




Review

# The Biological Role and Translational Implications of the Long Non-Coding RNA *GAS5* in Breast Cancer

Ilaria Grossi <sup>†</sup> , Eleonora Marchina <sup>†</sup>, Giuseppina De Petro  and Alessandro Salvi <sup>\*†</sup> 

Division of Biology and Genetics, Department of Molecular and Translational Medicine, University of Brescia, 25123 Brescia, Italy; ilaria.grossi@unibs.it (I.G.); eleonora.marchina@unibs.it (E.M.); giuseppina.depetro@unibs.it (G.D.P.)

\* Correspondence: alessandro.salvi@unibs.it

<sup>†</sup> These authors contributed equally to this work.

**Simple Summary:** *GAS5* is a lncRNA that was identified decades ago as a key player in arresting the growth of mouse fibroblasts. *GAS5* may fold into an RNA structure that competes with the intracellular glucocorticoid receptor for binding to DNA targets located on the promoters of apoptosis-associated genes, thus controlling their expression. *GAS5* expression has been examined in a variety of human cancers, and its levels have been found to be lower in cancer tissues than in adjacent non-tumor tissues. In breast cancer (BC), *GAS5* has mainly been described as tumor suppressor lncRNA capable of promoting apoptosis and inhibiting cell proliferation. Low expression of *GAS5* is correlated with poor prognosis, and its expression is modulated by certain chemotherapeutic agents and during the acquisition of drug resistance. Due to its role and expression trend, new experimental approaches to augment or restore the cellular levels of *GAS5* may have important translational implications in BC as well as in other malignancies.

**Abstract:** The lncRNA *GAS5* plays a significant role in tumorigenicity and progression of breast cancer (BC). In this review, we first summarize the role of *GAS5* in cell biology, focusing on its expression data in human normal tissues. We present data on *GAS5* expression in human BC tissues, highlighting its downregulation in all major BC classes. The main findings regarding the molecular mechanisms underlying *GAS5* dysregulation are discussed, including DNA hypermethylation of the CpG island located in the promoter region of the gene. We focused on the action of *GAS5* as a miRNA sponge, which is able to sequester microRNAs and modulate the expression levels of their mRNA targets, particularly those involved in cell invasion, apoptosis, and drug response. In the second part, we highlight the translational implications of *GAS5* in BC. We discuss the current knowledge on the role of *GAS5* as candidate prognostic factor, a responsive molecular therapeutic target, and a circulating biomarker in liquid biopsies with clinical importance in BC. The findings position *GAS5* as a promising druggable biomolecule and stimulate the development of strategies to restore its expression levels for novel therapeutic approaches that could benefit BC patients in the future.

**Keywords:** *GAS5*; breast cancer; lncRNAs; ncRNAs



**Citation:** Grossi, I.; Marchina, E.; De Petro, G.; Salvi, A. The Biological Role and Translational Implications of the Long Non-Coding RNA *GAS5* in Breast Cancer. *Cancers* **2023**, *15*, 3318. <https://doi.org/10.3390/cancers15133318>

Academic Editors: Sabrina Strano and Mario Cioco

Received: 30 May 2023

Revised: 20 June 2023

Accepted: 21 June 2023

Published: 23 June 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

### 1.1. Breast Cancer

Breast cancer (BC) is the most common malignancy in women worldwide. It is a heterogeneous disease promoted by different risk factors, including the exposition to endogenous and exogenous estrogens, lifestyle, excessive food and alcohol consumption, obesity, and toxic environmental factors (pollutions, heavy metals, and chemicals) [1]. Approximately 15% of breast cancers are due to genetic predisposition and hereditary factors. Pathogenetic mutations in *BRCA1* and *BRCA2* (Breast Cancer Associated gene 1 and 2) genes have been extensively described in the literature [2,3], as well as other

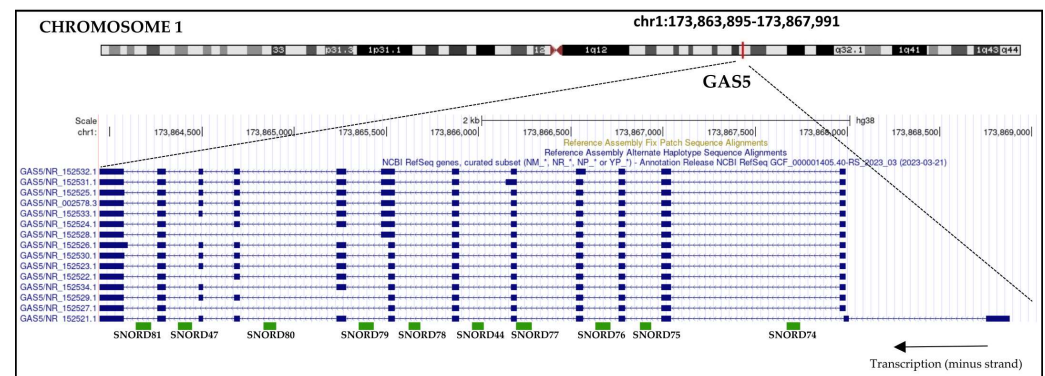
susceptibility genes with high or intermediate penetrance [4,5]. These genes are necessary for DNA repair, and their deactivation results in chromosomal instability that contributes to tumor transformation and carcinogenesis. The healthy mammary epithelium consists of the luminal cells (that produce milk) and basal cells that have contraction ability and favor milk secretion. BC is histologically classified into invasive ductal carcinoma (IDC), also known as invasive BC of no special type (NST), which accounts for 50% to 80% of all invasive cancers, and into “special subtypes” of BC. Special subtypes account for up to 25% of all BCs and include more than 17 distinct histological special types, with invasive lobular carcinoma (ILC) being the most common subtype. ILC accounts for approximately 10% of all invasive carcinomas [6,7]. BC is further classified based on the status of the intracellular estrogen receptor and progesterone receptor and the cell surface receptor HER2 (human epidermal growth factor receptor 2). On the basis of the expression profile, BC is further classified in five subtypes: (1) luminal A (approximately 50–60% of cases); (2) normal-like, which is a subtype of the luminal A group, but with worse prognosis (approximately 10% of the luminal A type); (3) luminal B (approximately 10% of cases); (4) HER2-enriched (approximately 20% of cases); and (5) basal-like triple-negative (TNBC) tumors (approximately 10% of cases). TNBC has a high risk of recurrence, with the worst prognosis among the different subtypes [8]. The curative options for BC are as follows: (a) surgical resection (75% of cases) of the tumor nodule/nodules to preserve the anatomy of the mammary gland when it is possible, or total removal (mastectomy); (b) radiotherapy used to destroy the remaining cancer cells after the surgery or to treat advanced BC (i.e., bone metastasis); (c) the use of hormone therapy for luminal BC with estrogen receptor antagonists in pre- and postmenopausal women; (d) targeted therapy such as the monoclonal antibody trastuzumab (Herceptin); and (e) treatment with inhibitors of CDK4/6 (cyclin-dependent kinase type 4 and 6) to inhibit cell cycle progression [9]. Following-up on patients with BC is essential for the best outcome, to monitor the response to therapies, and to verify the absence of relapse or metastasis in other organs such as the bones, lungs, liver, or brain.

