatively, when horizontal pleiotropy was present, we used the MR-PRESSO outlier-corrected method to correct the IVW test by removing outlier SNPs. We conducted further sensitivity analyses using alternative Mendelian randomization methods that provide consistent estimates of the causal effect even when some instrumental variables are invalid, at the cost of reduced statistical power including: (1) Weighted median estimator (WME); (2) weighted mode-based estimator (WMBE); and (3) MR-Egger regression. Robust causal estimates were defined as those that were significant at an FDR of 5% and either (1) showed no evidence of heterogeneity (MR-PRESSO global test $P > 0.05$) or horizontal pleiotropy (Egger intercept $P > 0.05$); or (2) in the presence of heterogeneity or horizontal pleiotropy, the WME-, WMBE-, MR-Egger- or MR-PRESSO-corrected estimates were significant ($P < 0.05$). All statistical analyses were conducted using R v.4.0.3. Mendelian randomization analysis was performed using the 'TwoSampleMR' v.0.5.5 package⁵⁵.

Website and data distribution

In anticipation of the need to coordinate many international partners around a single meta-analysis effort, we created the COVID-19 HGI website (<https://covid19hg.org>). We were able to centralize information, recruit partner studies, rapidly distribute summary statistics and present preliminary interpretations of the results to the public. Open meetings are held on a monthly basis to discuss future plans and new results; video recordings and supporting documents are shared (<https://covid19hg.org/meeting-archive>). This centralized resource provides a conceptual and technological framework for organizing global academic and industry groups around a shared goal. The website source code and additional technical details are available at <https://github.com/covid19-hg/covid19hg>.

To recruit new international partner studies, we developed a workflow in which new studies are registered and verified by a curation team (<https://covid19hg.org/register>). Users can explore the registered studies using a customized interface to find and contact studies with similar goals or approaches (<https://covid19hg.org/partners>). This helps to promote organic assembly around focused projects that are

adjacent to the centralized effort (<https://covid19hg.org/projects>). Visitors can query study information, including study design and research questions. Registered studies are visualized on a world map and are searchable by institutional affiliation, city and country.

To encourage data sharing and other forms of participation, we created a rolling acknowledgements page (<https://covid19hg.org/acknowledgements>) and directions on how to contribute data to the central meta-analysis effort (<https://covid19hg.org/data-sharing>). Upon the completion of each data freeze, we post summary statistics, plots and sample size breakdowns for each phenotype and contributing cohort (<https://covid19hg.org/results>). The results can be explored using an interactive web browser (<https://app.covid19hg.org>). Several computational research groups carry out follow-up analyses, which are made available for download (<https://covid19hg.org/in-silico>). To enhance scientific communication to the public, preliminary results are described in blog posts by the scientific communications team and shared on Twitter. The first post was translated to 30 languages with the help of 85 volunteer translators. We compile publications and preprints submitted by participating groups and summarize genome-wide significant findings from these publications (<https://covid19hg.org/publications>).

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this paper.

Data availability

Summary statistics generated by the COVID-19 HGI are available at <https://www.covid19hg.org/results/r5/> and are available in the GWAS Catalog (study code GCST011074). The analyses described here include the freeze-5 data. COVID-19 HGI continues to regularly release new data freezes. Summary statistics for non-European ancestry samples are not currently available due to the small individual sample sizes of these groups, but results for lead variants of 13 loci are reported in Supplementary Table 3. Individual level data can be requested directly from contributing studies, listed in Supplementary Table 1. We used publicly available data from GTEx (<https://gtexportal.org/home/>), the Neale lab (<http://www.nealelab.is/uk-biobank/>), Finucane lab (<https://www.finucanelab.org>), the FinnGen Freeze 4 cohort (https://www.finnngen.fi/en/access_results) and the eQTL catalogue release 3 (<http://www.ebi.ac.uk/eql/>).

Code availability

The code for summary statistics lift-over, the projection PCA pipeline including precomputed loadings and meta-analyses are available on GitHub (<https://github.com/covid19-hg/>) and the code for the Mendelian randomization and genetic correlation pipeline is available on GitHub at <https://github.com/marcoralab/MRCovid>.

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Author contributions Author contributions are provided within the author list.

Competing interests A full list of competing interests is supplied as Supplementary Table 13.

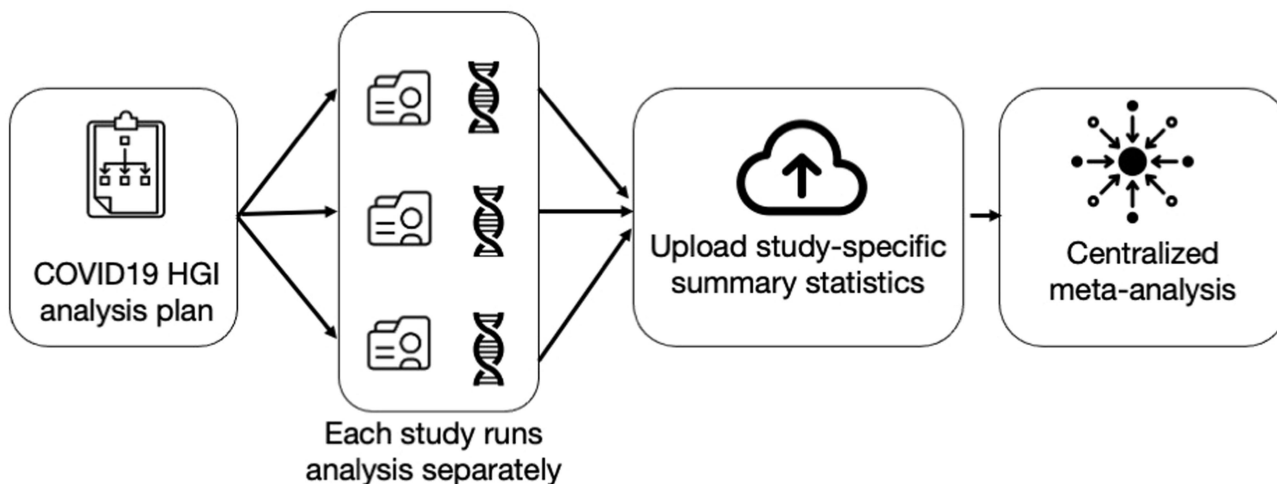
Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41586-021-03767-x>.

