

Journal Pre-proof



Monitoring and Management of Hemolytic Disease of the Fetus and Newborn Based on an International Expert Delphi Consensus

Hiba J. Mustafa, MD, Enaja V. Sambatur, MD, Alireza A. Shamshirsaz, MD, Sonia Johnson, MD, Kenneth J. Moise, Jr., MD, Ahmet A. Baschat, MD, E.J.T. (Joanne) Verweij, MD, Ali Javinani, MD, Mark D. Kilby, MD, DSc, Enrico Lopriore, MD, Rebecca Rose, MD, Roland Devlieger, MD, Saul Snowise, MD, Ulrich J. Sachs, MD, Asma Khalil, MD, MSc, On behalf of the HDFN Delphi Working Group

PII: S0002-9378(24)01130-X

DOI: <https://doi.org/10.1016/j.ajog.2024.11.003>

Reference: YMOB 15928

To appear in: *American Journal of Obstetrics and Gynecology*

Received Date: 8 September 2024

Revised Date: 6 November 2024

Accepted Date: 7 November 2024

Please cite this article as: Mustafa HJ, Sambatur EV, Shamshirsaz AA, Johnson S, Moise Jr KJ, Baschat AA, Verweij EJT(J), Javinani A, Kilby MD, Lopriore E, Rose R, Devlieger R, Snowise S, Sachs UJ, Khalil A, On behalf of the HDFN Delphi Working Group, Monitoring and Management of Hemolytic Disease of the Fetus and Newborn Based on an International Expert Delphi Consensus, *American Journal of Obstetrics and Gynecology* (2024), doi: <https://doi.org/10.1016/j.ajog.2024.11.003>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2024 Elsevier Inc. All rights are reserved, including those for text and data mining, AI training, and similar technologies.

1 **Monitoring and Management of Hemolytic Disease of the Fetus and Newborn Based on an**
2 **International Expert Delphi Consensus**

3 **Hiba J. Mustafa, MD¹; Enaja V. Sambatur, MD²; Alireza A. Shamshirsaz, MD²; Sonia**
4 **Johnson, MD³; Kenneth J Moise Jr, MD⁴; Ahmet A. Baschat, MD⁵; E.J.T. (Joanne)**
5 **Verweij, MD⁶; Ali Javinani, MD²; Mark D. Kilby, MD, DSc⁷; Enrico Lopriore, MD⁸;**
6 **Rebecca Rose, MD⁹; Roland Devlieger, MD^{10,11}; Saul Snowise, MD¹²; Ulrich J. Sachs,**
7 **MD¹³; Asma Khalil, MD, MSc¹⁴**

8 **· .On behalf of the HDFN Delphi Working Group†**

9 ¹The Fetal Center at Riley Children's and Indiana University Health, Division of Maternal-Fetal
10 Medicine, Indiana University School of Medicine, Riley Children's Hospital, Indianapolis, IN, USA

11 ²Fetal Care and Surgery Center, Division of Maternal-Fetal Medicine and Surgery, Boston
12 Children's Hospital, Harvard Medical School, Boston, MA, USA

13 ³Fetal Medicine Unit, Liverpool Women's Hospital, University of Liverpool, UK

14 ⁴Dell Medical School – University of Texas at Austin and the Comprehensive Fetal Care Center,
15 Dell Children's Hospital, Austin, TX, USA

16 ⁵The Johns Hopkins Center for Fetal Therapy, Johns Hopkins University, Baltimore, MD, USA

17 ⁶Department of Obstetrics, Division of Fetal Therapy, Leiden University Medical Center,
18 Leiden, The Netherlands

19 ⁷Fetal Medicine Centre, Birmingham Women's & Children's Foundation Trust, Birmingham,
20 UK, and University of Birmingham, UK.(ORCID: [0000-0001-9987-4223](https://orcid.org/0000-0001-9987-4223))

21 ⁸Neonatology and Fetal Medicine, Leiden University Medical Center, Leiden, The Netherlands

22 ⁹Division of Neonatology, Indiana University School of Medicine, Riley Children's Hospital, IN,
23 USA

24 ¹⁰Maternal-Fetal Medicine, University Hospital Leuven, Belgium

25 ¹¹Mother and Child Unit, Department of Development and Regeneration, KU Leuven, Belgium

26 ¹²Midwest Fetal Care Center, Children's Minnesota, MN, USA

27 ¹³Immunology and Transfusion Medicine, Justus-Liebig-University, Germany

28 ¹⁴Fetal Medicine Unit, St George's Hospital, St George's University of London, London, UK

29

30 †Collaborators are listed at the end of the article.

31

32 **The authors report no conflicts of interest.**

33 **No external funding has been received for this study.**

34

35 **Corresponding author:**

36 **Hiba J. Mustafa, MD**

37 Maternal-Fetal Medicine, Indiana University School of Medicine, Riley Children's Hospital,

38 Indianapolis, IN

39 hmustafa@iu.edu

40

41 **Abstract word count: 288**

42 **Full-text word count: 2980**

43 **Table numbers: 6**

44 **Figure numbers: 1**

45

46

Condensation Page

47 **Tweetable statement:** The Delphi method facilitated the development of a consensus-based
48 clinical workflow informing the clinical management of pregnancies at risk or affected by red
49 cell alloimmunization.

50 **Short title:** Haemolytic Disease of the Fetus and Newborn Delphi

51

AJOG at a Glance

52 **Why was this study conducted?** To reach a Delphi-generated international expert consensus on
53 the monitoring and the management of hemolytic disease of the fetus and newborn.

54 **What are the key findings?**

- 55 ● The expert panel agreed on using cell-free DNA to determine fetal genotype in
56 pregnancies with red blood cell alloimmunization
- 57 ● Antibody titers of ≥ 16 are considered a critical threshold requiring fetal monitoring via
58 ultrasound in non-anti-Kell alloimmunized pregnancies.
- 59 ● The earliest middle cerebral artery (MCA) Doppler ultrasound assessment is to be started
60 at 16 weeks gestation, to be performed weekly, and to be continued until delivery.
- 61 ● Intravenous immunoglobulin infusions are to be considered in pregnancies with prior
62 fetal or neonatal death due to hemolytic disease or a history of intrauterine transfusion
63 before 24 weeks in a previous pregnancy. Consensus related to indications, gestational
64 age at initiation, MCA Dopplers, and dosage of infusions was reached.
- 65 ● The timing of the second and third intrauterine transfusion (IUT) can be determined by a
66 combination of weekly MCA Dopplers and the calculated rate of hemoglobin decline,

67 which is approximately 0.9 g/dL per day in 10-14 days after the first transfusion and 0.6
68 g/dL per day in 2-3 weeks after the second transfusion. 78.5% of participating experts
69 perform IUT until a gestational age of 35^{0/7} to 35^{6/7} weeks.

- 70 ● Delivery timing in alloimmunized pregnancies in which no IUT was indicated to be
71 between 37^{0/7} - 38^{6/7} weeks. However, if IUT was performed then timing to be two to
72 three weeks following the last transfusion.
- 73 ● Regarding postnatal management, the thresholds for phototherapy and exchange
74 transfusion are to be determined by the American Academy of Pediatrics (AAP) 2022
75 guidelines. Anemia labs should be investigated prior to discharge in all neonates of
76 pregnancies complicated with alloimmunization. If IUT was required, anemia labs should
77 be repeated in one week following discharge if the initial labs were normal.
- 78 ● The panel agreed that the hemoglobin cut-off level to consider transfusion following
79 hospital discharge is 7 g/dL and the newborns need to be monitored frequently until 2-3
80 months of age.

81

82 **What does this study add to what is already known?**

83 The findings of this Delphi can be used to create a standardized approach in the
84 monitoring and management of pregnancies and newborns affected by maternal
85 alloimmunization, particularly related to aspects where clinical and research knowledge gaps
86 exist.

87

ABSTRACT

88 The study aimed to develop structured, expert-based clinical guidance on the prenatal and
89 postnatal management of hemolytic disease of the fetus and newborn. A Delphi procedure was
90 conducted among an international panel of experts in fetal medicine, neonatology, and
91 hematology. Experts were selected based on their expertise, relevant publications, and
92 affiliations. The domains were (i) prenatal workup, (ii) prenatal monitoring and management,
93 (iii) intrauterine transfusion, (iv) delivery, and (v) postnatal management. The pre-defined cut-off
94 for consensus was $\geq 70\%$ agreement. One hundred-seven experts representing 25 countries across
95 six continents completed the first round, and 100 (93.5%) completed the subsequent rounds.

96 75.3% agreed on using cfDNA to determine fetal antigen status, particularly for RhD, Kell, and
97 Rhc antigens. The critical titer, requiring fetal monitoring via ultrasound, is considered when the
98 threshold of ≥ 16 is for non-Kell antigens. 70.0% agreed on the use of maternal IVIg in
99 pregnancies with prior intrauterine transfusion (IUT) < 24 weeks or fetal/neonatal death due to
100 HDFN. The minimum GA for IUT is 16 to 18 weeks, and the maximum is $35^{0/7}$ to $35^{6/7}$ weeks.