### 1.2. Long Non-Coding RNAs

Long non-coding RNAs (lncRNAs) belong to the non-coding RNAs (ncRNAs) category. Based on their size, Mattick et al. recently suggested dividing the ncRNAs into three classes: (a) small RNAs less than 50 nucleotides in length (i.e., microRNAs and siRNAs); (b) ncRNAs generally 50–500 nucleotides in length, including RNA polymerase III (Pol III) transcripts such as tRNAs and 5S rRNA, small RNAs transcribed by RNA polymerase (Pol II) (i.e., small nuclear, snRNAs), and intron-derived small nucleolar RNAs (snoRNAs); (c) lncRNAs more than 500 nucleotides in length that are generally transcribed by Pol II [10]. GENCODE [11] and FANTOM consortium [12] identified ~20,000 and ~30,000 human lncRNA genes, respectively. More than 100,000 human lncRNAs transcripts have been annotated and catalogued in these databases. LncRNAs are transcribed by RNA polymerase I, Pol II, and Pol III, and are also processed from introns. With respect to protein-coding genes, lncRNAs can be classified into sense lncRNAs, antisense lncRNAs, bidirectional lncRNAs, intron lncRNAs, intergenic lncRNAs, and enhancer lncRNAs. The majority of lncRNAs are spliced and polyadenylated, but unlike mRNAs, they lack translational potential. While many lncRNAs are localized in the nucleus, a significant portion functions in cytoplasmic processes, and their expression is often cell and tissue-specific [13,14]. Numerous studies have described the involvement of lncRNAs in many physiological and pathological contexts. They participate in chromatin architecture and gene expression by regulating the transcription, RNA splicing, RNA stability, and protein translation [15]. LncRNAs play pivotal roles in numerous malignancies, and the aberrant expression of tumor-suppressive and/or oncogenic lncRNAs is frequently found in cancer cells. They are involved in carcinogenesis processes, such as proliferation, cell migration, invasion, epithelial–mesenchymal transition (EMT), and apoptosis [16,17].

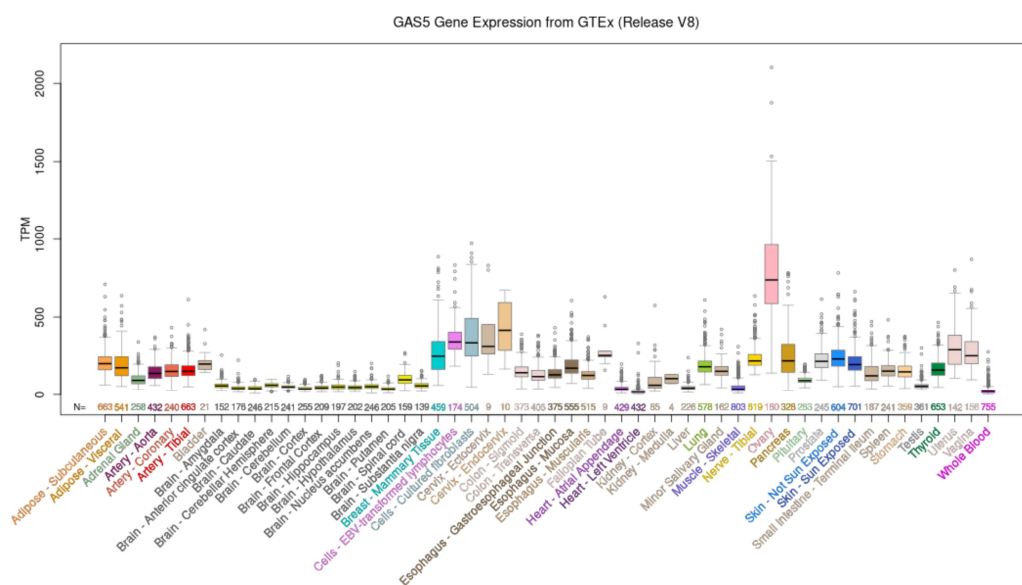
### 1.3. GAS5

The growth arrest-specific 5 (*GAS5*) gene encodes for a lncRNA that was identified using a subtraction library approach to clone cDNA from growth-arrested NIH 3T3 mouse fibroblasts cells [18]. The human *GAS5* gene is located at 1q25.1, and it encodes for more than 20 different transcript variants with lengths of 510–725 nucleotides. The *GAS5* gene includes up to 13 exons (transcript variant NR\_152521.1), as well as 10 small nucleolar RNA-coding sequences (snoRNA) in its introns (Figure 1).



**Figure 1.** Human *GAS5* gene locus, structure, and transcript variants. *GAS5* is a lncRNA encoded by a sequence located at 1q25.1. In The Genome Browser database, 15 different RNA isoforms have been annotated with lengths of 510–725 nucleotides. *GAS5* transcript isoforms include up to 13 exons. *GAS5* is a gene harboring 10 small nucleolar RNA genes (snoRNA) in its introns (SNORD genes). (From Genome Browser; <https://genome.ucsc.edu> modified, accessed on 2 May 2023).

The database LncBook 2.0 (integrating human long non-coding RNAs with multi-omics annotations) [19] provides conservation features of human lncRNA genes across 40 vertebrates. The conservation levels are expressed as Q50 = low conservation, Q75 = intermediate conservation, and Q90 = high conservation. According to the database, the level of *GAS5* conservation varies between Q50 and Q90 in apes (Q50 in *Gorilla gorilla gorilla*, Q75 in *Pan troglodytes*, and Q90 in *Nomascus leucogenys*). In rodents, *GAS5* shows Q50 conservation in *Mus musculus* and *Rattus norvegicus*, but Q90 in *Heterocephalus glaber*. In birds such as *Gallus gallus*, it exhibits Q50 conservation, while in the fish *Danio rerio* and the amphibian *Xenopus tropicalis*, it is not conserved. These data confirm an intermediate level of sequence conservation of *GAS5* across different species. Human *GAS5* is a well-studied lncRNA that widely contributes to various cell biology functions. *GAS5* acts as a decoy for a glucocorticoid receptor (GR) by mimicking the glucocorticoid response element (GRE) present on target DNA. The binding of *GAS5* with the GR blocks the binding between GR and GRE, inhibiting the transcription of genes modulated by GR that are involved in cell survival and apoptosis [20,21]. In addition, *GAS5* promotes osteoblast differentiation [22] and regulates the expression of insulin receptors in adipocytes [23]. According to the NIH Genotype–Tissue Expression (GTEx) project (<https://commonfund.nih.gov/GTEx>, accessed on 26 April 2023), *GAS5* is ubiquitously expressed in human tissues (Figure 2). Interestingly, high levels of *GAS5* expression were detected in female tissues and organs (breast, cervix, uterus, ovary), with the highest expression found in the ovary. This suggests its key role in the physiology and pathology of these anatomical districts.



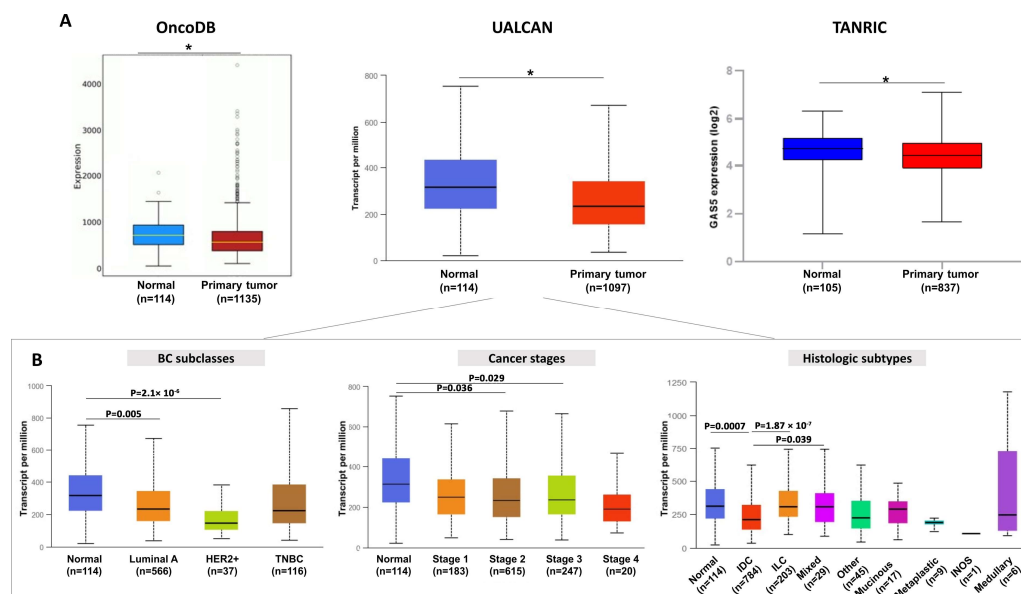
**Figure 2.** *GAS5* gene expression from GTEx (Release V8). The NIH Genotype–Tissue Expression (GTEx) project (<https://commonfund.nih.gov/GTEx>, accessed on 26 April 2023) was created to establish a sample and data resource for studies on gene expression in multiple human tissues. This track shows the median gene expression levels in 52 tissues and two cell lines (EBV-transformed lymphocytes and cultured fibroblasts) based on RNA-seq data from the GTEx final data release (V8, August 2019) obtained from UCSC genome browser (<https://genome.ucsc.edu>, accessed on 26 April 2023). This release is based on data from 17,382 tissue samples obtained from 948 adult post-mortem individuals.