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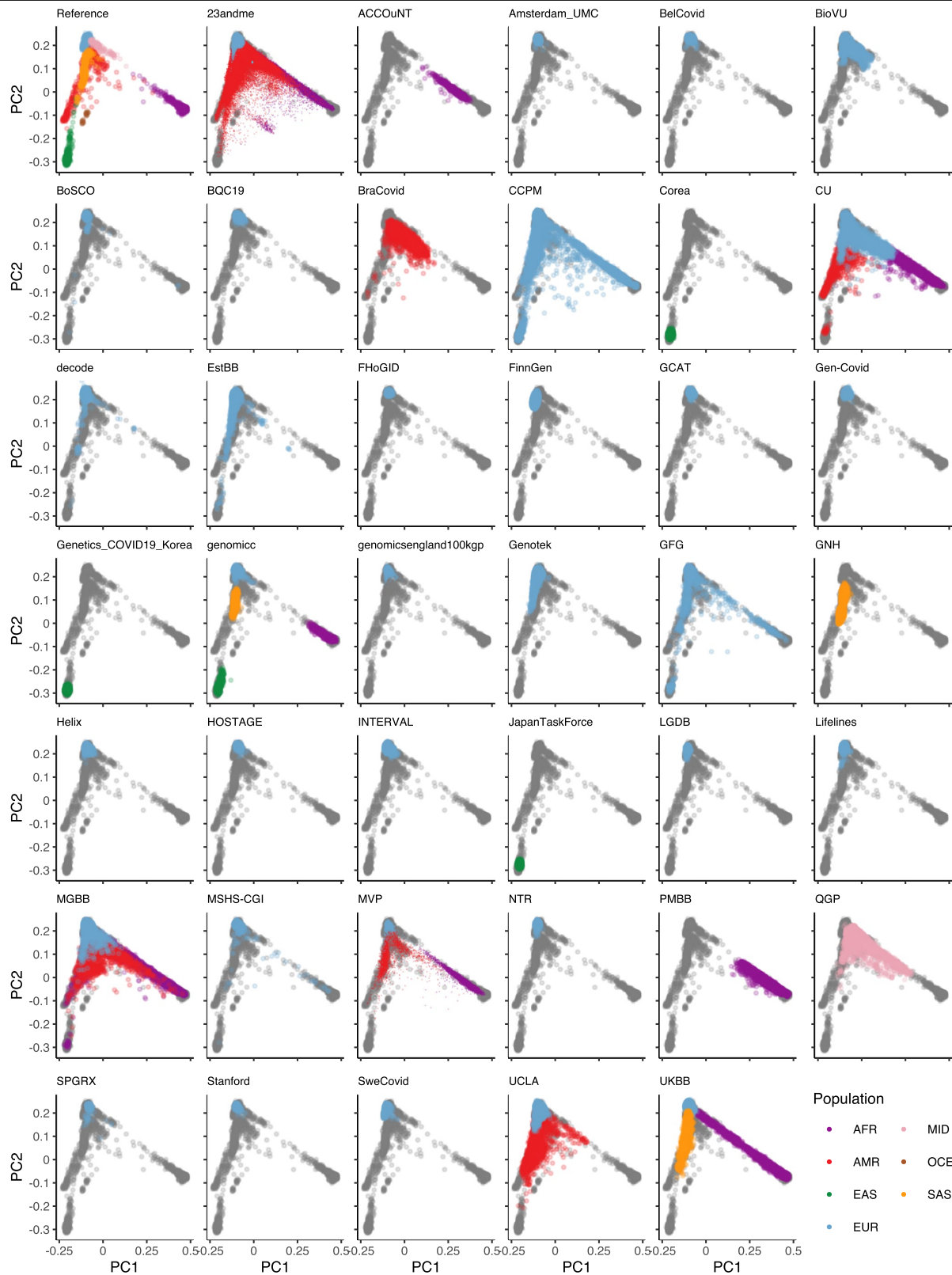
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	Critically ill COVID-19+	Hospitalized COVID-19+	Reported SARS-CoV-2 infection
Cases	Lab confirmed SARS-CoV-2 infection AND hospitalized for COVID-19 AND death OR respiratory support	Lab confirmed SARS-CoV-2 infection AND hospitalized for COVID-19	Lab confirmed SARS-CoV-2 infection OR Physician-reported COVID-19 OR self-reported COVID-19 via questionnaire
	N=6,179	N=13,641	N=49,562
Controls	Everybody that is not a case, e.g. population	Everybody that is not a case, e.g. population	Everybody that is not a case, e.g. population
	N=1,483,780	N=2,070,709	N=1,770,206
	↓ 6 loci	↓ 9 loci	↓ 7 loci
	13 unique loci		