101 Postnatal management consensus was reached for the following: anemia labs should be
102 investigated in the affected neonates before hospital discharge (92.0% agreement), and if they
103 received IUT, the labs should be repeated within one week of discharge (84.0% agreement).

104 96.0% agreed that exchange transfusions should be centralized in hospitals with sufficient
105 exposure and experience, and 92.0% agreed that the hemoglobin cut-off level to consider
106 transfusion following hospital discharge is 7g/dL, and the newborns need to be monitored until
107 2-3 months of age (96.0% agreement).

108 **Keywords:** Fetal; Erythroblastosis; Pregnancy; Delphi; Consensus, IVIg, IUT, anemia,
109 hemolytic disease, cordocentesis, PUBS

110

111 INTRODUCTION

112 Red blood cell (RBC) alloimmunization has historically been an unknown and tragic cause
113 of perinatal death. Still, its trajectory has significantly changed with the worldwide implementation
114 of prenatal screening and the administration of Rho(D) Immune Globulin (RhIG)¹. However, the
115 RhD antigen is not the only RBC antigen that can cause alloimmunization; numerous other
116 antigens can cause hemolytic disease of the fetus and newborn (HDFN)^{2,3}. The mortality and
117 morbidity of HDFN in the post-RhIG era are heavily determined by prenatal screening,
118 monitoring, and management, which are shown to be directly related to the socioeconomic status
119 of the countries⁴.

120 The perinatal mortality of HDFN has been reported to be 20-25%, meaning that one out of
121 five pregnancies diagnosed with RBC alloimmunization can still lead to the perinatal death of the
122 fetus or newborn⁴. Moreover, it is estimated that nearly one-third of the patients are diagnosed
123 with hydrops at the time of diagnosis⁴. This significantly highlights the importance of screening in
124 at-risk populations, the optimal prenatal management of pregnancies diagnosed with fetal anemia,
125 and the meticulous postnatal care of affected newborns.

126 The prenatal screening, diagnosis, and monitoring of patients with HDFN have been
127 addressed in several clinical guidelines from different scientific societies^{5,6}. These include
128 strategies for implementing and interpreting screening antibodies, fetal genotyping, middle
129 cerebral artery (MCA) Doppler evaluation, and fetal blood sampling. Regarding the treatment of
130 patients with fetal anemia, intrauterine transfusion (IUT) has been the standard of care, but
131 maternal administration of intravenous immunoglobulin (IVIg) has also been reported⁷. The
132 postnatal care of affected newborns has also been the subject of debate among neonatologists and
133 hematologists.

134 The Delphi methodology has been recommended as a qualitative method for reaching a
135 consensus on topics where existing literature cannot quantitatively address unresolved challenges⁸.
136 This method has been widely used in fetal medicine, as the rarity of conditions and ethical
137 challenges make it unlikely to run clinical trials to resolve controversies⁹⁻¹¹. Accordingly, in the
138 current study, we aim to address the controversies in screening, monitoring, and prenatal and
139 postnatal management of HDFN with consensus from a multidisciplinary and international panel
140 of experts.

141 **Delphi Methodology and Participants**

142 *Delphi design*

143 The Delphi methodology was used. This methodology consists of scoring a series of
144 structured statements that are revised, fed back to the participants, and repeated in multiple rounds
145 in increasing detail until consensus has been reached.¹² This procedure aims to refine participating
146 experts' opinions while minimizing confounding factors present in other group response
147 methods.¹³ The rationale for its use is that it is a well-established instrument with which to reach
148 a consensus from a panel of experts on research questions that cannot be answered with empirical
149 evidence and complete certainty. Participants provided informed consent before commencing the
150 first round and were reminded of their right to anonymity, and the ability to withdraw before each
151 subsequent round. Institutional Review Board (IRB) exempt approval was obtained from Indiana
152 University with IRB # 21347

153 *Panel selection*

154 The study core group (the authors of this study) identified key stakeholder experts (working
155 group) who consisted of general obstetricians, maternal-fetal medicine specialists, neonatologists,
156 and hematologists. Eligibility for experts' participation was based on at least one of the following
157 inclusion criteria: expertise in the management of HDFN, based on a relevant publication record.
158 Second, membership in pertinent scientific organizations, including the Society for Maternal-Fetal
159 Medicine (SMFM), the North American Fetal Therapy Network (NAFTNet), the International
160 Fetal Medicine and Surgery Society (IFMSS), and the International Society for Prenatal Diagnosis
161 (ISPD). Lastly, invitees were asked to nominate other specialists with relevant expertise. Potential

162 participants were sent an invitational email with a detailed description of the background, goals,
163 methodology, and selection criteria. The intended sample size was above 50 participants to ensure
164 sufficient international representation of expert views.

165 First round

166 Five domains were used to structure the first round: (i) prenatal workup (ii) prenatal
167 monitoring and management (iii) IUT (iv) labor and delivery, and (v) postnatal workup and
168 management.

169 Response options included multiple choice answers or a 5-point Likert scale (with 1
170 representing Strongly Agree and 5 as Strongly Disagree). A pre-defined cut-off for group
171 consensus on an item or group of similar answers was $\geq 70\%$ ¹⁴. Items with 60-69% agreement were
172 reconsidered in the next round, while those with $< 60\%$ agreement reflected a lack of consensus,
173 and follow-up questions regarding these items were not posed in the subsequent round unless
174 rewords were felt to be necessary. Participants were able to provide feedback or suggest additional
175 items which were used to adjust the questions and answer choices by the research core group.

176 Subsequent round

177 Items that reached consensus were presented to the panel for confirmation in the second
178 round. Items with significant agreement (60-69%) were reconsidered following rephrasing the
179 question-and-answer options, or a new question was added to clarify. Items with $< 60\%$ agreement
180 were determined to be no consensus items. Additional suggested items were discussed among the
181 steering group before introduction in the next round.

182 Data collection and analysis

183 Data were collected in each round using online questionnaires that were presented to
184 panelists through a unique token-secured link for each round. Responses were captured in REDCap
185 version 13.7.19 (Vanderbilt University, Nashville, TN). Non-responders received reminder emails
186 after two and four weeks and were excluded from subsequent survey rounds if no response was
187 obtained. The panel categorized and considered newly suggested items carefully for their
188 applicability in this procedure. Experts' demographics and practice characteristics were collected.
189 Analyses were performed using REDCap and presented in frequency tables.

190 Participants

191 One hundred and seven experts participated in the first round of which 100 (93.5%)
192 completed the subsequent round. Experts' demographic characteristics are outlined in Table 1. The
193 expert panel represented 25 countries across six continents. Among our panel, 76 (71.0%) were
194 maternal-fetal medicine specialists, 22 (20.6%) neonatologists, and 9 (8.4%) hematologists.
195 Experts that had more than 20 years of practice experience made up 48.6% of the expert panel.
196 53.3% of the experts assessed 15 or more cases of fetal anemia per year (Table 1).

197 **Prenatal Workup of Pregnancies at Risk or Complicated with Alloimmunization**

198 The expert panel agreed on using cell-free DNA (cfDNA) to determine fetal genotype
199 (75.3% agreement). Related to the use of cfDNA, experts agreed (>70% agreement) on using
200 cfDNA for RhD, Kell, and Rhc antigens and agreed on using it in all twin gestations, both
201 monochorionic and dichorionic (89.2% agreement). Additionally, while there was no consensus,
202 there was a significant agreement to not continue monitoring antibody titers (63.3% agreement)

203 and to not initiate or continue MCA Dopplers monitoring (66.7% agreement) if cfDNA showed
204 fetus is not at risk for HDFN (Table 2). We conducted a subgroup analysis on the responses from
205 US, non-US (including Europe), and Europe. In pregnancies with maternal alloimmunization, if
206 cfDNA suggests that the fetus is not at risk for HDFN, the proportion of experts who agreed that
207 monitoring maternal antibody titers is not required was 71.4%, 59.6%, and 57.5%, respectively. In
208 pregnancies with RBC alloimmunization and elevated titers, if cfDNA testing suggests that the
209 fetus is not at risk for hemolytic disease, the following percentages of experts from the mentioned
210 regions agreed not to perform MCA Doppler for fetal anemia surveillance: 64.3%, 66.0%, and
211 70.0%.

212 There was consensus that 16 should be the critical titer threshold for all antigens known to
213 cause HDFN, except Kell (86.4% agreement). While there was no consensus for Kell antigens,
214 experts had 62.9% agreement on using titers of 4 to define critical titers requiring fetal anemia
215 imaging monitoring (Table 2). 64.2% of experts agreed on not repeating antibody titers once they
216 reach a predefined critical level but rather initiating MCA Dopplers for fetal anemia monitoring.
217 The MCA Doppler assessment should be started at 16 weeks gestation (80.2% agreement), weekly
218 assessment (61.7% agreement), to be continued until delivery (93.8% agreement) (Table 2).
219 Regarding ultrasound parameters to assess for fetal anemia, experts did not reach consensus on the
220 use of prenatal surveillance with non-stress tests (NST) or biophysical profile (BPP) (44.4%
221 agreement).