## 2. *GAS5* Expression and Functions in BC

### 2.1. *GAS5* Expression Is Dysregulated in BC Tissues

A growing number of studies have suggested that lncRNAs are abnormally expressed in human cancer. The expression patterns of certain lncRNAs, both in solid biopsies and biological fluids, are highly cancer-specific and can be strongly associated with clinicopathological characteristics, including the overall survival of cancer patients. All these features address the potential of lncRNAs as promising tools useful in cancer diagnosis, disease monitoring, and prognosis [24,25]. Concerning *GAS5*, several studies have reported that its expression is typically reduced in different cancer types, including hepatocellular carcinoma (HCC) [26,27], head and neck squamous cell carcinoma, glioblastoma, bladder, and non-small cell lung cancer (NSCLC) [28]. In BC, *GAS5* is generally downregulated in solid biopsies, as indicated by the bioinformatics analysis that we performed by consulting the OncoDB [29], UALCAN [30], and TANRIC [31] databases (Figure 3A). These publicly available online databases provide analyses of RNA-seq data from TCGA by considering the breast-invasive carcinoma (BRCA) dataset. The downregulation of *GAS5* has been further observed among the major BC subclasses (Luminal A and HER2 positive) and in the predominant histological subtype (IDC, invasive ductal carcinoma) of the BRCA dataset (Figure 3B). Different published works have determined the *GAS5* levels in solid biopsies using qRT-PCR analysis and confirmed *GAS5* downregulation in tumor tissues from BC patients. Arshi et al. showed that *GAS5* expression in BCs is associated with the age of patients. *GAS5* levels were lower in the tissues of younger group (<45 years) compared with the older group (>45 years) [32]. Li et al. showed that *GAS5* levels were significantly decreased in HER2-positive BC specimens compared to corresponding non-tumor tissues ( $n = 20$ ) [33]. Furthermore, low levels of *GAS5* were significantly associated with advanced TNM stages and grading, as well as poor overall survival (OS) and disease-free survival (DFS) [33,34]. Interestingly, *GAS5* expression was found to be positively correlated with unc-51 like autophagy activating kinase (*ULK*) 1/2 mRNA in BC clinical samples [35]. The

over-expression of *GAS5* in BC cells up-regulated ULK1 and ULK2 protein expression, but not other autophagy-related proteins (Atgs), and led to autophagosome formation suggesting that the involvement of *GAS5* in autophagy should be further explored. Finally, *GAS5* was significantly downregulated in primary triple-negative BC (TNBC). In this BC type, the expression of *GAS5* was associated with tumor size, clinical stage, lymph node metastasis, and survival, suggesting that a high level of tissue *GAS5* could favor a good clinical outcome for BC patients [35–37]. Together, these results strongly support that the evaluation of *GAS5* expression in solid biopsies may have potential prognostic relevance in BC patients.

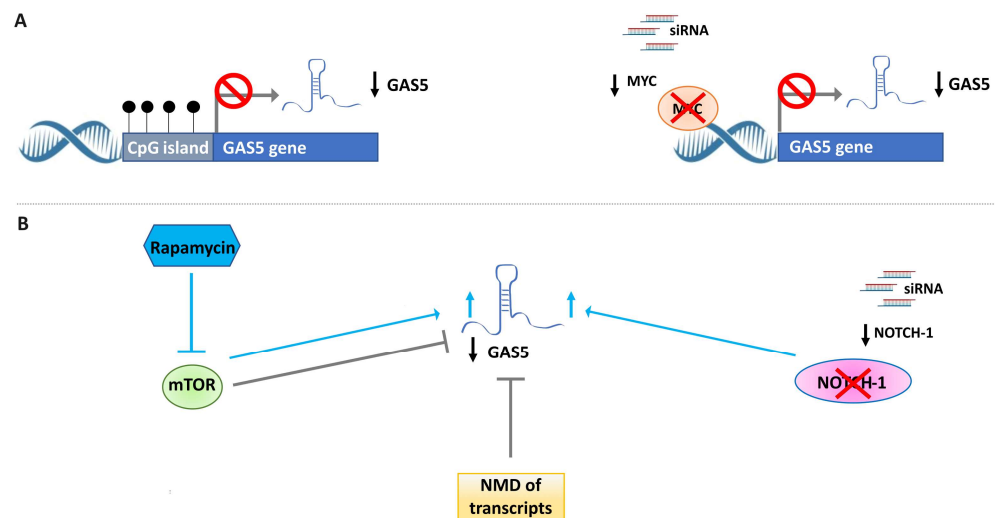


**Figure 3.** *GAS5* expression levels in TCGA datasets obtained by consulting different databases. (A) *GAS5* expression levels in primary BC tissues and normal tissues from TCGA datasets, evaluable in OncoDB, UALCAN, and TANRIC databases. (B) *GAS5* expression levels in the BRCA dataset from UALCAN based on BC subclasses, cancer stages, and histological subtypes. IDC, infiltrating ductal carcinoma; ILC, infiltrating lobular carcinoma; Mixed, mixed histology; INOS, infiltrating carcinoma not otherwise specified. \*  $p < 0.05$ .

## 2.2. Different Mechanisms Are Involved in the Downregulation of *GAS5* in BC

As for other lncRNAs, the expression of *GAS5* can be regulated by different mechanisms at the transcriptional or post-transcriptional level (Figure 4). Among these, DNA methylation that occurs in the CpG islands located in the promoter region of the gene can affect the expression of the lncRNA. As indicated in UCSC Human Genome Browser (GRCh38/hg38; consulted in March 2023) [38], the *GAS5* gene is located on human chromosome 1q25.1 (chr1:173,858,997–173,867,989), and a CpG island has been identified upstream of the *GAS5* gene (chr1:173,868,035–173,868,779). Li et al. evaluated the DNA methylation of *GAS5* in TNBC tissues and cells using a methylation-specific PCR (MSP) assay [39]. They showed a significant increase in DNA methylation levels in TNBC tissues versus adjacent normal tissues. Regarding BC cell lines, they found that *GAS5* was methylated and poorly expressed in TNBC cell lines (MDA-MB-231, MDA-MB-468, HCC1937, and MDA-MB-453), but partially demethylated and up-regulated in non-malignant human breast epithelial cells, MCF10A. Treating TNBC cells with the demethylating agent 5-Aza-2'-deoxycytidine (5-aza) was shown to determine the decrease in DNA methylation levels and the restoration of *GAS5* levels [39]. These data, also obtained in other cancers (such as melanoma [40] and cervical cancer [41]), support the finding that aberrant DNA methylation is one of the mechanisms involved in the downregulation of *GAS5*. At the post-transcriptional level, the expression of *GAS5* is modulated by the interplay between the

mTOR and the nonsense-mediated decay (NMD) pathways. The mTOR pathway takes part in the regulation of 5' TOP-containing mRNAs through its downstream effectors (p70S6K and eIF4E/4E-BP1) [42]. The NMD pathway is involved in the degradation of transcripts containing stop codons in early exons. As reported in the literature, *GAS5* is considered a 5'-terminal oligopyrimidine tract (5'-TOP) gene [43], and is characterized by a short reading frame terminating with a stop codon within exon 3 (of 12) that is recognized by the NMD pathway. In actively growing cells, the activation of the mTOR signaling pathway regulates *GAS5* expression [44]. In support of this, the inhibition of mTOR activity by rapamycin determines the increase in cellular *GAS5* in different cell types, including BC cells [45,46]. Two additional well-known oncogenic pathways, Notch-1 and MYC, seem to be involved in the regulation of *GAS5* in BC cells. Pei et al. reported that the silencing of Notch-1 using siRNAs significantly increased the level of *GAS5* in the T47D cell line [47]. Similarly, the lentiviral vector-mediated siRNA knockdown of MYC led to a decrease in *GAS5* expression levels in MCF7 cells [48]. Thus, *GAS5* expression may be regulated depending on the modulation of Notch-1 and MYC levels. Further in vitro and in vivo studies are required to elucidate the type of regulation, direct or indirect, exerted by Notch-1 and MYC on the expression of *GAS5* in BC.