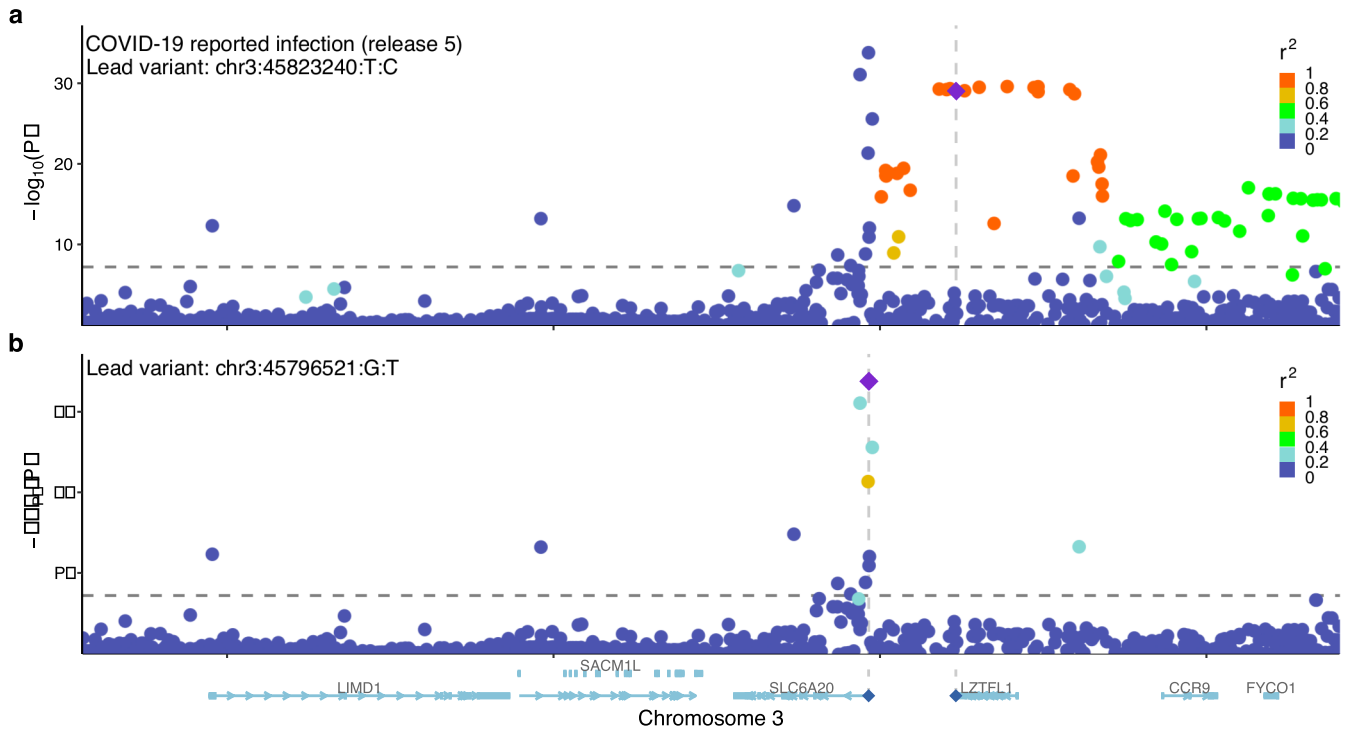
Extended Data Fig. 1 | Analytical summary of the COVID-19 HGI meta-analysis. Using the analytical plan set by the COVID-19 HGI, each individual study runs their analyses and uploads the results to the Initiative, who then runs the meta-analysis. There are three main analyses that each study can contribute summary statistics to: critically ill COVID-19, hospitalized COVID-19 and reported SARS-CoV-2 infection. The phenotypic criteria used to define cases are listed in the dark grey boxes, along with the numbers of cases

(*N*) included in the final all-ancestries meta-analysis. Controls were defined in the same way across all three analyses as everybody that is not a case—for example, population controls (light grey box). Sensitivity analyses—not reported in this extended data figure—also included mild and/or asymptomatic cases of COVID-19 as control individuals. Sample number (*N*) of control individuals differed between the analyses due to the difference in the number of studies contributing data to these.



Extended Data Fig. 2 | Projection of contributing studies samples into the same PC space. We asked participating studies to perform a PC projection using the 1000 Genomes Project and Human Genome Diversity Project as a reference, with a common set of variants. For each panel (except for the reference), coloured points correspond to contributed samples from each cohort, whereas grey points correspond to the reference samples from the 1000 Genomes Project. Colour represents a genetic population that each

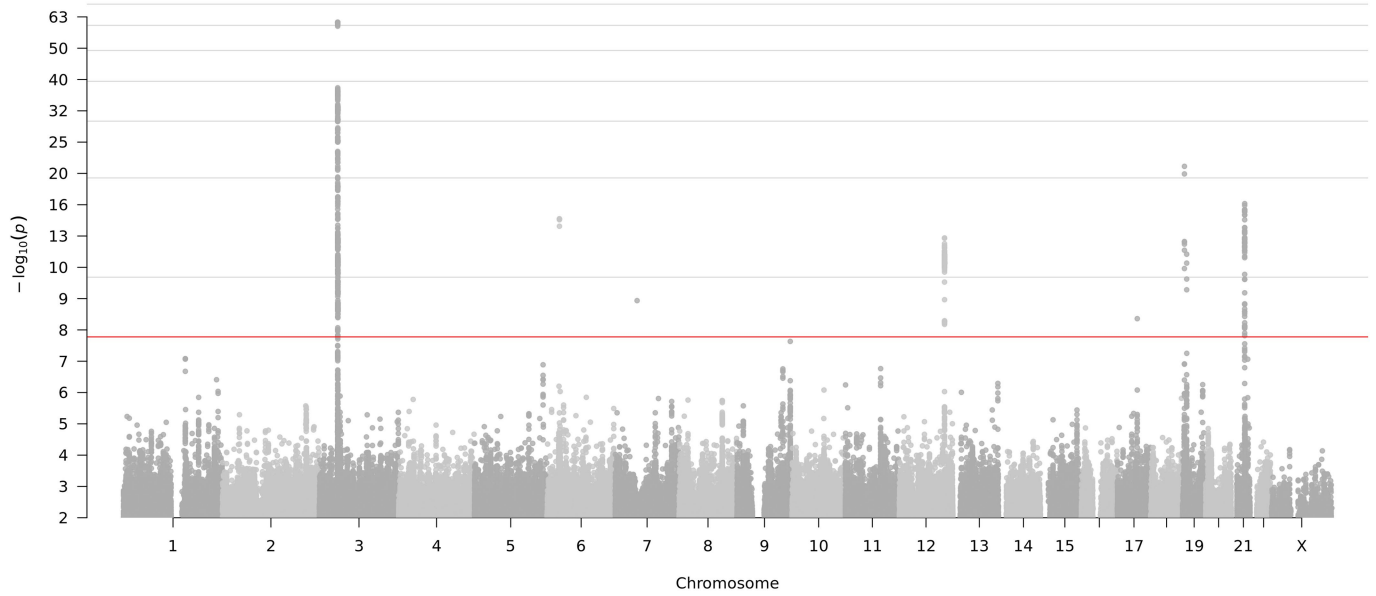
cohort specified. As 23andMe, Genomics England 100,000 Genomes Project (GenomicsEngland100kpg), and Million Veterans Program (MVP) only submitted PCA images, we overlaid their submitted transparent images using the same coordinates, instead of directly plotting them. Populations are defined as African (AFR), admixed American (AMR), East Asian (EAS), European (EUR), Middle Eastern (MID) and South Asian (SAS), Oceanian (OCE).