222 A few items that did not reach consensus include anti-M management and how often to
223 repeat titers. Further practices related to prenatal testing and monitoring that were included in the
224 Delphi can be seen in Table 2.

225 We included anti-M alloimmunization due to controversy in the literature about its role. It's
226 crucial to determine if it leads to alloimmunization, using lab methods like serological testing,
227 RBC phenotyping, and molecular genotyping. Although consensus wasn't reached, this topic is
228 vital for accurate diagnosis and management.

229 There are varying recommendations in current clinical guidelines concerning the prenatal
230 assessment of fetal anemia risk using cfDNA. The American College of Obstetrics and
231 Gynecology (ACOG) states that the use of cfDNA is a “reasonable alternative” for fetal RhD
232 testing in patients who are at risk and decline amniocentesis to perform PCR on fetal amniocytes.¹⁵

233 Guidelines recommend screening for RBC alloimmunization by testing for antibodies to RBC
234 antigens and measuring titer levels.^{5,16} ACOG indicates that critical titer thresholds, which prompt
235 further assessment, range from 8 to 32 due to laboratory variations. Typically, titers of 8 or less
236 may warrant monitoring titers every 4 weeks.⁵ The Royal College of Obstetricians and
237 Gynaecologists (RCOG) identifies critical thresholds for anti-D antibodies as >15 IU/mL, though
238 this is not recommended as best practice. Additionally, RCOG recommends referral to a fetal
239 medicine center with anti-D levels ≥ 4 IU/mL. Their recommendation of best practice for anti-Kell
240 antibodies is that referral should occur as soon as they are detected due to the risk of severe anemia
241 at lower titer levels.¹⁶ It is important to note the difference in methodology of alloimmunization
242 assessment between the ACOG and RCOG guidelines. ACOG uses titers which are a dilution of
243 the antibody level in maternal serum to a level where agglutination is not seen and RCOG reports

244 the autoanalyzer method which directly measures the antibody level in the maternal serum.^{5,16} In
245 a 16-year unselected cohort of 1026 Kell-immunized pregnancies, a titer of ≥ 4 was determined
246 to be used as the target for regular clinical monitoring.¹⁷ A Canadian study verifies that their results
247 correspond with the current “critical titer threshold” of 8-32.^{18,19} The expert panel agreed on the
248 critical titer threshold of non-Kell antibodies as 16 and significant agreement but not consensus
249 for Kell critical threshold ≥ 4 . Although consensus was reached on the critical antibody titers, the
250 provider must be certain of their lab methodology as the consensus from this study applies only to
251 titers generated from the indirect Coombs test, unlike the various other methodologies used to
252 determine maternal alloantibody titers/levels.

253 In addressing the differences in Rh antibody assessment methods, it is crucial to emphasize
254 that our Delphi study findings are based solely on the indirect Coombs method. It is recommended
255 that readers be fully aware of the specific methodology used by their laboratories. Additionally,
256 there are various methods for Rh antibody detection, including gel microcolumn agglutination
257 assay and automated solid phase, as well as autoanalyzer measurements. Notably, some European
258 centers utilize an autoanalyzer measurement of maternal serum Rh antibodies ≥ 15 IU/ml to
259 identify fetuses at risk for moderate-severe anemia, while others suggest lower thresholds (≥ 6
260 IU/ml and ≥ 3.5 IU/ml) based on recent studies.²⁰ These differences underscore the importance of
261 understanding the specific methods and cutoffs used in practice to accurately interpret and apply
262 our findings.

263 **Prenatal Management of Pregnancies with Alloimmunization**

264 Regarding the maternal administration of IVIg, 54/77 (70.0%) experts agreed that it should
265 be considered an option for managing pregnancies with HDFN. Based on our results, IVIg is
266 indicated in pregnancies with prior fetal or neonatal death due to HDFN or a history of IUT before
267 24 weeks in a previous pregnancy. It is recommended to be started at 10-14 weeks of gestation,
268 (77.4%) with no loading dose and a maintenance dosage of 1 g/kg per week (75.5%) . MCA
269 Doppler monitoring needs to be done weekly while receiving IVIg, and if it suggests fetal anemia,
270 IVIg should be stopped and IUT needs to be offered (92.5%) (Table 3).

271 With regard to monitoring these patients, American Medical Society guidelines do not have a
272 recommendation for an exact gestational age at which to initiate MCA Doppler assessments. The
273 SMFM guideline suggests that MCA Doppler assessment be initiated at a GA when fetal blood
274 sampling procedures or IUTs are technically feasible, such as 18-20 weeks gestation. However,
275 weekly assessment has been recommended following 24 weeks gestation.^{5,21} RCOG mentions that
276 if the fetus is antigen-positive and maternal antibody titers are at a critical level, MCA Dopplers
277 should be initiated and monitored weekly. ACOG and RCOG advised caution with monitoring
278 MCA Doppler beyond 34-35 and 36 weeks gestation, respectively, due to their decreased
279 sensitivity for the detection of fetal anemia beyond this gestational age.^{5,16} Our expert panel agreed
280 on the initiation of MCA Doppler assessments starting at 16 weeks and continuing weekly until
281 delivery.

282

283 **Intrauterine Transfusion**

284 Consensus was not reached regarding the minimum gestational age (GA) for intravenous
285 (IV) IUT. There was consensus that the maximum GA should be 35^{0/7} to 35^{6/7} weeks (78.5%
286 agreement). The expert panel was asked if the threshold of MCA peak systolic velocity (PSV) is
287 similar in early IUT (<20 weeks) as IUT >20 weeks and the panel recommended that IUT should
288 be offered for pregnancies less than 20 weeks when the MCA PSV is persistently higher than 1.5
289 MoM (92.9% agreement). In patients where cord IV IUT is not feasible, our panel agreed that
290 intra-peritoneal (IP) and intra-hepatic access should be considered. However, a combination of
291 intra-peritoneal transfusion (IPT) and intravascular transfusion (IVT) should not be routinely used
292 in the same procedure (92.3% agreement) (Table 4).

293 To calculate the transfusion volume, hemoglobin and/or hematocrit can be used (92.9%
294 agreement), and for IVT, the same calculation formula should be used regardless of GA (88.5%
295 agreement). Concerning monitoring, the timing of the second and third IUT should be determined
296 by a combination of weekly MCA-PSV and the calculated rate of hemoglobin decline, which is
297 approximately 0.9 g/dL per day in 10-14 days after the first IUT and 0.6 g/dL per day in 2-3 weeks
298 after the second IUT (75.7% agreement). Repeat intrauterine transfusions are typically considered
299 when the MCA-PSV exceeds 1.5 MoM. This indicates moderate to severe fetal anemia. As for the
300 Hb level, repeat transfusions are usually considered when the Hb level falls below 10 g/dL.

301 In dichorionic-diamniotic twin pregnancies, fetal blood sampling should not be performed
302 on both fetuses when only one has an abnormal MCA Doppler finding (85.9% agreement) (Table
303 4).

304 The Dodd et al. clinical trial, in 2018, concluded that both MCA-PSV and estimation of
305 fetal hemoglobin drop calculation can be used to determine the timing of the second IUT due to
306 the lack of statistically significant differences in outcomes and complications between the two
307 groups.²² In line with this study, our expert panel also recommended that the timing of the
308 subsequent IUT should be determined by a combination of weekly MCA-PSV and hemoglobin
309 drop calculations. Additionally, it is important to note that while the expert panel reached
310 significant agreement on the target hemoglobin concentration for intrauterine transfusions (IUT)
311 at 14-16 g/dL, evidence in the literature suggests that, in cases of severely anemic fetuses, it is
312 advised not to increase the fetal hemoglobin by more than four-fold the starting hemoglobin level
313 in one IUT session. Instead, these patients should undergo a subsequent IUT 48 hours later to
314 achieve the target hemoglobin concentration.

315 Procedural-related practices, the panel agreed on using a 20 gauge needle after 22-24
316 weeks, while 22 gauge prior to that (81.4% agreement). Type and first line of maternal anesthesia
317 is not affected by GA (85.9% agreement) with local or local with maternal sedation is the most
318 commonly used type (88.5%). While no consensus was reached, 67.1% agreed that fetal paralytic
319 medicine should be considered in IUT, 68.1% agreed on using hemoglobin of 14-16 g/dL or
320 hematocrit of 40-45% as a target level for transfusion, and to proceed for emergency cesarean
321 delivery only if no improvement in fetal status following a trial of intrauterine resuscitation for
322 fetal bradycardia (90.0%) (Table 4).

323 Regarding IPT, experts who performed at least 5 procedures (49/78, 62.8%) were given
324 open-ended questions given that the procedure is infrequent. Practices were variable related to IPT
325 volume of transfusion and timing of transfusion following IPT. Still, overall experts are

326 considering between 5-10 mL of transfusion for pregnancies at ≤ 20 weeks and considering MCA
327 Doppler monitoring 1-2 weekly to trigger possible repeat transfusion after one week.