**Figure 4.** Schematic representation of mechanisms involved in the regulation of *GAS5* expression in BC. (A) *GAS5* expression can be downregulated by DNA methylation (left). MYC silencing (right) induced by siRNA molecules can decrease *GAS5* expression. Black circles indicate methylated cytosines. (B) Different pathways, including mTOR, NMD, and NOTCH-1, can be involved in the post-transcriptional regulation of *GAS5* in BC. Nonsense mediated decay, NMD.

### 2.3. *GAS5* Functions as ceRNA of miRNAs in BC

The main biological function described in BC for *GAS5* is as competitive endogenous RNA (ceRNA). ceRNAs are ncRNAs which competitively bind microRNAs (miRNAs) and sequester miRNAs from their original target transcripts. Consequently, they avoid the degradation or expression inhibition of target transcripts induced by miRNAs at the post-transcriptional and translational levels. As in other cancers, *GAS5* can interact with different miRNAs and thus weaken the effect of miRNAs on mRNA targets. At the time of writing this review, we found five studies on BC reporting that *GAS5* acts as a ceRNA for four miRNAs with oncogenic functions (onco-miRs), including miR-21-5p, miR-221-3p, miR-196a-5p, and miR-378-5p [36,37,49,50], and the tumor-suppressor miR-216b [51]. miR-21-5p is the most commonly upregulated miRNA in solid and hematological malignancies and exerts its function by regulating tumor suppressor genes, like PTEN, TPM1 and PDC4. Zhang et al. verified the interaction between *GAS5* and miR-21-5p using RNA immunoprecipitation and RNA-pulldown assays [49]. Furthermore, in vitro experiments demonstrated the down and upregulation of *GAS5* after transfection with

miR-21-5p mimics and anti-miR-21-5p, respectively. These data suggested the reciprocal regulation of miR-21-5p and GAS5 in BC cells. In triple-negative BC cells, GAS5 has been shown to act as a ceRNA of miR-196a-5p, which targets FOXO1 [36]. The upregulation of GAS5 in TNBC cells (i) increases FOXO1 expression, (ii) inhibits downstream PI3K/AKT activation, and (iii) decreases cell invasion. The over-expression of miR-196a-5p partially rescued the effects induced by the ectopic expression of GAS5 in TNBC cells. In the same cell lines, Zheng et al. reported that GAS5 promoted apoptosis by acting as a ceRNA of miR-378-5p which negatively regulated SUFU [37]. It is known that SUFU functions as a negative regulator of the Hedgehog signaling pathway and has a key role in promoting apoptosis. In BC cells resistant to adriamycin treatment, direct binding between GAS5 and miR-221-3p was verified using a luciferase reporter assay and an RNA immunoprecipitation assay [50]. GAS5 competed for miR-221-3p binding to regulate the expression of its target, dickkopf 2 (DKK2). The role of the GAS5/miR-221-3p/DKK2 axis in adriamycin resistance is explained in detail below. Finally, GAS5 may also interact with miR-216b in BC cells, as suggested by the results of the dual-luciferase assay. In the same work, the authors showed that the inhibition of GAS5 using siRNA determined (i) the decrease in the invasion ability of BC cells, and (ii) inhibition of E-cadherin, as well as the up-regulation of N-cadherin and Vimentin in cervical cancer cells (SiHa cells) [51]. These data suggest that GAS5 may be involved in epithelial–mesenchymal transition (EMT) in cancer, and further studies will be fundamental in determining the role of GAS5 in metastasis formation in BC.

In addition to its ceRNA role, GAS5 has been recently described as mitochondria-associated lncRNA. Sang et al. reported that GAS5 localizes in mitochondria, where it modulates tricarboxylic acid (TCA) flux by suppressing the formation of the metabolic enzymes complex formed by canonical members of the TCA cycle (fumarate hydratase, FH; malate dehydrogenase, MDH2; citrate synthase, CS) [34]. In BC cells, GAS5 levels are decreased by glucose deprivation. A decreased availability of mitochondria-associated GAS5 results in the formation of the CS-MDH2-FH complex, promoting the TCA cycle. These data indicate an important functional role for GAS5 as a regulator of metabolism in BC cells.

### 3. GAS5 as a Potential Biomarker and Therapeutic Target in BC

#### 3.1. GAS5 Levels Modulate the Response to Apoptosis-Inducing Agents in BC Cells

In breast and other cancer cell lines, GAS5 plays an important role in affecting cell growth and determining apoptosis [52]. As previously described, in BC cells, GAS5 sponged miR-21 [49] and miR-196a-5p [36], determining the upregulation of the tumor suppressors PTEN, PDCD4, and FOXO1, which are key factors in the induction of apoptosis. By considering the close relationship between the activation of apoptosis and cancer therapy (i.e., radiotherapy and chemotherapy), the intracellular levels of GAS5 may have important implications for cancer therapy responses due to its apoptosis-promoting activity. Consistent with this observation, Pickard et al. demonstrated that low GAS5 levels attenuated the effects of chemotherapeutic agents (5-FU and docetaxel) and UV-C irradiation in terms of apoptosis induction, cell viability, and colony formation ability in *in vitro* BC models [46]. Particularly, reductions of 50–70% in GAS5 levels in MCF7 and T-47D cells were strongly associated with attenuated cell death in response to treatment with 5-FU, docetaxel, or UV-C irradiation [46]. Conversely, MCF-7 cells transfected with the GAS5-overexpressing vector pCMVSPORT6 were more sensitive to UV irradiation and cisplatin-induced apoptosis [53]. However, it has been observed that, in the same cell line, the effect on apoptosis caused by TKI imatinib is independent of the cellular GAS5 level. Taken together, these data suggest that GAS5 is able to modulate the action of anti-cancer treatments in a selective way, likely due to the mechanisms of action of the different drugs. These findings could have important implications from a therapeutic point of view. Indeed, the increase in intracellular levels of GAS5 before the administration of chemo- or radio-based therapies may represent an innovative approach to improve the action of the treatments. Importantly, this goal could be achieved by acting on the mechanisms involved in GAS5 regulation, such

as DNA methylation and/or the mTOR pathway. As previously described, the demethylating agents, as well as the mTOR inhibitors, restore *GAS5* levels in BC cells. Therefore, functional studies should be addressed to prove the efficacy of innovative anti-cancer tools based on the combination of epigenetic drugs (i.e., the demethylating agent 5-aza) or mTOR inhibitors (i.e., rapalogues) with conventional treatments.

### 3.2. *GAS5* Levels Are Modulated by Different Anticancer Drugs

*GAS5* expression levels are expected to impact the treatment response of BC cells. However, relevant studies have reported that different anti-cancer drugs alter the *GAS5* levels of BC cells. Among these, treatment with TKI sorafenib results in a significant increase in *GAS5* levels in HCC-1937 and MCF-7 cells. The authors hypothesized that the increased *GAS5* levels may be an indirect downstream effect of the sorafenib treatment [26]. Jiang et al. demonstrated that metformin, a lipophilic biguanide with discussed antitumor effects, increases *GAS5* levels in BC cells. The authors demonstrated that the levels of mTOR, phospho-m-TOR, and mitogen-activated kinase p-P70S6K proteins decreased after metformin treatment of breast cancer cells resistant to tamoxifen. The upregulation of *GAS5* resulted in growth inhibition of BC cells and increased apoptosis [54]. Similarly, natural compounds with antitumor properties have been found to modulate *GAS5* levels, including curcumin, a chemo-protective natural agent extracted from the rhizome of *Curcuma longa*. The exposure of three BC cell lines to dendrosomic-delivered curcumin significantly affected the increase in *GAS5* expression as well as certain anti-cancer effects such as a decrease in cell viability, migration, and apoptosis induction. Interestingly, knocking down *GAS5* using siRNAs limited the effects on cell proliferation, apoptosis, and migration, suggesting that it might play a role in the anticancer action of curcumin in BC [55].