Extended Data Fig. 3 | Locus-zoom plots of the 3p21.31 region for reported SARS-CoV-2 infection. a, A standard plot without exclusion. Here, the severity lead variant rs10490770 (chr. 3: 45823240T:C) is shown as a lead variant.

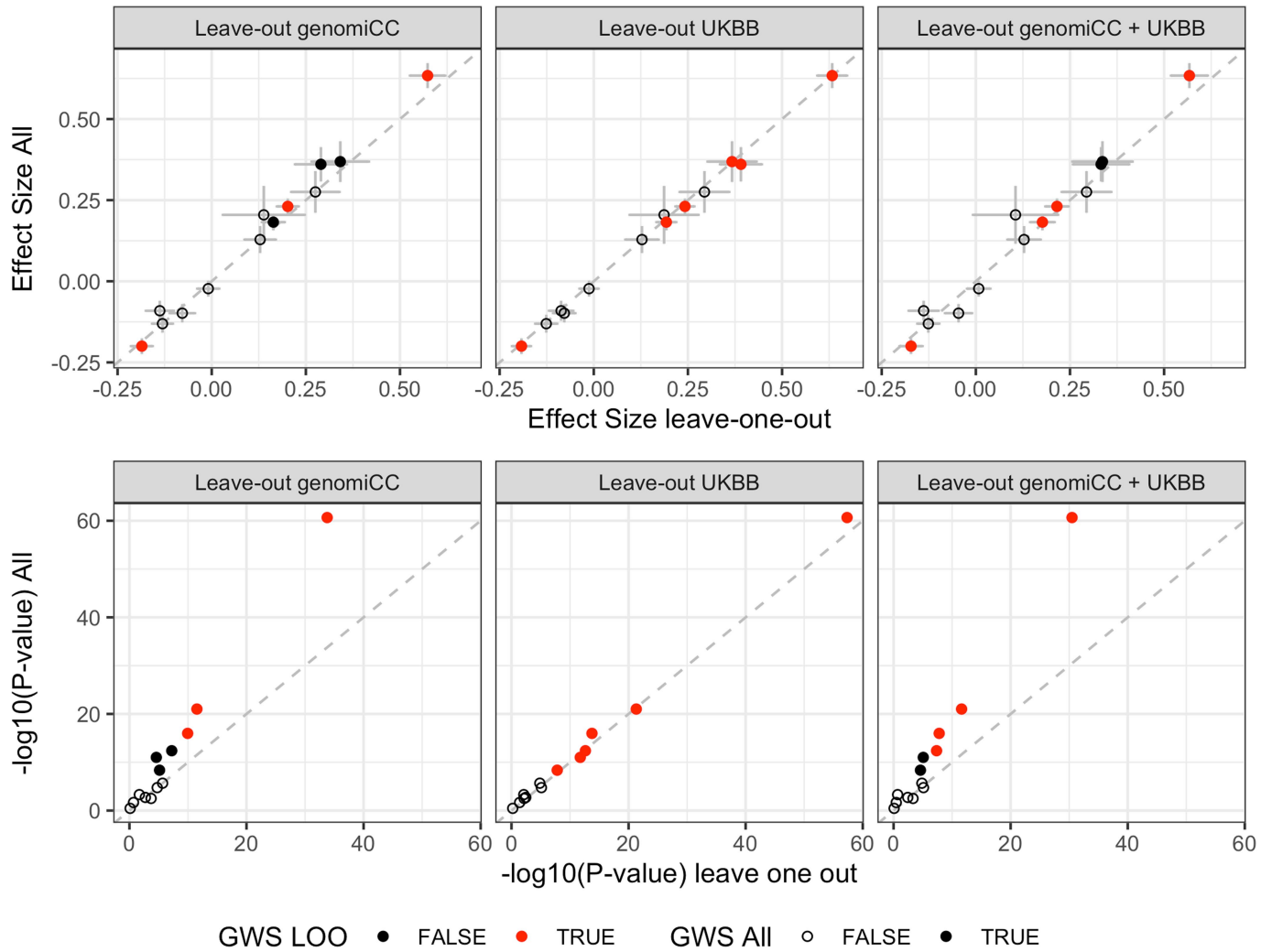
b, Additional independent susceptibility signal(s) after excluding variants with $r^2 > 0.05$ with rs10490770. The susceptibility lead variant rs2271616 (chr. 3: 45796521G:T) is highlighted.