328 **Delivery Management**

329 Regarding delivery, the route needs to be determined by obstetric indications (98.7%). The
330 timing of delivery for patients with critical antibody titer and no IUT is recommended to be 37^{0/7}
331 - 38^{6/7} weeks (100.0%) and if IUT was needed to be two to three weeks following the last IUT
332 (89.6%). The panel agreed on delayed cord clamping (71.4%) but not cord milking in pregnancies
333 that had IUT (85.7%) (Table 5).

334 Based on the results outlined in the sections above, the participating experts were able to
335 generate a monitoring and clinical management workflow as seen in Figure 1.

336 **Postnatal Workup and Management**

337 In this section, 18 statements were presented to 25 experts from various fields and
338 specialties involved in the postnatal care of these neonates (Table 6). Our panel agreed that the
339 postnatal treatment of fetal anemia should include intensive phototherapy, blood transfusion, and
340 exchange transfusion, without the routine administration of IVIg. The thresholds for phototherapy
341 and exchange transfusion should be determined by the American Academy of Pediatrics (AAP)
342 2022 guidelines (72% agreement). Anemia labs should be investigated in neonates of pregnancies
343 complicated with alloimmunization before hospital discharge (92.0% agreement) and if IUT was
344 needed for the labs to be repeated in one week following discharge if they were not anemic at birth
345 (84.0% agreement). In newborns of pregnancies that had critical titers but did not require IUT,

346 96% agreed that anemia labs should be performed before hospital discharge. 96.0% agreed that
347 exchange transfusions should be centralized in hospitals with sufficient exposure and experience
348 and 92.0% agreed that they do not use erythropoietin-stimulating agents in the first weeks of life
349 to reduce the need for blood transfusion. The panel agreed that the hemoglobin cut-off level to
350 consider transfusion following hospital discharge is 7 g/dL (92.0% agreement), and the newborns
351 need to be monitored frequently until 2-3 months of age (96.0% agreement) (Table 6).

352 Regarding the postnatal management of neonates affected by HDFN, the 2022 AAP
353 guidelines mentioned that the use of IVIg is an optional therapy, and similarly, our panel did not
354 reach a consensus on its routine use.²³ It is important to note that with updates to AAP guidelines,
355 the relevance of the consensus reached on the use of neonatal IVIg may be altered.

356 Although 92% of the expert panel stated that they are not routinely using erythropoietin-
357 stimulating agents in the first few weeks of life to reduce the need for transfusion, a recently
358 published randomized controlled trial showed that darbepoetin alfa decreased the number of
359 transfusion episodes.²⁴

360 **Strengths and Limitations**

361 The strengths of our study include the use of the well-established Delphi procedure and the
362 inclusion of a diverse group of international experts. Our selection criteria based on clinical and
363 academic experience resulted in a high degree of expertise among our participants. Moreover, a
364 relatively low attrition rate was achieved across rounds. We were able to provide insight into how
365 experts synthesize conflicting data, and demonstrate choices that are made when no high-quality
366 data exist, and build on current knowledge gaps alloimmunization monitoring and management.

367 This may drive the further collection of evidence for treatment efficacy but also provide a useful
368 guide for shared decision-making and treatment assessment.

369 Limitations include that the Delphi output reflects the contemporary interpretation of
370 existing literature which can change over time. As a summary of expert opinion, it also provides
371 different insight than that provided by a systematic review or society guidelines. Additionally,
372 given the presentation of consensus results in follow-up rounds, participants may have altered their
373 initial thoughts to prioritize the consensus views to emphasize group unanimity²⁵. This was
374 minimized by masking individual expert opinions that could steer the group in a particular
375 direction, adding relevant questions raised by individual participants guided by a working group,
376 and the independent nature of the questionnaire itself. Another limitation is the overrepresentation
377 of Western world countries and the underrepresentation of countries from Africa, Asia, and South
378 America. This represents the views of a selected group of participants, and it cannot be known
379 how representative it is of the wider community. Lastly, this Delphi consensus can be used as a
380 guideline for areas where consensus was reached. However, in areas where consensus was not
381 reached, providers should apply a patient-by-patient approach to determine the clinical course of
382 action.

383 **Scope for future research:**

384 **1. Prevention Strategies:** Exploring new preventive measures, such as novel
385 immunoglobulin therapies or vaccines, to reduce the incidence of HDFN. This includes
386 awaiting the completion of the phase III randomized, double-blind, placebo-controlled trial
387 of nipocalimab to prove the potential benefit in preventing the need for serial IUTs in
388 HDFN.

- 389 **2. Long-term Outcomes:** Studying the long-term health impacts on children who were
390 affected by HDFN, including neurodevelopmental outcomes and quality of life.
- 391 **3. Global Health Initiatives:** Addressing disparities in HDFN care by conducting research
392 in low-resource settings and developing cost-effective treatment protocols.
- 393 **4. Ethical and Social Considerations:** Investigating the ethical, legal, and social
394 implications of HDFN management, including informed consent and access to care.

395 **Conclusions**

396 Although experts agreed on many aspects of monitoring and management of red cell
397 alloimmunization and HDFN, the results of this Delphi show apparent practice variations
398 worldwide. This Delphi survey facilitated the development of a consensus-based clinical
399 workflow that can be used to enhance clinical practice, improve outcomes, and facilitate future
400 research. Non-consensus items should be viewed as areas where clinical judgment remains crucial,
401 and where further evidence and expert discussion are needed to develop more definitive guidelines.

402

403 **COLLABORATORS**

404 The following are members of the HDFN Delphi working group that provided consent to be
 405 included in the acknowledgment:

Name	Institution	City	Country
Ahmed A Nassr	Baylor College of Medicine and Texas Children's Fetal Center	Houston	United States
Ahmet Baschat	Johns Hopkins University	Baltimore	United States
Alexander Hohnecker	Klinikum Dritter Orden München	München/Munich	Germany
Alireza Shamshirsaz	Harvard Medical School	Boston	United States
Angel Luciano	Johns Hopkins All Childrens Hospital	St Petersburg	United States
Anne Debeer	UZ Leuven	Leuven	Belgium
Annegret Geipel	University Hospital Bonn	Bonn	Germany
Antoni Borrell	Hospital Clinic Barcelona	Barcelona	Spain
Asma Khalil	St George's Hospital, University of London	London	United Kingdom
Aurora Viejo Llorente	Hospital La Paz	Madrid	Spain
Beate Mayer	Charité - Universitätsmedizin Berlin	Berlin	Germany
Borna Poljak	Liverpool Women's Hospital	Liverpool	United Kingdom
C. Ellen van der Schoot	Sanquin Research	Amsterdam	Netherlands
Catherine Taillefer	CHU Ste-Justine/Université de Montréal	Montréal	Canada
Christof Dame	Charité - Universitätsmedizin Berlin	Berlin	Germany
Christoph Berg	Universitätsklinikum Köln	Cologne	Germany
Conrado Milani Coutinho	Hospital das Clínicas da Faculdade de Medicina de Ribeirão Preto da Universidade de São Paulo	Ribeirão Preto	Brazil
Derek P. de Winter	Leiden University Medical Center	Leiden	Netherlands
Dick Oepkes	Leiden University Medical Center	Leiden	Netherlands
EJT Verweij	LUMC	Leiden	Netherlands
Elena Carreras Moratonas	Hospital Universitari Vall d'Hebron	Barcelona	Spain
Eleonor Tiblad	Karolinska Institutet	Stockholm	Sweden
Ellen Bendel-Stenzel	Mayo Clinic	Rochester, MN	United States
Emeline Maisonneuve	Centre Hospitalier Universitaire Vaudois	Lausanne	Switzerland
Enrico Lopriore	Leiden University Medical Center	leiden	Netherlands
Evangelia Vlachodimitropoulou	King's College London	London	United Kingdom
Federico Prefumo	IRCCS Istituto Giannina Gaslini	Genova	Italy
Fernando Maia, Peixoto-Filho	Universidade Estadual do Rio de Janeiro - UERJ	Rio de Janeiro	Brazil
Francisca S. Molina	Hospital Universitario Clínico San Cecilio.	Granada	Spain
Gerardo Sepulveda Gonzalez	Instituto de Salud Fetal	Monterrey NL Mexico	Mexico
Glenn Gardener	Mater Mothers Hospital	Brisbane	Australia