### 3.3. *GAS5* Has Regulatory Functions in Drug Resistance of BC

Drug resistance remains a clinical challenge in the treatment of BC. An understanding of the mechanisms involved in the development of resistance is urgently needed to reduce the side effects of treatments and identify new therapeutic strategies for improving the outcomes of BC patients. Numerous studies have shown that lncRNAs can be considered important players in the regulation of drug resistance [56]. *GAS5* has been implicated in the activation of drug resistance through the regulation of different pathways. One of these pathways involves the membrane-associated protein ABCB1, a member of the superfamily of ATP-binding cassette (ABC) transporters that is responsible for decreased drug accumulation in multidrug-resistant cells and often mediates the development of anticancer drug resistance. Using RNA-seq technology, Chen et al. identified *GAS5* as one of the most downregulated lncRNAs in BC MCF7 cells resistant to adriamycin (ADR), an anthracycline widely used as a chemotherapeutic drug for BC. The over-expression of *GAS5* significantly increased the sensitivity of resistant cells to ADR by reducing cell proliferation capacity and increasing apoptosis [50]. Moreover, the upregulation of *GAS5* affected both the efflux function of ABCB1 and its expression. Mechanistically, the authors showed that *GAS5* acts as a ceRNA by competing for miR-221-3p binding to regulate its target DKK2, which is a critical receptor for activating the Wnt/ $\beta$ -catenin signaling pathway. Importantly, in vivo experiments have confirmed that the upregulation of *GAS5* reduces miR-221-3p and ABCB1 expression, increases DKK2 expression, and confirms the ability of *GAS5* to sensitize resistant cells to ADR. These important data suggest that *GAS5* could revert ABCB1-mediated ADR resistance in ER-positive BC cells (MCF7) via the miR-221-3p/DKK2 axis by repressing the Wnt/ $\beta$ -catenin pathway. The role of lncRNA *GAS5* in increasing tamoxifen sensitivity has been demonstrated in MCF-7 resistant cells and in vivo in nude mice. *GAS5* may act as a ceRNA for miR-222, resulting in increased levels of the miRNA target PTEN. Knocking down miR-222 in tamoxifen-resistant MCF7 cells increased levels of *GAS5*, leading to higher sensitivity to the drug [57]. Similarly, *GAS5* overexpression also promoted sensitivity to ADR in triple-negative BC cells (MDA-MB-231 and MDA-MB-468) [39]. Approximately 13–15% of BCs are characterized by an amplification of ERBB2 that



results in the activation of the HER2 pathway [1]. The epidermal growth factor 2 (ERBB2, formerly HER2 or HER2/neu) is a transmembrane receptor tyrosine kinase in the epidermal growth factor receptor family that correlates with a higher death rate in the absence of systemic therapy. Patients with HER2-positive BC benefit from HER2-targeted therapy, including anti-HER2 antibodies (such as trastuzumab) and TKIs (such as lapatinib) [58]. Li et al. identified *GAS5* as one of the most downregulated lncRNAs in HER2-overexpressing BC SKBR-3 cells made resistant to trastuzumab (SKBR-3/Tr). Since *GAS5* functions as a ceRNA for miR-21, and miR-21 promotes tumor proliferation and invasion by targeting PTEN, the low levels of *GAS5* in SKBR-3/Tr cells are related to low PTEN levels, favoring the aggressive properties of cancer cells. Interestingly, treatment with the TKI lapatinib suppressed the proliferation of SKBR-3/Tr cells by upregulating PTEN and *GAS5* [33].

### 3.4. *GAS5* May Sensitize BC Cells to Radiation Therapy

Radiation therapy is an integrative component of BC treatment [1]. In early BC, radiation therapy following surgery reduces locoregional recurrences and improves patient survival. In advanced BC, radiation therapy plays a crucial role in alleviating symptoms caused by metastasis, including those affecting the brain and bones [59]. However, cancer cells can become radioresistant, limiting the clinical benefit of this therapeutic approach. The role of *GAS5* in the radiation response shows promise. As mentioned above, *GAS5* was initially shown to impair the effects induced by UV-C irradiation, which was used as an in vitro apoptotic stimulus for BC cells [46,53]. More recently, Ma et al. reported that ionizing radiation inhibited *GAS5* expression in BC cells. Interestingly, the overexpression of *GAS5* in irradiated BC cells promoted apoptosis and significantly increased unrepaired DNA damage, which is one of the direct effects induced by ionizing radiation, leading to cell cycle arrest [60]. The authors also found that the upregulation of miR-21, known to be sponged by *GAS5*, reversed the effects of *GAS5* on the radiosensitivity of BC cells. These data suggest the potential role of *GAS5* in sensitizing BC cells to ionizing radiation through miR-21 and support further in vivo experiments to explore this function.

### 3.5. *GAS5* Levels in Liquid Biopsies from BC Patients

Relevant evidence has indicated that the *GAS5* transcript is stable and detectable in biological fluids (i.e., plasma and serum). Many studies have evaluated the circulating levels of *GAS5* in plasma or serum from cancer patients and the promising results obtained in this field make circulating *GAS5* a potentially interesting non-invasive tool for cancer diagnosis, prognosis, and treatment response. In NSCLC and HCC, low levels of *GAS5* were found in plasma from cancer patients compared with healthy controls [26,61]. Interestingly, *GAS5* was also detected in exosomes isolated from the serum of NSCLC patients, displayed a significantly low level compared with healthy controls, and correlated with tumor size and TNM stage [62]. The circulating levels of *GAS5* in patients with solid cancers can be modulated in response to different anti-cancer therapies, including chemotherapy [63,64] and TKI [65]. Concerning BC, sparse but promising data have been published regarding the circulating levels of *GAS5*. Han et al. reported no significant differences in plasmatic levels of *GAS5* between BC patients and the healthy individuals. However, they found that *GAS5* plasma levels were significantly decreased at the postoperative stage compared with the preoperative stage in surgery-treated BC patients. A negative correlation was further shown between lymph node metastasis states and plasmatic *GAS5* levels after surgery [66]. Toraih et al. reported that *GAS5* was downregulated in the serum of BC patients compared with healthy subjects and non-cancer patients at risk of developing BC. Interestingly, lower *GAS5* levels in the serum of BC patients were related to metastasis and recurrence [67]. These data serve as a starting point for future research aiming to confirm the clinical significance of circulating *GAS5* as a biomarker for BC. The availability of larger cohorts of patients, as well as the use of standardized and sensitive protocols for lncRNA analysis (such as droplet-digital PCR [68]), should be considered to achieve this goal.

#### 4. Conclusions

The lncRNA *GAS5* was discovered decades ago and has been predominantly studied in human cancer, particularly in breast cancer, where its downregulation is correlated with unfavorable clinical outcomes. Limited information is available on the role of human *GAS5* in cell biology and development, and to our knowledge, there is currently no characterization of its high expression in normal human female organs (breast, ovary, cervix and uterus) at a single-cell resolution in a spatial context. This lack of basic knowledge does not allow for a deep understanding of the role of *GAS5* in normal breast tissue and in BC. Nevertheless, altered expression of *GAS5* has been reported in diseases other than cancer, including neurodegenerative disorders (i.e., Alzheimer's [69]), autoimmune diseases (i.e., rheumatoid arthritis [70]), knee osteoarthritis [71], and preeclampsia [72]. Concerning BC, the extensive knowledge of *GAS5* expression in all subtypes of BC tissues highlights real translational implications of the data reported in the literature thus far. Given the advancements in RNA-based biotechnology, it is worth further exploring the translational potential of this promising RNA molecule. Several factors contribute to the importance of *GAS5* in breast cancer research: (i) *GAS5* downregulation is associated with an unfavorable clinical outcome of BC patients; (ii) altered *GAS5* levels have been detected following anti-cancer therapies, including chemotherapy and radiotherapy; and (iii) *GAS5* plays a role in sensitizing cancer cells to different types of treatment, promoting apoptosis, and in inhibiting the proliferation of cancer cells. As a tumor-suppressive transcript, therapeutic approaches aimed at augmenting *GAS5* expression in BC may be crucial in the development of novel drugs. However, it is important to address key challenges associated with ncRNA therapeutics, such as specificity, delivery, and tolerability [73].