Critically ill COVID-19+ (N.cases=6,179,N.controls=1,483,780)



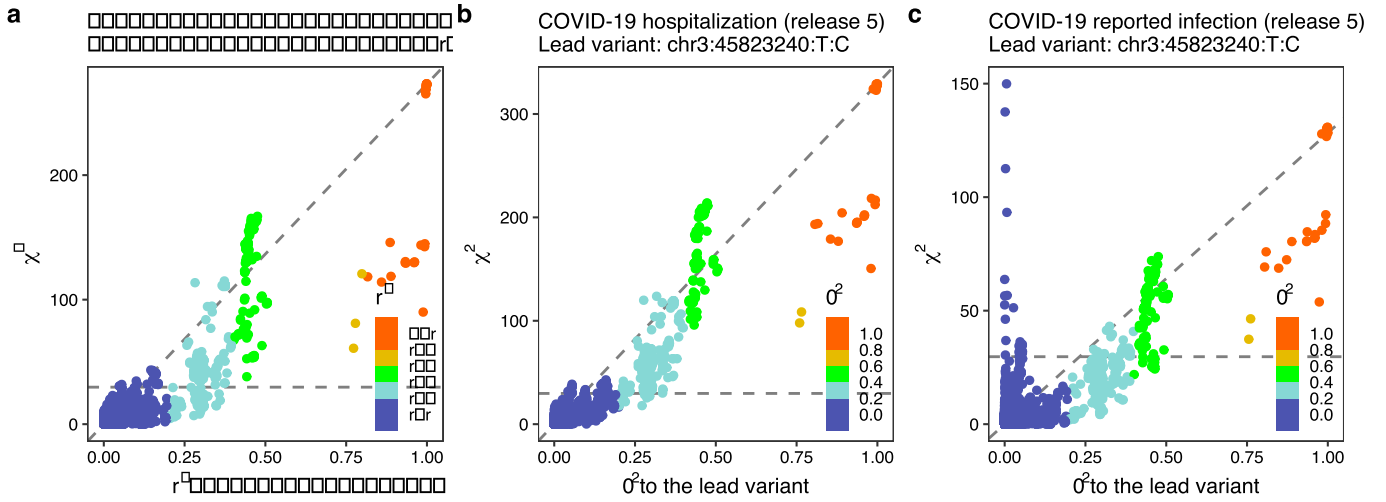
Extended Data Fig. 4 | Genome-wide meta-analysis association results for critical illness due to COVID-19. The locus on chromosome 6 is the HLA locus, which was removed from the list of reported loci in Supplementary Table 2 due to the high heterogeneity in effect size estimated between studies included in

the analysis. The locus on chromosome 7 was also not reported in Supplementary Table 2 due to missingness across studies—that is, the high number of studies in the meta-analysis that did not report summary statistics for this region. There are two association peaks on chromosome 19.



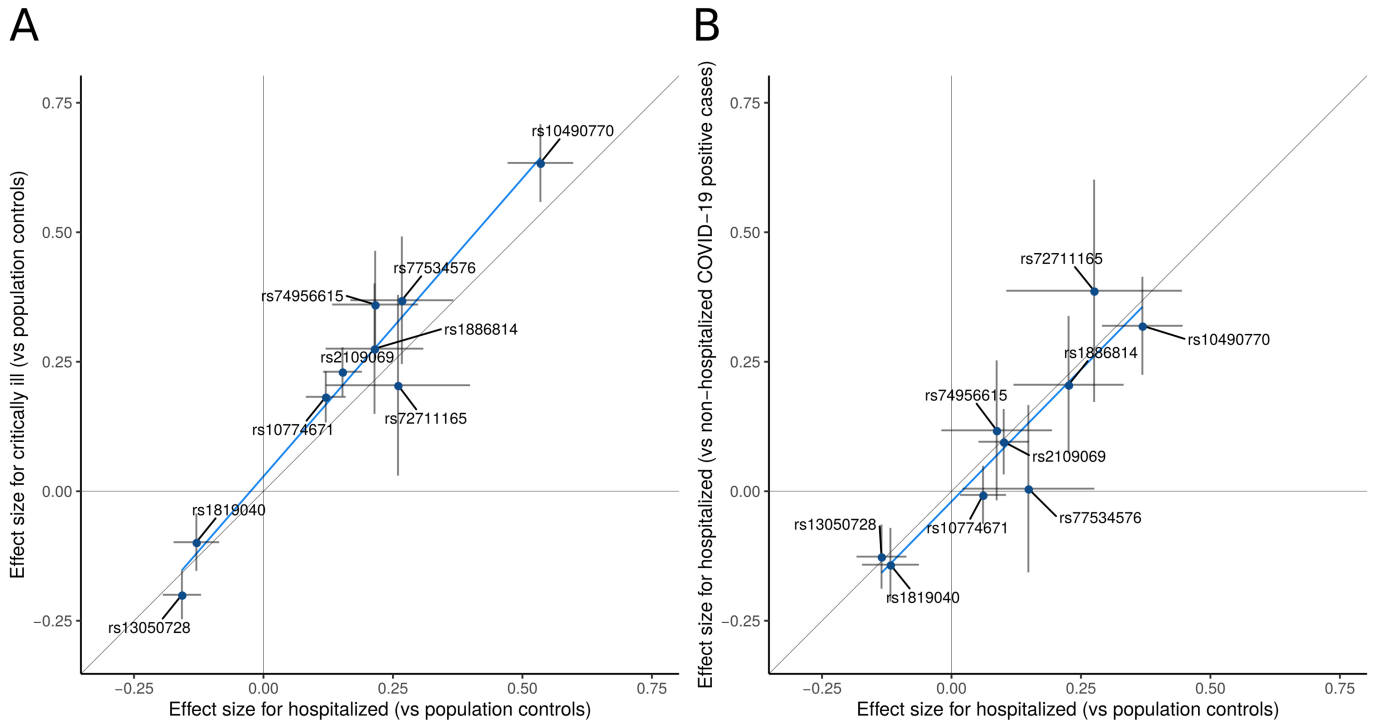
Extended Data Fig. 5 | Sensitivity analyses for overlapping controls in genomICC and UK Biobank. Comparison of the beta effect sizes (top) and unadjusted *P* values (bottom) of the 13 lead variants, using data from the COVID-19 critical illness meta-analysis in all the cohorts to leaving out genomICC (cases, *n* = 4,354; controls, *n* = 1,474,655; total, *n* = 1,479,009), leaving out the UK Biobank (UKBB; cases, *n* = 5,870; controls, *n* = 1,155,203; total, *n* = 1,161,073) and leaving out both genomICC and UK Biobank (cases, *n* = 4,045; controls, *n* = 1,146,078; total, *n* = 1,150,123) (from left to right, respectively). Top, dots and grey bars represent the beta effect size estimates ± standard

error from the corresponding GWAS meta-analysis. Bottom, dots represent two-sided *P* values from the corresponding GWAS meta-analysis. Filled dots indicate variants that showed genome-wide significance in the full meta-analysis of critical illness due to COVID-19, and empty dots represent variants that were not significant for critical illness but were significant for either hospitalization due to COVID-19 or reported SARS-CoV-2 infection. Red dots represent variants that showed genome-wide significance in the leave-one-out analysis for genomICC, UK Biobank or genomICC and UK Biobank.