Haruhiko Sago	Sanno Birth Center	Tokyo	Japan
Helen Liley	Mater Mothers' Hospital	Brisbane, QLD	Australia
Hiba Mustafa	Indiana University and Riley Children's Hospital	Indianapolis	United States
Ingrid Schwach	Federal University of São Paulo(1) Faculty of Medical Sciences of Santa Casa of São Paulo (2)	São Paulo	Brazil
Ivonne Bedei	Justus-Liebig University	Giessen	Germany
James Castleman	Birmingham Women's and Children's NHS Foundation Trust	Birmingham	United Kingdom
Jana Lozar Krivec	University Medical Centre Ljubljana	Ljubljana	Slovenia
Jean-marie jouannic	CNRHP Trousseau Hospital, APHP Sorbonne University	paris	France
Jena Miller	Johns Hopkins Center for Fetal Therapy	Baltimore	United States
Joana Filipa Pereira Nunes	Unidade Local de Saúde São João	Porto	Portugal
Johanna Middeldorp	Leiden University Medical Center	Leiden	Netherlands
Josep M Martinez	BCNatal Hospital Clinic and Sant Joan de Deu	Barcelona	Spain
Karin Sundberg	Rigshospitalet	Copenhagen	Denmark
Katherine Bligard	Washington University School of Medicine	Saint Louis	United States
Katherine Kohari	Yale	New Haven	United States
Keisuke Ishii	Osaka Women's and Children's Hospital	Izumi	Japan
Kenneth Moise	Dell Medical School - UT Austin	Austin	United States
Kévin Le Duc	Centre Hospitalier Universitaire de Lille	Lille	France
Liesbeth Lewi	UZ Leuven	Leuven	Belgium
Lizelle Van Wyk	Stellenbosch University	Cape Town	South Africa
Lucas Otaño	Hospital Italiano de Buenos Aires	Buenos Aires	Argentina
Luming SUN	Shanghai First Maternity & Infant Hospital of TongJi University	Shanghai	China
Lut Geerts	Stellenbosch University	Cape Town	South Africa
M Angeles, SANCHEZ-DURAN	HOSPITAL VALL HEBRON	BARCELONA	Spain
Mar Bennasar	BCNatal. Maternal Fetal and Neonatal Center of Barcelona. Hospital Clínic.	Barcelona	Spain
Marcella Vaena	Instituto Fernandes Figueira-FIOCRUZ	Rio de Janeiro	Brazil
Maria M Gil	Hospital Universitario de Torrejon	Madrid	Spain
Mark F Weems	University of Tennessee Health Science Center	Memphis	United States
Mark Kilby	Birmingham Women's and Children's Foundation Trust / University of Birmingham, UK	Birmingham	United Kingdom
Masja de Haas	Sanquin	Amsterdam	Netherlands
Matthew Saxonhouse	Wake Forest School of Medicine	Charlotte	United States
Mauro Schenone	Mayo Clinic	Rochester, MN	United States
Mert Ozan Bahtiyar	Yale School of Medicine	New Haven	United States
Michael v. Zaretsky, Md	Colorado Fetal Care Center	Aurora, Colorado	United States
Miguel Angel Martinez Rodriguez	Medicina Fetal Mexico	Guadalajara	Mexico
Mounira Habli	Cincinnati Children Hospital	Cincinnati	United States
Nahla Khalek, MD, MPH, MEd	Richard D. Wood Jr. Center for Fetal Diagnosis and Treatment at Children's Hospital of Philadelphia	Philadelphia	United States
Natalie Frost	Dell Children's Medical Center	Austin	United States

Pamela Griffiths	Phoenix Children's	Phoenix	United States
Paul Maurice	CNRHP Trousseau Hospital AP-HP. Sorbonne University	Paris	France
Pe'er Dar	Montefiore Medical Center/ Albert Einstein College of medicine	Bronx NY	United States
Peter Lindgren	Karolinska Institutet	Stockholm	Sweden
Petya Chaveeva	Dr Shterev Hospital	Sofia	Bulgaria
Philipp Klaritsch	Medical University of Graz	Graz	Austria
Prathima Radhakrishnan	Bangalore Fetal Medicine Centre	Bengaluru	India
Rahel Schuler	Department of General Pediatrics and Neonatology, Justus-Liebig- University	Giessen	Germany
Ramen Chmait, MD	University of Southern California	Los Angeles	United States
Rebecca Rose	Indiana University	Indianapolis	United States
Renske van 't Oever	Leiden University Medical Center (LUMC)	Leiden	Netherlands
Riina Jernman	Helsinki University Hospital, Obstetrics and Gynecology	Helsinki	Finland
Robert Christensen	University of Utah and Intermountain Health	Salt Lake City	United States
Robert Cincotta	Mater Mothers Hospital	Brisbane	Australia
Roland Axt-Fliedner	Division of Prenatal Medicine&Fetal Therapy	Gießen&Marburg	Germany
Roland Devlieger	University Hospital Leuven	Leuven	Belgium
Roopali Donepudi	Baylor College of Medicine	Houston	United States
Sailesh Kumar	Mater Mother's Hospital and Mater Research Institute	Brisbane	Australia
Stefan Verlohren	Charité - Universitätsmedizin Berlin	Berlin	Germany
Stephen P. Emery, MD	UPMC Magee-Womens Hospital	Pittsburgh, PA	United States
Susanna Sainio	Finnish Red Cross Blood Service	Vantaa	Finland
Suwan Mehra	Advocate Childrens Hospital	Chicago	United States
Tanja Premru-Srsen	Department of Perinatology, UMC Ljubljana; Faculty of Medicine, University of Ljubljana	Ljubljana	Slovenia
Tobias Legler, MD	University Medical Center Goettingen	Goettingen	Germany
Ulrich J. Sachs	Giessen University Hospital	Giessen	Germany
Vandana Basal	Nowrosjee Wadia Maternity Hospital	Mumbai	India
Vedran Stefanovic	Department of OB/GYN, Helsinki University Hospital	Helsinki	Finland
William Goodnight	University of North Carolina Health Fetal Care Center	Chapel Hill	United States
Yair Blumenfeld	Stanford University	Palo Alto, CA	United States

406

407 **ACKNOWLEDGEMENTS**

408 We would like to thank our expert panel for participation in our Delphi procedure.

409

410

411 **References**

- 412 1 Mittendorf, R. & Williams, M. A. Rho(D) immunoglobulin (RhoGAM): how it came into being.
413 *Obstet Gynecol* **77**, 301-303, doi:10.1097/00006250-199102000-00029 (1991).
- 414 2 Yu, D., Ling, L. E., Krumme, A. A., Tjoa, M. L. & Moise, K. J., Jr. Live birth prevalence of hemolytic
415 disease of the fetus and newborn in the United States from 1996 to 2010. *AJOG Glob Rep* **3**,
416 100203, doi:10.1016/j.xagr.2023.100203 (2023).
- 417 3 Koelewijn, J. M. *et al.* Diagnostic value of laboratory monitoring to predict severe hemolytic
418 disease of the fetus and newborn in non-D and non-K-alloimmunized pregnancies. *Transfusion*
419 **60**, 391-399, doi:10.1111/trf.15631 (2020).
- 420 4 Donepudi, R. V. *et al.* Perinatal survival following intrauterine transfusion for red cell
421 alloimmunized pregnancies: systematic review and meta-regression. *Am J Obstet Gynecol* **230**,
422 e1-e2, doi:10.1016/j.ajog.2023.10.016 (2024).
- 423 5 ACOG Practice Bulletin No. 192: Management of Alloimmunization During Pregnancy. *Obstet*
424 *Gynecol* **131**, e82-e90, doi:10.1097/aog.0000000000002528 (2018).
- 425 6 Green-top Guideline No. 65: The Management of Women with Red Cell Antibodies during
426 Pregnancy. *The Obstetrician & Gynaecologist* **16**, 224-224,
427 doi:https://doi.org/10.1111/tog.12125 (2014).
- 428 7 Mustafa, H. J. *et al.* Intravenous immunoglobulin for the treatment of severe maternal
429 alloimmunization: individual patient data meta-analysis. *Am J Obstet Gynecol*,
430 doi:10.1016/j.ajog.2024.03.044 (2024).
- 431 8 Niederberger, M. & Spranger, J. Delphi Technique in Health Sciences: A Map. *Front Public Health*
432 **8**, 457, doi:10.3389/fpubh.2020.00457 (2020).
- 433 9 Krispin, E. *et al.* Consensus protocol for management of early and late twin-twin transfusion
434 syndrome: Delphi study. *Ultrasound Obstet Gynecol* **63**, 371-377, doi:10.1002/uog.27446 (2024).
- 435 10 Oyelese, Y. *et al.* Vasa previa in singleton pregnancies: diagnosis and clinical management based
436 on an international expert consensus. *Am J Obstet Gynecol*, doi:10.1016/j.ajog.2024.03.013
437 (2024).
- 438 11 Khalil, A. *et al.* Consensus definition and essential reporting parameters of selective fetal growth
439 restriction in twin pregnancy: a Delphi procedure. *Ultrasound in Obstetrics & Gynecology* **53**, 47-
440 54, doi:https://doi.org/10.1002/uog.19013 (2019).
- 441 12 Murphy MK *et al.* Consensus development methods, and their use in clinical guideline
442 development. *Health technology assessment (Winchester, England)* **2** (1998).
- 443 13 Sinha IP, Smyth RL & PR, W. Using the Delphi technique to determine which outcomes to
444 measure in clinical trials: recommendations for the future based on a systematic review of
445 existing studies. *PLoS medicine* **8**, doi:10.1371/journal.pmed.1000393 (2011).
- 446 14 Hohmann, E., Cote, M. P. & Brand, J. C. Research pearls: expert consensus based evidence using
447 the Delphi method. *Arthroscopy: The Journal of Arthroscopic & Related Surgery* **34**, 3278-3282
448 (2018).
- 449 15 ACOG Clinical Practice Update: Paternal and Fetal Genotyping in the Management of
450 Alloimmunization in Pregnancy. *Obstetrics & Gynecology* **144**, e47-e49,
451 doi:10.1097/aog.0000000000005630 (2024).
- 452 16 RCOG. The Management of Women with Red Cell Antibodies during Pregnancy. (2014).
453 <https://www.rcog.org.uk/media/oykp1rtg/rbc_gtg65.pdf>.
- 454 17 Slootweg, Y. M. *et al.* Predicting anti-Kell-mediated hemolytic disease of the fetus and newborn:
455 diagnostic accuracy of laboratory management. *Am J Obstet Gynecol* **219**, 393.e391-393.e398,
456 doi:10.1016/j.ajog.2018.07.020 (2018).