**Author Contributions:** Conceptualization, A.S.; writing—original draft preparation, I.G., A.S., E.M. and G.D.P.; writing—review and editing, I.G., A.S., E.M. and G.D.P.; supervision, A.S.; funding acquisition, A.S., E.M. and G.D.P. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported by Lega Italiana Lotta ai Tumori (LILT) grant number 02/2023, Liberamente Association (Gambara, BS, Italy) grant number 03/2023, and the University of Brescia (local grants) grant numbers 60/2021 and 60/2022. The funding bodies played no role in the design of the study, the collection, analysis, and interpretation of data, or the writing of the manuscript.

**Data Availability Statement:** The data generated in this study can be found in the Genome Browser, (<https://genome.ucsc.edu>, accessed on 2 May 2023), OncoDB (<https://oncodb.org/>, accessed on 26 April 2023), UALCAN (<https://ualcan.path.uab.edu>, accessed on 26 April 2023), and Tanric <https://bioinformatics.mdanderson.org/public-software/tanric/>, accessed on 26 April 2023) databases.

**Conflicts of Interest:** The authors declare no conflict of interest.

#### References

1. Harbeck, N.; Penault-Llorca, F.; Cortes, J.; Gnant, M.; Houssami, N.; Poortmans, P.; Ruddy, K.; Tsang, J.; Cardoso, F. Breast cancer. *Nat. Rev. Dis. Prim.* **2019**, *5*, 66. [[CrossRef](#)] [[PubMed](#)]
2. Marchina, E.; Fontana, M.G.; Speziani, M.; Salvi, A.; Ricca, G.; Di Lorenzo, D.; Gervasi, M.; Caimi, L.; Barlati, S. BRCA1 and BRCA2 genetic test in high risk patients and families: Counselling and management. *Oncol. Rep.* **2010**, *24*, 1661–1667. [[CrossRef](#)] [[PubMed](#)]
3. Abeni, E.; Grossi, I.; Marchina, E.; Coniglio, A.; Incardona, P.; Cavalli, P.; Zorzi, F.; Chiordera, P.; Paties, C.; Crosatti, M.; et al. DNA methylation variations in familial female and male breast cancer. *Oncol. Lett.* **2021**, *21*, 10–25. [[CrossRef](#)] [[PubMed](#)]
4. Bogdanova, N.; Helbig, S.; Dörk, T. Hereditary breast cancer: Ever more pieces to the polygenic puzzle. *Hered. Cancer Clin. Pract.* **2013**, *11*, 12. [[CrossRef](#)]
5. Yadav, S.; Boddicker, N.J.; Na, J.; Polley, E.C.; Hu, C.; Hart, S.N.; Gnanaolivu, R.D.; Larson, N.; Holtegaard, S.; Huang, H.; et al. Contralateral Breast Cancer Risk Among Carriers of Germline Pathogenic Variants in ATM, BRCA1, BRCA2, CHEK2, and PALB2. *J. Clin. Oncol.* **2023**, *41*, 1703–1713. [[CrossRef](#)]
6. Weigelt, B.; Geyer, F.C.; Reis-Filho, J.S. Histological types of breast cancer: How special are they? *Mol. Oncol.* **2010**, *4*, 192–208. [[CrossRef](#)] [[PubMed](#)]