Extended Data Fig. 6 | Comparison of χ^2 statistics and r^2 values to the lead variant in the 3p21.31 region. a-c, Data are shown for critical illness (a), hospitalization (b) and reported SARS-CoV-2 infection (c). The left blue peak in

c, which is uncorrelated with the lead variants in the region, indicates that there are independent signals.



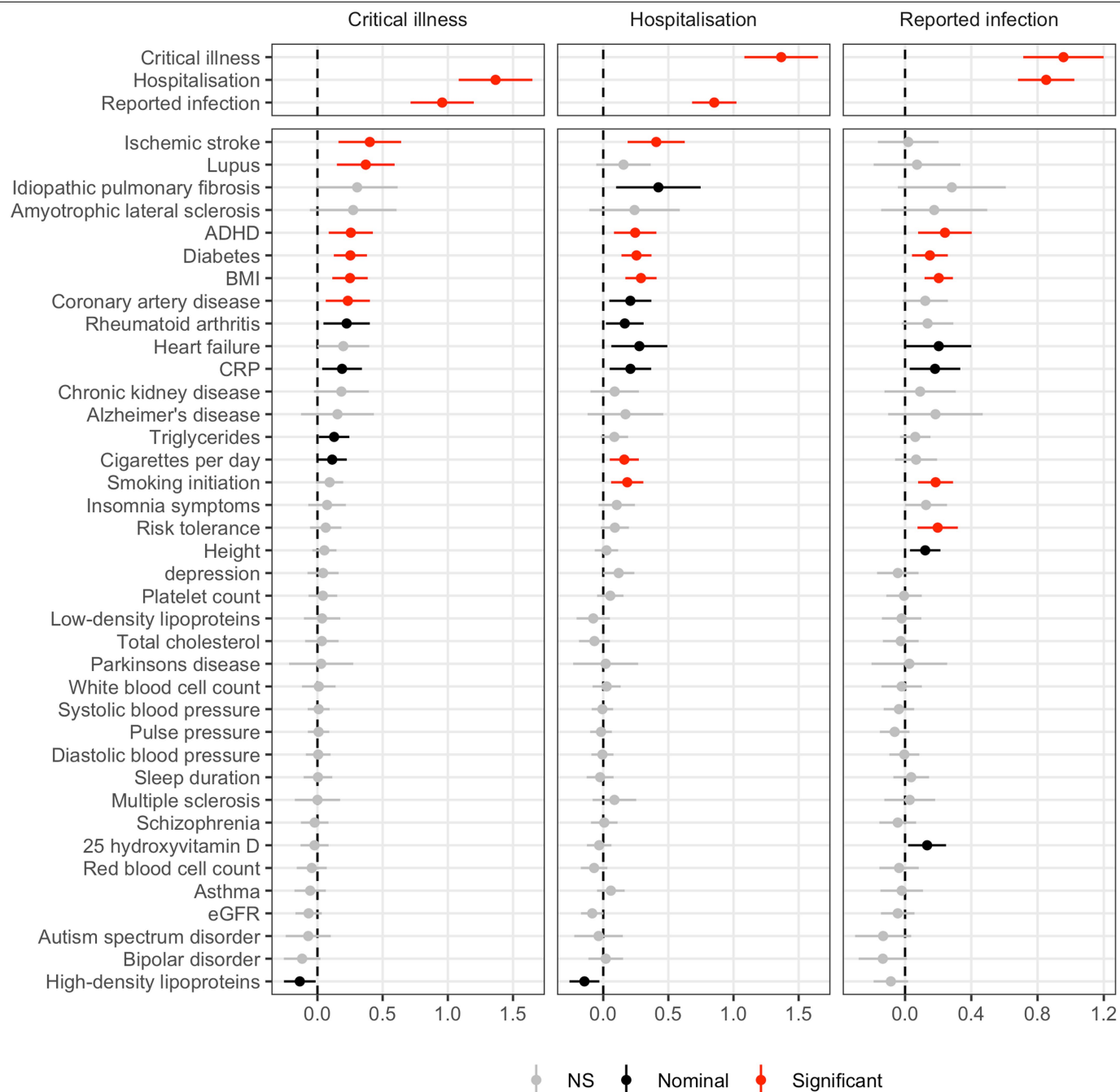
Extended Data Fig. 7 | Comparison of the effect sizes of lead variants between pairs of COVID-19 meta-analyses. Comparison of effect sizes for the nine variants associated with severity of COVID-19 disease. **a**, Comparing hospitalized cases of COVID-19 versus population controls ($n = 10,428$ cases and $n = 1,483,270$ controls) and critically ill cases of COVID-19 versus population controls ($n = 6,179$ cases and $n = 1,483,780$ controls). **b**, Hospitalized cases of COVID-19 versus population controls ($n = 5,806$ cases and $n = 1,144,263$ controls) and hospitalized cases of COVID-19 versus non-hospitalized cases of

COVID-19 ($n = 5,773$ cases and $n = 15,497$ controls). Sample sizes for hospitalized cases of COVID-19 versus population controls differ between **a** and **b** due to differences in the sampling of studies selected for the analysis. This selection included all studies that were able to contribute data to the respective analyses that the data were compared to (shown on the y axis) in each panel. Dots represent the effect size beta estimates, bars represent the 95% confidence interval of the estimates. Effect size estimates and P values for heterogeneity tests (Cochran's Q , two-tailed test) are reported in Supplementary Table 3.