- 457 18 Judd, W. J. Practice guidelines for prenatal and perinatal immunohematology, revisited.
458 *Transfusion* **41**, 1445-1452, doi:10.1046/j.1537-2995.2001.41111445.x (2001).
- 459 19 Zwingerman, R., Jain, V., Hannon, J., Zwingerman, N. & Clarke, G. Alloimmune Red Blood Cell
460 Antibodies: Prevalence and Pathogenicity in a Canadian Prenatal Population. *J Obstet Gynaecol*
461 *Can* **37**, 784-790, doi:10.1016/s1701-2163(15)30148-1 (2015).
- 462 20 Nicolaidis, K. H. & Rodeck, C. H. Maternal serum anti-D antibody concentration and assessment
463 of rhesus isoimmunisation. *Bmj* **304**, 1155-1156, doi:10.1136/bmj.304.6835.1155 (1992).
- 464 21 Mari, G. *et al.* Society for Maternal-Fetal Medicine (SMFM) Clinical Guideline# 8: the fetus at risk
465 for anemia—diagnosis and management. *American journal of obstetrics and gynecology* **212**,
466 697-710 (2015).
- 467 22 Dodd, J. M. *et al.* Fetal middle cerebral artery Doppler to time intrauterine transfusion in red-cell
468 alloimmunization: a randomized trial. *Ultrasound Obstet Gynecol* **51**, 306-312,
469 doi:10.1002/uog.18807 (2018).
- 470 23 Kemper, A. R. *et al.* Clinical practice guideline revision: management of hyperbilirubinemia in the
471 newborn infant 35 or more weeks of gestation. *Pediatrics* **150** (2022).
- 472 24 Ree, I. M. C. *et al.* Darbepoetin alfa to reduce transfusion episodes in infants with haemolytic
473 disease of the fetus and newborn who are treated with intrauterine transfusions in the
474 Netherlands: an open-label, single-centre, phase 2, randomised, controlled trial. *Lancet*
475 *Haematol* **10**, e976-e984, doi:10.1016/s2352-3026(23)00285-5 (2023).
- 476 25 Baron, R. S. So right it's wrong: Groupthink and the ubiquitous nature of polarized group
477 decision making. (2005).

478

479

480 **Table 1: Experts demographics and practice characteristics**

Characteristic	Respondents (n=107)
Age	
25-44	23 (21.5)
45-54	45 (42.1)
55-64	30 (28.0)
65+	9 (8.4)
Region of practice †	
United States	30 (28.0)
Germany	15 (14.0)
Netherlands	12 (11.2)
United Kingdom	9 (8.4)
Each of Australia, Brazil (total N=8)	4 (3.7)
Each of France, Finland, Belgium, Slovenia (Total N=12)	3 (2.8)
Each of India, Japan, Canada, South Africa, Sweden, Mexico (Total N=12)	2 (1.9)
Each of Argentina, Austria, Bulgaria, China, Denmark, Italy, Portugal, Singapore, Switzerland (Total N=9)	1 (0.9)
Speciality	
Obstetrics	76 (71.0)
Maternal-Fetal Medicine, performing intrauterine transfusion	71 (66.4)
Maternal-Fetal Medicine, not performing intrauterine transfusion	5 (4.7)
Neonatology	22 (20.6)
Hematology or immunohematology	9 (8.4)
Academic rank	
Professor	36 (33.6)
Specialist/Consultant	35 (32.7)
Associate / Assistant professor	31 (29.0)
Other	5 (4.7)
Practice setting	
University/Academic hospital-based practice	93 (86.9)
Community academic hospital-based practice	5 (4.7)
Private practice (independently or health system/hospital owned)	7 (6.5)
Other	2 (1.9)
Years in practice	
<5-9	18 (16.8)
10-19	37 (34.6)
>20	52 (48.6)
HDFN related participants' characteristics	
Number of assessments for fetal anemia per annum	
<5	5 (4.7)
5-14	29 (27.1)
15-34	29 (27.1)
>35	28 (26.2)
Uncertain	16 (15.0)
Number of intrauterine transfusions for fetal anemia per annum*	
<5	26 (26.3)

Characteristic	Respondents (n=107)
5-14	42 (42.4)
15-34	22 (22.2)
>35	3 (3.0)
Uncertain	14 (13.1)
Number of individuals who perform intrauterine transfusions at the centre	
1-2	40 (37.4)
3-4	53 (49.5)
>5	14 (13.1)
Number of IVIG procedures per annum**	
None /I do not offer IVIG	26 (26.5)
<5	60 (61.2)
5-24	5 (5.1)
>25	1 (1.0)
Uncertain	15 (14)
Number of deliveries following treatment for fetal anemia per annum*	
None	4 (4.0)
<5	26 (26.3)
5-24	53 (53.5)
>25	12 (12.1)
Uncertain	12 (11.2)
Number of HDFN pediatric cases per annum***	
None	1 (1.1)
<5	18 (19.8)
5-24	46 (50.5)
>25	16 (17.6)
Uncertain	26 (24.3)
Published papers in HDFN	65 (60.7)
Principal investigator and/or First author	46/65 (70.8)

481 *HDFN (Hemolytic Disease of the Fetus and Newborn); IVIG (Intravenous Immunoglobulins)*
482 *†N=100; *N=99 as 8 out of 107 participants indicated 'Not applicable to my specialty'; **N=98 as 9 out of 107*
483 *participants indicated 'Not applicable to my specialty'; ***N=91 as 16 out of 107 participants indicated 'Not*
484 *applicable to my specialty',*
485 *All percentages are presented in parentheses ()*
486

487 **Table 2: Prenatal Workup and Monitoring**

	Round in which item was included	
	1	2
	N=81	N=81
cfDNA		
cfDNA to determine fetal genotype in maternal alloimmunization	61/81 (75.3)	
Fetal antigens to test **		
RhD antigen	61/61 (100.0)	
Kell antigen	51/61 (86.9)	
Rhc antigen	44/61 (72.1)	
RhE antigen	36/61 (59.0)	
RhC antigen	31/61 (50.8)	
Rhe antigen	23/61 (37.7)	
Fya antigen	12/61 (19.7)	
JKa antigen	6/61 (9.8)	
Gestational age to initiate testing **		
10 weeks	18/61 (29.5)	
11 weeks	15/61 (24.6)	
12 weeks	16/61 (26.2)	
13 or 14 weeks	4/61 (6.6)	
cfDNA use to determine fetal genotyping in twin gestations	33 (89.2)	
MCA doppler surveillance should not be performed in alloimmunized pregnancies with high titres if cfDNA suggests no risk of HFDN	41/61 (67.2)	40/60 (66.7)
Antibody titers monitoring should not be continued in alloimmunized pregnancies if cfDNA suggests no risk of HFDN	41/61 (67.2)	38/60 (63.3)
Antibody titers		
Critical threshold for all antibodies known to cause HDFN, except Kell		
1:4 or 1:8	7/81 (8.6)	
1:16	54/81 (66.6)	70/81 (86.4)
1:32	10/81 (12.3)	
1:64 or >1:128	10/81 (12.3)	
Critical threshold for Kell antibodies		
Any positive Indirect Coombs test	20/81 (24.7)	
1:4	49/81 (60.5)	51/81 (62.9)
1:16	3/81 (3.7)	
≥1:32	0 (0.0)	
Critical threshold for Anti-M pregnancies		
Quantification of IgM vs IgG to determine high risk pregnancies	28/81 (34.6)	
Any positive indirect Coombs test †	4/81 (14.3)	
1:4	1/81 (3.6)	
1:16	13/81 (46.4)	
1:32	6/81 (21.4)	
1:64	4/81 (14.3)	
Antibody titres should NOT be repeated if they have reached critical threshold	53/81 (65.4)	52/81 (64.2)
Frequency of repeat titres if no critical threshold reached		
Weekly	1/81 (1.2)	