7. Ellis, I.O.; Galea, M.; Broughton, N.; Locker, A.; Blamey, R.W.; Elston, C.W. Pathological prognostic factors in breast cancer. II. Histological type. Relationship with survival in a large study with long-term follow-up. *Histopathology* **1992**, *20*, 479–489. [[CrossRef](#)]
8. Koboldt, D.C.; Fulton, R.S.; McLellan, M.D.; Schmidt, H.; Kalicki-Veizer, J.; McMichael, J.F.; Fulton, L.L.; Dooling, D.J.; Ding, L.; Mardis, E.R.; et al. Comprehensive molecular portraits of human breast tumours. *Nature* **2012**, *490*, 61–70.
9. Loibl, S.; Poortmans, P.; Morrow, M.; Denkert, C.; Curigliano, G. Breast cancer. *Lancet* **2021**, *397*, 1750–1769. [[CrossRef](#)]
10. Mattick, J.S.; Amaral, P.P.; Carninci, P.; Carpenter, S.; Chang, H.Y.; Chen, L.L.; Chen, R.; Dean, C.; Dinger, M.E.; Fitzgerald, K.A.; et al. Long non-coding RNAs: Definitions, functions, challenges and recommendations. *Nat. Rev. Mol. Cell Biol.* **2023**, *17*, 430–447. [[CrossRef](#)]
11. Frankish, A.; Diekhans, M.; Jungreis, I.; Lagarde, J.; Loveland, J.E.; Mudge, J.M.; Sisu, C.; Wright, J.C.; Armstrong, J.; Barnes, I.; et al. GENCODE 2021. *Nucleic Acids Res.* **2021**, *49*, 917. [[CrossRef](#)] [[PubMed](#)]
12. Hon, C.-C.; Ramilowski, J.A.; Harshbarger, J.; Bertin, N.; Rackham, O.J.L.; Gough, J.; Denisenko, E.; Schmeier, S.; Poulsen, T.M.; Severin, J.; et al. An atlas of human long non-coding RNAs with accurate 5' ends. *Nature* **2017**, *543*, 199–204. [[CrossRef](#)] [[PubMed](#)]
13. Marchese, F.P.; Raimondi, I.; Huarte, M. The multidimensional mechanisms of long noncoding RNA function. *Genome Biol.* **2017**, *18*, 206. [[CrossRef](#)]
14. Bhat, S.A.; Ahmad, S.M.; Mumtaz, P.T.; Malik, A.A.; Dar, M.A.; Urwat, U.; Shah, R.A.; Ganai, N.A. Long non-coding RNAs: Mechanism of action and functional utility. *Non-Coding RNA Res.* **2016**, *1*, 43–50. [[CrossRef](#)] [[PubMed](#)]
15. Statello, L.; Guo, C.J.; Chen, L.L.; Huarte, M. Gene regulation by long non-coding RNAs and its biological functions. *Nat. Rev. Mol. Cell Biol.* **2021**, *22*, 96–118. [[CrossRef](#)]
16. Gao, N.; Li, Y.; Li, J.; Gao, Z.; Yang, Z.; Li, Y.; Liu, H.; Fan, T. Long Non-Coding RNAs: The Regulatory Mechanisms, Research Strategies, and Future Directions in Cancers. *Front. Oncol.* **2020**, *10*, 598817. [[CrossRef](#)]
17. Fang, Y.; Fullwood, M.J. Roles, Functions, and Mechanisms of Long Non-coding RNAs in Cancer. *Genom. Proteom. Bioinform.* **2016**, *14*, 42–54. [[CrossRef](#)]
18. Schneider, C.; King, R.M.; Philipson, L. Genes specifically expressed at growth arrest of mammalian cells. *Cell* **1988**, *54*, 787–793. [[CrossRef](#)]
19. Li, Z.; Liu, L.; Feng, C.; Qin, Y.; Xiao, J.; Zhang, Z.; Ma, L. LncBook 2.0: Integrating human long non-coding RNAs with multi-omics annotations. *Nucleic Acids Res.* **2023**, *51*, D186–D191. [[CrossRef](#)]
20. Zhou, Y.; Chen, B. GAS5-mediated regulation of cell signaling. *Mol. Med. Rep.* **2020**, *22*, 3049–3056. [[CrossRef](#)]
21. Goustin, A.; Thepsuwan, P.; Kosir, M.; Lipovich, L. The Growth-Arrest-Specific (GAS)-5 Long Non-Coding RNA: A Fascinating lncRNA Widely Expressed in Cancers. *Non-Coding RNA* **2019**, *5*, 46. [[CrossRef](#)] [[PubMed](#)]
22. Li, M.; Xie, Z.; Li, J.; Lin, J.; Zheng, G.; Liu, W.; Tang, S.; Cen, S.; Ye, G.; Li, Z.; et al. Gas5 protects against osteoporosis by targeting upf1/smad7 axis in osteoblast differentiation. *Elife* **2020**, *9*, e59079. [[CrossRef](#)] [[PubMed](#)]
23. Shi, Y.; Parag, S.; Patel, R.; Lui, A.; Murr, M.; Cai, J.; Patel, N.A. Stabilization of lncRNA GAS5 by a Small Molecule and Its Implications in Diabetic Adipocytes. *Cell Chem. Biol.* **2019**, *26*, 319–330.e6. [[CrossRef](#)] [[PubMed](#)]
24. Silva, A.; Bullock, M.; Calin, G. The Clinical Relevance of Long Non-Coding RNAs in Cancer. *Cancers* **2015**, *7*, 2169–2182. [[CrossRef](#)]
25. Qian, Y.; Shi, L.; Luo, Z. Long Non-coding RNAs in Cancer: Implications for Diagnosis, Prognosis, and Therapy. *Front. Med.* **2020**, *7*, 902. [[CrossRef](#)]
26. Faranda, T.; Grossi, I.; Manganelli, M.; Marchina, E.; Baiocchi, G.; Portolani, N.; Crosatti, M.; De Petro, G.; Salvi, A. Differential expression profiling of long non-coding RNA GAS5 and miR-126-3p in human cancer cells in response to sorafenib. *Sci. Rep.* **2019**, *9*, 9118. [[CrossRef](#)]
27. Chang, L.; Li, C.; Lan, T.; Wu, L.; Yuan, Y.; Liu, Q.; Liu, Z. Decreased expression of long non-coding RNA GAS5 indicates a poor prognosis and promotes cell proliferation and invasion in hepatocellular carcinoma by regulating vimentin. *Mol. Med. Rep.* **2016**, *13*, 1541–1550. [[CrossRef](#)]
28. Pickard, M.R.; Williams, G.T. Molecular and cellular mechanisms of action of tumour suppressor GAS5 lncRNA. *Genes* **2015**, *6*, 484–499. [[CrossRef](#)]
29. Tang, G.; Cho, M.; Wang, X. OncoDB: An interactive online database for analysis of gene expression and viral infection in cancer. *Nucleic Acids Res.* **2022**, *50*, D1334–D1339. [[CrossRef](#)]
30. Chandrashekar, D.S.; Karthikeyan, S.K.; Korla, P.K.; Patel, H.; Shovon, A.R.; Athar, M.; Netto, G.J.; Qin, Z.S.; Kumar, S.; Manne, U.; et al. UALCAN: An update to the integrated cancer data analysis platform. *Neoplasia* **2022**, *25*, 18–27. [[CrossRef](#)]
31. Li, J.; Han, L.; Roebuck, P.; Diao, L.; Liu, L.; Yuan, Y.; Weinstein, J.N.; Liang, H. TANRIC: An interactive open platform to explore the function of lncRNAs in cancer. *Cancer Res.* **2015**, *75*, 3728–3737. [[CrossRef](#)] [[PubMed](#)]
32. Arshi, A.; Sharifi, F.S.; Khorramian Ghahfarokhi, M.; Faghhi, Z.; Doosti, A.; Ostovari, S.; Mahmoudi Maymand, E.; Ghahramani Seno, M.M. Expression Analysis of MALAT1, GAS5, SRA, and NEAT1 lncRNAs in Breast Cancer Tissues from Young Women and Women over 45 Years of Age. *Mol. Ther. Nucleic Acids* **2018**, *12*, 751–757. [[CrossRef](#)] [[PubMed](#)]
33. Li, W.; Zhai, L.; Wang, H.; Liu, C.; Zhang, J.; Chen, W.; Wei, Q. Downregulation of lncRNA GAS5 causes trastuzumab resistance in breast cancer. *Oncotarget* **2016**, *7*, 27778–27786. [[CrossRef](#)] [[PubMed](#)]
34. Sang, L.; Ju, H.; Yang, Z.; Ge, Q.; Zhang, Z.; Liu, F.; Yang, L.; Gong, H.; Shi, C.; Qu, L.; et al. Mitochondrial long non-coding RNA GAS5 tunes TCA metabolism in response to nutrient stress. *Nat. Metab.* **2021**, *3*, 90–106. [[CrossRef](#)] [[PubMed](#)]