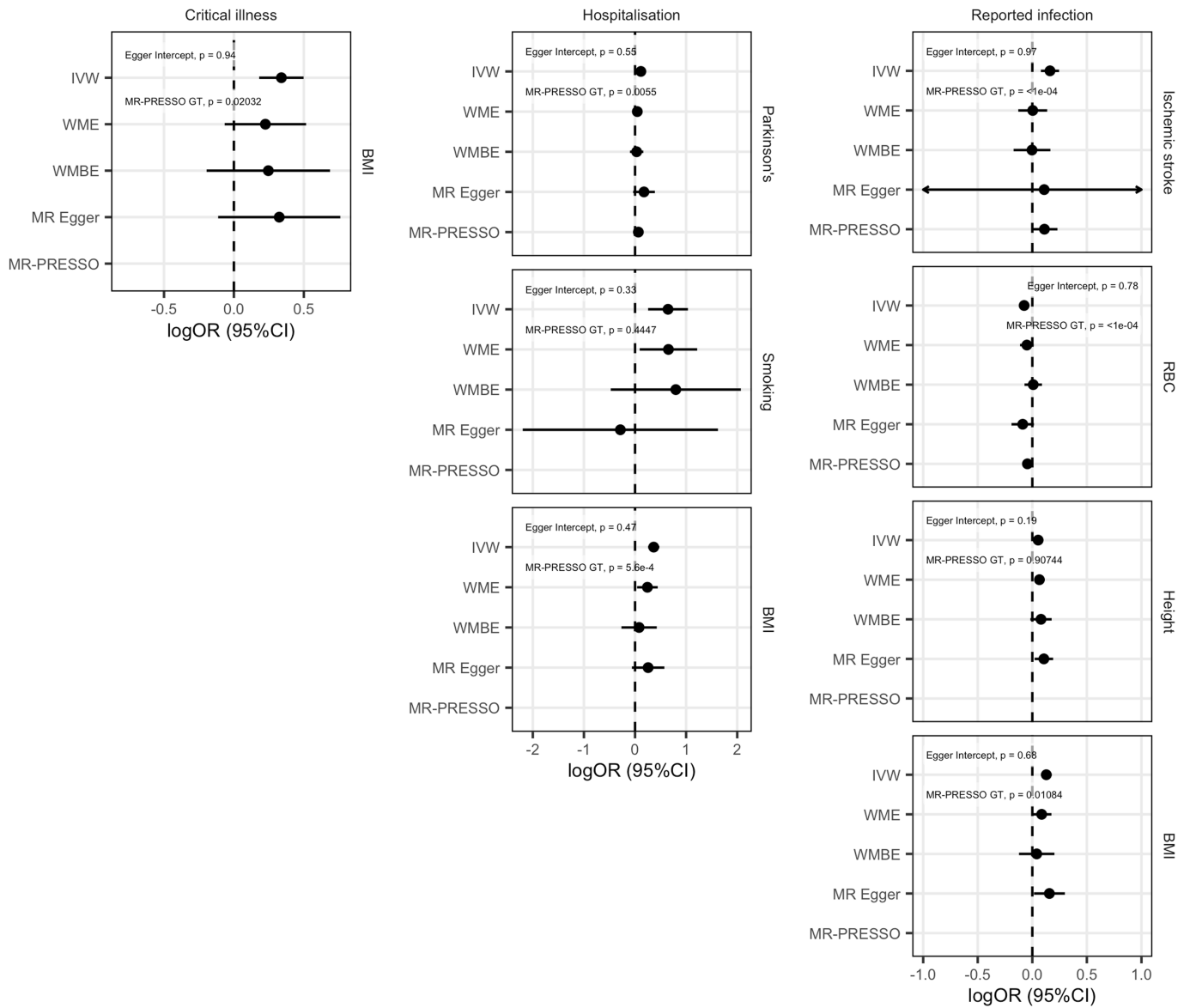
Extended Data Fig. 8 | PheWAS for genome-wide significant lead variants.

Selected phenotypes associated with genome-wide significant COVID-19 variants (see Supplementary Table 6 for a complete list). We report those associations for which a lead variant from a previous GWAS result was in high LD ($r^2 > 0.8$) with the index COVID-19 variants. The colour represents the z-scores of correlated risk increasing alleles for the trait. The total number of associations for each COVID-19 variant is highlighted in the grey box.



Extended Data Fig. 9 | Genetic correlation with COVID-19 phenotypes. Each column shows the genetic correlation results for the three COVID-19 phenotypes (European-ancestry analyses only): critical illness, hospitalization and reported SARS-CoV-2 infection. The traits that the genetic correlation is run against are listed on the left. Significant correlations (FDR < 0.05) are

shown with their 95% confidence intervals in red, nominally significant correlations ($P < 0.05$) are in black and non-significant correlations are in grey. Two-sided P values were calculated using LDSC for genetic correlations and exact estimates, unadjusted standard errors and two-sided P values are available in Supplementary Table 11.



Extended Data Fig. 10 | Mendelian randomization sensitivity analyses. Genetic correlations and Forest plots displaying the causal estimates for each of the sensitivity analyses used in the Mendelian randomization analysis for

trait pairs that were significant at an FDR of 5%. Two-sided P values were estimated using IVW, WME, WMBE and MR-PRESSO analyses. RBC, red blood cell count.

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Software and code

Policy information about [availability of computer code](#)

Data collection

Data analysis Each individual study that contributed genetic-phenotype association summary statistics to the consortium carried out their association analyses independently of the consortium (study-specific information outlined in Supplementary Table 1). However, the consortium did release phenotyping and analysis guidelines as a recommendation (<https://www.covid19hg.org/>). For quality control of genotype data we recommended using the Ricopili pipeline (PMID: 31393554). For genotype phasing and imputation we recommended the TopMed Imputation Server (PMID: 27571263) or Michigan Imputation Server (PMID: 27571263). For genome-wide association study (GWAS), we recommended SAIGE (PMID: 30104761), but some studies used PLINK (PMID: 17701901). Each study then submitted their GWAS summary statistics to the consortium for meta-analysis.