	Round in which item was included	
	1	2
	N=81	N=81
Every 2 weeks	21/81 (25.9)	
Every 4 weeks	42/81 (51.9)	
Once per trimester	1/81 (1.2)	
Changes with titers and gestational age	17/81 (20.9)	
Antibody titres should be obtained first in the current pregnancy to decide if MCA Dopplers are required, despite the level of the high antibodies in a prior pregnancy without treatment	49/81 (60.5)	54/81 (66.6)
Ultrasound		
Parameters to assess for fetal anemia		
Fetal middle cerebral artery (MCA) Doppler assessments	79/81 (97.5)	
Evaluation for hydrops fetalis	79/81 (97.5)	
Detailed anatomical survey (if not previously obtained)	69/81 (85.2)	
Cardiac function assessment (cardiac size, regurgitation, others)	59/81 (72.8)	
Antepartum surveillance (eg CTG/NST, BPP)	36/81 (44.4)	
UA Doppler assessment, including in the absence of FGR	30/81 (37.0)	
Measurements of fetal liver and spleen	22/81 (27.2)	
Earliest gestational age for MCA Doppler to reliably detect fetal anemia		
14 weeks	6/81 (7.4)	
15 weeks	4/81 (4.9)	
16 weeks	49/81 (60.5)	65/81 (80.2)
17 weeks	3/81 (3.7)	
18 weeks	19/81 (23.5)	
Frequency of MCA Doppler assessment once antibody titre threshold is reached and GA < 28 weeks		
Twice weekly	4/81 (4.9)	
Once weekly	50/81 (61.7)	50/81 (61.7)
Once biweekly	18/81 (22.2)	
Once every four weeks	2/81 (2.5)	
Individualized per titer and gestational age	7/81 (8.6)	
Gestational age at which MCA Doppler monitoring should cease if no IUT is required		
Up to 32 weeks	1/81 (1.2)	
Up to 35 weeks	10/81 (12.3)	
Up to 37 weeks	21/81 (26)	
Until delivery	49/81 (60.5)	76 (93.8)
The frequency of MCA Doppler monitoring should not change after a certain GA	58/81 (71.6)	
It should be increased if there was an incremental rise in MoMs (but this remains <1.5 in absence of other ultrasound signs)	53/81 (65.4)	53/81 (65.4)
MCA dopplers should cease if repeat antibody titres are below critical threshold	56/81 (69.1)	56/81 (69.1)

488 * Dark grey shading represents consensus (defined as $\geq 70\%$ agreement), light grey light grey represents significant
489 agreement (of 60-69%), and white represents no agreement (<60%). A '-' in a cell means that the issue was not
490 addressed in that round.

491 GA= Gestational age UA= umbilical doppler FGR= Fetal growth restriction MCA= middle cerebral artery MoMs,
492 † Test was not available in the region for 16 (19.8%) participants; **N=61; † N=28

493 All percentages are presented in parentheses ()

494 **Table 3: Prenatal Management with Intravenous Immunoglobulins**

	Round in which item was included	
	1 (N=77)	2
For eligible pregnancies, IVIg should be considered	54/77 (70.0)	
Indications in fetal anemia		
Prior fetal or neonatal death due to HDFN	38/53 (71.7)	
History of IUT at <24 weeks in the previous pregnancy	37/53 (70.0)	
Current MCA Dopplers >1.5 MoM at <16-18 weeks regardless of obstetric history	11/53 (20.8)	
Current critical antibody titres at <16-18 weeks + confirmed Current fetal genotype at risk, regardless of obstetric history	7/53 (13.2)	
History of IUT at any GA in the previous pregnancy	7/53 (13.2)	
GA at initiation of IVIg		
6-10 weeks	3/53 (5.7)	
10-14 weeks	41/53 (77.4)	
14-18 weeks or > 18 weeks	9/53 (17.0)	
Maximal GA at which IVIg should not be offered		
>14 weeks	3/53 (5.7)	
>16 weeks	6/53 (11.3)	
>17 weeks	1/53 (1.9)	
>18 weeks	6/53 (11.3)	
>20 weeks	19/53 (35.8)	
No GA limit	18/53 (33.9)	
GA at which IVIg should be stopped, given no signs of fetal anemia		
Up to 24 weeks	9/53 (17.0)	
Up to 26 weeks	2/53 (3.8)	
Up to 28 weeks	1/53 (1.9)	
Up to 32 weeks	9/53 (17.0)	
Up to 35 weeks	13/53 (24.5)	
Up to 37 weeks	7/53 (13.2)	
Until delivery occurs regardless of GA	12/53 (22.6)	
IVIg Dosing		
No loading dose + 1g/kg/week	40/53 (75.5)	
2 g/kg loading dose + 1g/kg/week every week after	5/53 (9.4)	
No loading dose + 2 g/kg every 3 weeks administered as 1 g/kg/day over 2 days	4/53 (7.5)	

	Round in which item was included	
	1	2
	(N=77)	
No loading dose + 0.5 g/kg/week	4/53 (7.5)	
MCA Doppler monitoring once every week while on IVIg	49/53 (92.5)	
If suggestive of fetal anemia, IVIG should be stopped and IUT started	49/53 (92.5)	

495 * Dark grey shading represents consensus (defined as $\geq 70\%$ agreement), light grey light grey represents
 496 significant agreement (of 60-69%), and white represents no agreement ($< 60\%$). A '-' in a cell means that
 497 the issue was not addressed in that round.

498 GA= Gestational age IVIg= Intravascular immunoglobulins IUT= intrauterine transfusion CS=
 499 Caesarean section MoM= Multiple of the Median

500 All percentages are presented in parentheses ()

501

502

503

504

505 **Table 4: Intrauterine transfusion**

	Round in which item was included	
	1	2
	(N=78)	(N=70)
Minimum GA for intravascular IUT in a non-hydropsic fetus		
14 weeks	1/78 (1.3)	
16 weeks	18/78 (23.1)	
18 weeks	36/78 (46.2)	
20 weeks	16/78 (20.5)	
22 weeks	5/78 (6.4)	
24 weeks	2/78 (2.6)	
Minimum GA for intravascular IUT in a hydropsic fetus		
14 weeks	6/78 (7.7)	
16 weeks	27/78 (34.6)	
18 weeks	34/78 (43.6)	
20 weeks	11/78 (14.1)	
IUT should be considered for pregnancies <20 weeks when the MCA is persistently >1.5 MoM	53/78 (67.9)	65/70(92.9)
Interventions in a non-hydropsic fetus when intracord IUT is not technically feasible		
Intraperitoneal transfusion	64/78 (82.1)	
Intrahepatic vein transfusion	58/78 (74.4)	
Abort the procedure and re-attempt later	17 (21.8)	
Interventions in a hydropsic fetus when intracord IUT is not technically feasible		
Intraperitoneal transfusion	60/78 (76.9)	
Intrahepatic vein transfusion	58/78 (74.4)	
Abort the procedure and re-attempt later	7/78 (9.0)	
Test to determine transfused volume in intravascular IUT		
Hb and/or Hct	47/78 (60.3)	65/70 (92.9)
Hct	17/78 (21.8)	
Hb	14/78 (17.9)	
In intravascular IUT the same calculation should be used if GA <24 weeks	69 (88.5)	
Intraperitoneal transfusion (IPT), experts who have performed at least 5 IPTs answered related open-ended questions (see results text)	49/78 (62.8)	
Use of peri-operative tocolysis in IUT	34/78 (43.6)	
Needle gauge choice should differ depending on GA at the procedure	47/78 (60.3)	42/70 (60)
22 gauge needle		

	Round in which item was included	
	1	2
	(N=78)	(N=70)
<20 weeks	21/47 (44.7)	
<22 weeks	10/47 (21.3)	
<24 weeks	7/47 (14.9)	
<26 weeks	3/47 (6.4)	
<28 weeks	6/47 (12.8)	
20 gauge needle should be used following the 22-24 weeks, while 22 gauge prior	30/47 (63.8)	57/70 (81.4)
18 needle gauge should not be used in IUT	36/47 (76.6)	
Maternal anaesthesia methods in IUT		
Methods		
Local or local and maternal sedation	69/78 (88.5)	
Regional	6/78 (7.6)	
General	3/78 (3.8)	
First-line choice for maternal anaesthesia is not influenced by GA	67/78 (85.9)	
Fetal paralytic medication should be considered in IUT	51/78 (65.3)	47/70 (67.1)
Intravascular	30/51 (58.8)	
Intramuscular	21/51 (41.2)	
MCA dopplers should be used to determine timing for the second IUT	52/78 (66.7)	43/70 (61.4)
MCA dopplers should not be used to determine timing for third IUT	44/78 (56.4)	
Timing of second transfusion in a non-hydropic fetus	N=78	
48 hrs	1 (1.3)	
72 hrs	1 (1.3)	
1 week	11 (14.1)	
2 weeks	22 (28.2)	
3 weeks	4 (5.1)	
Decide based on MCA Doppler assessment	33 (42.3)	
None of the above	5 (6.4)	
Timing of third transfusion in a non-hydropic fetus	N=78	
1 week	1 (1.3)	
2 weeks	16 (20.5)	
3 weeks	15 (19.2)	
Decide based on MCA Doppler assessment	29 (37.2)	
Based on MCA and post-transfusion levels	17 (21.8)	
Timing of second transfusion in a hydropic fetus	N=78	

	Round in which item was included	
	1	2
	(N=78)	(N=70)
48 hrs	11 (14.1)	
72 hrs	17 (21.8)	
1 week	27 (34.6)	
2 weeks	4 (5.1)	
Based on MCA and post-transfusion levels	19 (24.4)	
Timing of the second and third transfusion should be determined by a combination of weekly MCA-PSV and closing Hb drop calculation (0.9/d after 1 st and 0.6/d after 2 nd)**	52/78 (66.7)	53/70 (75.7)
Combining intraperitoneal and intravascular IUT should not be routinely used at the same procedure	72/78 (92.3)	
Target level in intravascular IUT		
Between 14-16 g/dl or hematocrit 40-45% regardless of GA	50/78 (64.1)	47 (68.1)
Between 16-18 g/dl or hematocrit 45-50% regardless of GA	28/78 (35.9)	
Maximal gestational age of last IUT		
32-34+6 weeks	9 (11.4)	
35-35+6weeks	49/78 (62.8)	55/70 (78.5)
36-37 weeks	20 (25.6)	
Threshold for emergency caesarean in pregnancies with viable GA that develop fetal bradycardia during IUT		
Trial of intrauterine resuscitation	47/78 (60.2)	63/70 (90)
Trial of intrauterine resuscitation + atropine first	24/78 (30.8)	
Trial of atropine only first	4/78 (5.1)	
Immediately progress into emergency C-Section	5/78 (6.4)	
Setting of IUT with pre-viable GA		
The Office/Fetal Medicine Unit	44/78 (56.4)	
OR/Theatres	34/78 (43.6)	
Phenobarbital should not be used	72/78 (92.3)	
In DCDA twins, fetal blood sampling should not be performed on both fetuses when only one has triggered the MCA Doppler MoM for IUT	67/78 (85.9)	

506 * Dark grey shading represents consensus (defined as $\geq 70\%$ agreement), light grey represents significant agreement
507 (of 60-69%), and white represents no agreement ($< 60\%$). A '-' in a cell means that the issue was not addressed in
508 that round.

509 **Approximately in 10-14 days after 1st IUT and in 2-3 weeks after the 2nd IUT

510 † Variability in open responses. See Supplementary material.

511 GA= Gestational age Hb=hemoglobin Hct=Hematocrit IVIG= Intravascular immunoglobulins IUT= intrauterine
512 transfusion MoM= Multiple of the Median MCA PSV= Middle cerebral artery peak systolic flow

513 All percentages are presented in parentheses ()

514
515

Journal Pre-proof

516 **Table 5: Delivery Management**

517

	Round in which item was included	
	1	2
	(N=78)	(N=77)
GA at which delivery should be opted as opposed to IUT if initial presentation of >1.5 MoM		
32 weeks	1/78 (1.3)	
34 weeks	11/78 (14.1)	
35 weeks	33/78 (42.3)	
36 weeks	23/78 (29.5)	
37 weeks	10/78 (12.8)	
Timing of delivery following last successful IUT in absence of other indications		
2-3 weeks after last IUT	54/78 (69.2)	69/77 (89.6)
3-4 weeks after last IUT	9/78 (11.5)	
Other	15/78 (19.2)	
GA for delivery in pregnancies where critical antibody titres are reached and IUT was not required		
34-36 +6/7 weeks GA	9/78 (11.5)	
37-38 + 6/7 weeks GA	54/78 (69.2)	77 (100.0)
39-40 + 6/7 weeks GA	15 (19.2)	
Route of delivery should follow obstetric indication for vaginal or CS delivery	76/77 (98.7)	
Delayed cord clamping during delivery in an IUT pregnancy	55/77 (71.4)	
Cord milking should not be performed in an IUT pregnancy	66/77 (85.7)	

518 * Dark grey shading represents consensus (defined as $\geq 70\%$ agreement), light grey light grey represents significant
519 agreement (of 60-69%), and white represents no agreement ($< 60\%$). A '-' in a cell means that the issue was not
520 addressed in that round.

521 GA= Gestational age, IUT= intrauterine transfusion CS= Caesarean section MoM= Multiple of the Median

522 All percentages are presented in parentheses ()

523

524 **Table 6: Postnatal Management**

	Round in which item was included	
	1	2
	(N=25)	(N=25)
Postnatal treatments to be considered for fetal anemia	N=25	
Phototherapy	25 (100.0)	
Blood transfusion	25 (100.0)	
Exchange transfusion	23 (92.0)	
IVIg	14 (56.0)	
Treatment with erythropoietin	7 (28.0)	
Bilirubin level at birth to initiate phototherapy treatment	N=25	
<5 mg/dl	4 (16.0)	
5-10 mg/dl	8 (32.0)	
10-15 mg/dl	2 (8.0)	
15-20 mg/dl	2 (8.0)	
None of the above	9 (36)	
Rise in bilirubin should be calculated when monitoring hyperbilirubinemia	18/25 (72.0)	
Rate of rise to start aggressive phototherapy in first 24 hrs		
>0.2 mg/dl	6 (33.3)	
>0.3 mg/dl	3 (16.7)	
>0.4 mg/dl	4 (22.2)	
>0.5 mg/dl	5 (27.8)	
None of the above	7 (28.0)	
All neonates with a history of HDFN requiring IUT should be not started on phototherapy immediately	15/25 (60.0)	15/25 (60.0)
Neonatal IVIG should not be used routinely in cases of HDFN with hyperbilirubinemia in the first few days of life to prevent the need of exchange transfusions	21/25 (84.0)	
Hb threshold for transfusion in term neonates with HDFN who are critically ill	N=25	
<13 g/dl	1 (4.0)	
<12 g/dl	9 (36.0)	
<11 g/dl	2 (8.0)	
<10 g/dl	6 (24.0)	
< 9 g/dl	1 (4.0)	
< 8 g/dl	2 (8.0)	
None of the above	4 (16.0)	
Hb threshold for transfusion in term neonates with HDFN who are not critically ill	N=25	

	Round in which item was included	
	1	2
	(N=25)	(N=25)
< 13 g/dl	1 (4.0)	
< 11 g/dl	2 (8.0)	
< 10 g/dl	8 (32.0)	
< 8 g/dl	5 (20.0)	
< 7 g/dl	4 (16.0)	
None of the above	5 (20.2)	
Hb threshold for transfusion in preterm neonates with HDFN who are critically ill	N=25	
<12 g/dl	13 (52.0)	
<10 g/dl	7 (28.0)	
<8 g/dl	1 (4.0)	
None of the above	4 (16)	
Hb threshold for transfusion in preterm neonates with HDFN who are not critically ill	N=25	
<12 g/dl	2 (8.0)	
<11 g/dl	2 (8.0)	
<10 g/dl	6 (24.0)	
<9 g/dl	2 (8.0)	
<8 g/dl	4 (16.0)	
<7 g/dl	2 (8.0)	
None of the above	7 (28.0)	
AAP 2022 guidance use to determine thresholds of phototherapy and exchange transfusion	18/25 (72.0)	
Prior to discharge, in a newborn, of a pregnancy complicated by maternal alloimmunisation with no evidence of fetal anemia, Hb and bilirubin levels should be determined	23/25 (92.0)	
In a newborn of a pregnancy which has critical titres but did not require IUT, anemia laboratory investigations should be performed	24/25 (96.0)	
In a newborn of a pregnancy which required recent IUT and who was not anaemic at birth, anaemia laboratory investigations should be repeated at 1 week	21/25 (84.0)	
Exchange transfusions should be centralized in hospitals with sufficient exposure and experience	24/25 (96.0)	
Erythropoietin stimulating agent not used in the first few weeks of life to reduce the need for transfusions in neonates treated with IUT	23/25 (92.0)	
The Hb level to consider further transfusions following hospital discharge is <7 g/dl	23/25 (92.0)	
Follow up frequency in infancy		
4-8 weeks	3/25 (12.0)	
2-3 months	15/25 (60.0)	24/25 (96.0)
2- 6 months	5/25 (20.0)	

	Round in which item was included	
	1	2
	(N=25)	(N=25)
6-12 months	2/25 (8.0)	

525 * Dark grey shading represents consensus (defined as $\geq 70\%$ agreement), light grey represents significant agreement
526 (of 60-69%), and white represents no agreement ($< 60\%$). A '-' in a cell means that the issue was not addressed in
527 that round.

528 ** 2 experts indicated that other erythropoietin analogs may be used in their respective countries

529 IVIg= Intravenous immunoglobulins Hb= hemoglobin Hct=Hematocrit AAP: American Academy of Pediatrics

530 All percentages are presented in parentheses ()

531

Journal Pre-proof