LD score regression v 1.0.1 [PMID: 25642630] was used for heritability and partitioned heritability analyses. Variants for Mendelian randomization instruments were selected using PLINK version 1.90b6.18 (PMID: 17701901). Exposure and outcome datasets were harmonized, and MR statistical analysis conducted using R version 4.0.3. with the R-package TwoSampleMR version 0.5.5 (PMID: 29846171) (which included Fixed-effects IVW analysis (PMID: 24114802), weighted median estimator (WME) (PMID: 27061298), weighted mode based estimator (WMBE) and MR Egger regression (PMID: 26050253)) and additionally MR-PRESSO version 1.0 (PMID: 29686387).

Code availability statement: The code for summary statistics liftover, projection PCA pipeline including precomputed loadings and meta-analysis are available at <https://github.com/covid19-hg/> and the code for Mendelian randomization and genetic correlation pipeline at <https://github.com/marcoralab/MRcovid>.

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Data availability statement:

Summary statistics generated by COVID-19 HGI are available at <https://www.covid19hg.org/results/r5/> and are available on GWAS Catalog (study code GCST011074). The analyses described here utilize the freeze 5 data. COVID-19 HGI continues to regularly release new data freezes. Summary statistics for non-European ancestry samples are not currently available due to the small individual sample sizes of these groups, but results for 13 loci lead variants are reported in Supplementary Table 3. Individual level data can be requested directly from contributing studies, listed in Supplementary Table 1. We used publicly available data from GTEx (<https://gtexportal.org/home/>), the Neale lab (<http://www.nealelab.is/uk-biobank/>), Finucane lab (<https://www.finucanelab.org>), FinnGen Freeze 4 cohort (https://www.finnngen.fi/en/access_results), and eQTL catalogue release 3 (<http://www.ebi.ac.uk/eqtl/>).

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Life sciences study design

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Sample size	The consortium meta-analysed genome-wide association study (GWAS) summary statistics from any individual study that had included a minimum of n=50 cases and n=50 controls in their analysis. The cutoff at n=50 cases and n=50 controls was aimed at reducing noise to the meta-analysis, but also to be inclusive of studies that had not yet accumulated large numbers of COVID-19 patient data. No statistical calculation for adequate sample size was performed, but the results identifying multiple genomic regions at genome-wide significance threshold indicates adequate power for genetic discovery.
Data exclusions	Individual level phenotype and genotype data exclusions were performed by each individual study, following the consortium analysis plan recommendations (www.covid19hg.org). Possible reasons for sample exclusion included removing genetic ancestry outliers within a study (using principal components analysis), poor quality of genetic data or lack of phenotypic data for a sample. The consortium manually examined GWAS summary statistics data submitted by each study (for each submitted analysis separately), including sample size used for analysis, allele frequency check against gnomad reference panel, and distribution of test statistics. After meta-analysis, the results were checked for heterogeneity variant effects between contributing studies, and Table 1 excludes two genome-wide significant loci that were deemed to have extremely heterogeneous effects, but these variants are reported in the released consortium summary statistics (with heterogeneity test values).
Replication	No replication was performed. The consortium meta-analysed GWAS summary statistics, bringing together as many studies as possible to achieve the largest possible sample size and statistical power for association. This meant that the consortium included most large studies of COVID-19 host genetics that have been performed to date, so it was not possible to perform replication analyses in external cohorts. Therefore we performed manual checks on each study contributing summary statistics before entering them into the meta-analysis. In addition, after meta-analysis, we performed a check for heterogeneity between variant association estimates across studies contributing data. This allowed us to better understand whether the variant effects differed much between individual studies.
Randomization	No randomization was performed because there was no allocation of samples to experimental groups.
Blinding	Blinding was not relevant to the study. The case status and severity of symptoms was evaluated for each sample by investigators from each study respectively. The consortium recommended using covariates to control for confounding: age + age ² + sex + age*sex + 20 principal components (obtained using genetic data) + study specific covariates (if any). The consortium meta-analysed summary statistics from these case/control studies, not individual level data. Details of which variables each study used and how the calculated PCs for their analysis are available in Supplementary Table 1.

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<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
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<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input type="checkbox"/>	<input checked="" type="checkbox"/> Human research participants
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<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

Methods

n/a	Included in the study
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<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Human research participants

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Population characteristics

Summary statistics from 46 independent studies were included in consortium meta-analyses. Mean age of cases across studies was 55.3 years. The effective sample size for genetic ancestry populations was: n=11,598 Middle Eastern; n=28,918 South Asian; 43,332 East Asian; 48,714 African; 70,902 Ad-mixed American; 738,538 European. Population characteristics regarding age, sex and exact case and control sample numbers for each contributing study are given in Supplementary Table 1.

Recruitment

The consortium pre-defined phenotype criteria for cases and controls, but the specific recruitment was carried out independently by each contributing study. COVID-19 disease status (critical illness, hospitalization status) was assessed following the Diagnosis and Treatment Protocol for Novel Coronavirus Pneumonia (PMID: 32358325). The critically ill COVID-19 group included patients who were hospitalized due to symptoms associated with laboratory-confirmed SARS-CoV-2 infection and who required respiratory support or whose cause of death was associated with COVID-19. The hospitalized COVID-19 group included patients who were hospitalized due to symptoms associated with laboratory-confirmed SARS-CoV-2 infection. The reported infection cases group included individuals with laboratory-confirmed SARS-CoV-2 infection or electronic health record, ICD coding or clinically confirmed COVID-19, or self-reported COVID-19 (e.g. by questionnaire), with or without symptoms of any severity. Genetic ancestry-matched controls for the three case definitions were sourced from population-based cohorts, including individuals whose exposure status to SARS-CoV-2 was either unknown or infection- negative for questionnaire/electronic health record based cohorts.

Ethics oversight

Ethical statements for each contributing study are given in Supplementary Table 1.

Note that full information on the approval of the study protocol must also be provided in the manuscript.