35. Li, G.; Qian, L.; Tang, X.; Chen, Y.; Zhao, Z.; Zhang, C. Long non-coding RNA growth arrest-specific 5 (GAS5) acts as a tumor suppressor by promoting autophagy in breast cancer. *Mol. Med. Rep.* **2020**, *22*, 2460–2468. [[CrossRef](#)]
36. Li, S.; Zhou, J.; Wang, Z.; Wang, P.; Gao, X.; Wang, Y. Long noncoding RNA GAS5 suppresses triple negative breast cancer progression through inhibition of proliferation and invasion by competitively binding miR-196a-5p. *Biomed. Pharmacother.* **2018**, *104*, 451–457. [[CrossRef](#)]
37. Zheng, S.; Li, M.; Miao, K.; Xu, H. lncRNA GAS5-promoted apoptosis in triple-negative breast cancer by targeting miR-378a-5p/SUFU signaling. *J. Cell. Biochem.* **2020**, *121*, 2225–2235. [[CrossRef](#)]
38. Kent, W.J.; Sugnet, C.W.; Furey, T.S.; Roskin, K.M.; Pringle, T.H.; Zahler, A.M.; Haussler, D. The Human Genome Browser at UCSC. *Genome Res.* **2002**, *12*, 996–1006. [[CrossRef](#)]
39. Li, J.; Li, L.; Yuan, H.; Huang, X.W.; Xiang, T.; Dai, S. Up-regulated lncRNA GAS5 promotes chemosensitivity and apoptosis of triple-negative breast cancer cells. *Cell Cycle* **2019**, *18*, 1965–1975. [[CrossRef](#)]
40. Zhang, Y.J.; Xie, R.; Jiang, J.; Zhai, L.; Yang, C.H.; Zhang, J.; Wang, X.; Chen, D.X.; Niu, H.T.; Chen, L. 5-Aza-dC suppresses melanoma progression by inhibiting GAS5 hypermethylation. *Oncol. Rep.* **2022**, *48*, 123. [[CrossRef](#)]
41. Yang, W.; Xu, X.; Hong, L.; Wang, Q.; Huang, J.; Jiang, L. Upregulation of lncRNA GAS5 inhibits the growth and metastasis of cervical cancer cells. *J. Cell. Physiol.* **2019**, *234*, 23571–23580. [[CrossRef](#)] [[PubMed](#)]
42. Jefferies, H.B.J.; Fumagalli, S.; Dennis, P.B.; Reinhard, C.; Pearson, R.B.; Thomas, G. Rapamycin suppresses 5' TOP mRNA translation through inhibition of p70s6k. *EMBO J.* **1997**, *16*, 3693–3704. [[CrossRef](#)] [[PubMed](#)]
43. Smith, C.M.; Steitz, J.A. Classification of gas5 as a Multi-Small-Nucleolar-RNA (snoRNA) Host Gene and a Member of the 5'-Terminal Oligopyrimidine Gene Family Reveals Common Features of snoRNA Host Genes. *Mol. Cell. Biol.* **1998**, *18*, 6897–6909. [[CrossRef](#)]
44. Mourtada-Maarabouni, M.; Hasan, A.M.; Farzaneh, F.; Williams, G.T. Inhibition of Human T-Cell Proliferation by Mammalian Target of Rapamycin (mTOR) Antagonists Requires Noncoding RNA Growth-Arrest-Specific Transcript 5 (GAS5). *Mol. Pharmacol.* **2010**, *78*, 19–28. [[CrossRef](#)] [[PubMed](#)]
45. Yacqub-Usman, K.; Pickard, M.R.; Williams, G.T. Reciprocal regulation of GAS5 lncRNA levels and mTOR inhibitor action in prostate cancer cells. *Prostate* **2015**, *75*, 693–705. [[CrossRef](#)]
46. Pickard, M.R.; Williams, G.T. Regulation of apoptosis by long non-coding RNA GAS5 in breast cancer cells: Implications for chemotherapy. *Breast Cancer Res. Treat.* **2014**, *145*, 359–370. [[CrossRef](#)]
47. Pei, J.; Wang, B. Notch-1 promotes breast cancer cells proliferation by regulating lncRNA GAS5. *Int. J. Clin. Exp. Med.* **2015**, *8*, 14464–14471.
48. Tokgun, P.E.; Tokgun, O.; Kurt, S.; Tomatir, A.G.; Akca, H. MYC-driven regulation of long non-coding RNA profiles in breast cancer cells. *Gene* **2019**, *714*, 143955. [[CrossRef](#)]
49. Zhang, Z.; Zhu, Z.; Watabe, K.; Zhang, X.; Bai, C.; Xu, M.; Wu, F.; Mo, Y.Y. Negative regulation of lncRNA GAS5 by miR-21. *Cell Death Differ.* **2013**, *20*, 1558–1568. [[CrossRef](#)]
50. Chen, Z.; Pan, T.; Jiang, D.; Jin, L.; Geng, Y.; Feng, X.; Shen, A.; Zhang, L. The lncRNA-GAS5/miR-221-3p/DKK2 Axis Modulates ABCB1-Mediated Adriamycin Resistance of Breast Cancer via the Wnt/ $\beta$ -Catenin Signaling Pathway. *Mol. Ther. Nucleic Acids* **2020**, *19*, 1434–1448. [[CrossRef](#)]
51. Li, Y.; Guo, X.B.; Wei, Y.H. lncRNA GAS5 affects epithelial-mesenchymal transition and invasion of breast cancer cells by regulating miR-216b. *Eur. Rev. Med. Pharmacol. Sci.* **2020**, *24*, 4873–4881. [[PubMed](#)]
52. Kaur, J.; Salehen, N.; Norazit, A.; Rahman, A.A.; Murad, N.A.A.; Rahman, N.M.A.N.A.; Ibrahim, K. Tumor Suppressive Effects of GAS5 in Cancer Cells. *Non-Coding RNA* **2022**, *8*, 39. [[CrossRef](#)] [[PubMed](#)]
53. Mourtada-Maarabouni, M.; Pickard, M.R.; Hedge, V.L.; Farzaneh, F.; Williams, G.T. GAS5, a non-protein-coding RNA, controls apoptosis and is downregulated in breast cancer. *Oncogene* **2009**, *28*, 195–208. [[CrossRef](#)] [[PubMed](#)]
54. Jiang, Y.; Qian, T.; Li, S.; Xie, Y.; Tao, M. Metformin reverses tamoxifen resistance through the lncRNA GAS5-mediated mTOR pathway in breast cancer. *Ann. Transl. Med.* **2022**, *10*, 366. [[CrossRef](#)] [[PubMed](#)]
55. Esmatabadi, M.J.D.; Motamedrad, M.; Sadeghizadeh, M. Down-regulation of lncRNA, GAS5 decreases chemotherapeutic effect of dendrosomal curcumin (DNC) in breast cancer cells. *Phytomedicine* **2018**, *42*, 56–65. [[CrossRef](#)]
56. Liu, K.; Gao, L.; Ma, X.; Huang, J.J.; Chen, J.; Zeng, L.; Ashby, C.R.; Zou, C.; Chen, Z.S. Long non-coding RNAs regulate drug resistance in cancer. *Mol. Cancer* **2020**, *19*, 54. [[CrossRef](#)]
57. Filippova, E.A.; Fridman, M.V.; Burdennyy, A.M.; Loginov, V.I.; Pronina, I.V.; Lukina, S.S.; Dmitriev, A.A.; Braga, E.A. Long Noncoding RNA GAS5 in Breast Cancer: Epigenetic Mechanisms and Biological Functions. *Int. J. Mol. Sci.* **2021**, *22*, 6810. [[CrossRef](#)]
58. Waks, A.G.; Winer, E.P. Breast Cancer Treatment: A Review. *JAMA* **2019**, *321*, 288–300. [[CrossRef](#)]
59. Haussmann, J.; Corradini, S.; Nestle-Kraemling, C.; Bölke, E.; Njanang, F.J.D.; Tamaskovics, B.; Orth, K.; Ruckhaeberle, E.; Fehm, T.; Mohrmann, S.; et al. Recent advances in radiotherapy of breast cancer. *Radiat. Oncol.* **2020**, *15*, 71. [[CrossRef](#)]
60. Ma, Y.; Yu, L.; Yan, W.; Qiu, L.; Zhang, J.; Jia, X. lncRNA GAS5 Sensitizes Breast Cancer Cells to Ionizing Radiation by Inhibiting DNA Repair. *Biomed. Res. Int.* **2022**, *2022*, 1987519. [[CrossRef](#)]
61. Liang, W.; Lv, T.; Shi, X.; Liu, H.; Zhu, Q.; Zeng, J.; Yang, W.; Yin, J.; Song, Y. Circulating long noncoding RNA GAS5 is a novel biomarker for the diagnosis of nonsmall cell lung cancer. *Medecine* **2016**, *95*, e4608. [[CrossRef](#)] [[PubMed](#)]

62. Li, C.; Lv, Y.; Shao, C.; Chen, C.; Zhang, T.; Wei, Y.; Fan, H.; Lv, T.; Liu, H.; Song, Y. Tumor-derived exosomal lncRNA GAS5 as a biomarker for early-stage non-small-cell lung cancer diagnosis. *J. Cell. Physiol.* **2019**, *234*, 20721–20727. [[CrossRef](#)]
63. Fayda, M.; Isin, M.; Tambas, M.; Guveli, M.; Meral, R.; Altun, M.; Sahin, D.; Ozkan, G.; Sanli, Y.; Isin, H.; et al. Do circulating long non-coding RNAs (lncRNAs) (LincRNA-p21, GAS 5, HOTAIR) predict the treatment response in patients with head and neck cancer treated with chemoradiotherapy? *Tumor Biol.* **2016**, *37*, 3969–3978. [[CrossRef](#)] [[PubMed](#)]
64. Kresoja-Rakic, J.; Szpechcinski, A.; Kirschner, M.B.; Ronner, M.; Minatel, B.; Martinez, V.D.; Lam, W.L.; Weder, W.; Stahel, R.; Früh, M.; et al. miR-625-3p and lncRNA GAS5 in Liquid Biopsies for Predicting the Outcome of Malignant Pleural Mesothelioma Patients Treated with Neo-Adjuvant Chemotherapy and Surgery. *Non-Coding RNA* **2019**, *5*, 41. [[CrossRef](#)] [[PubMed](#)]
65. Manganelli, M.; Grossi, I.; Ferracin, M.; Guerriero, P.; Negrini, M.; Ghidini, M.; Senti, C.; Ratti, M.; Pizzo, C.; Passalacqua, R.; et al. Longitudinal circulating levels of mir-23b-3p, mir-126-3p and lncrna gas5 in hcc patients treated with sorafenib. *Biomedicines* **2021**, *9*, 813. [[CrossRef](#)]
66. Han, L.; Ma, P.; Liu, S.M.; Zhou, X. Circulating long noncoding RNA GAS5 as a potential biomarker in breast cancer for assessing the surgical effects. *Tumor Biol.* **2016**, *37*, 6847–6854. [[CrossRef](#)]
67. Toraih, E.A.; El-Wazir, A.; Al Ageeli, E.; Hussein, M.H.; Eltoukhy, M.M.; Killackey, M.T.; Kandil, E.; Fawzy, M.S. Unleash multifunctional role of long noncoding RNAs biomarker panel in breast cancer: A predictor classification model. *Epigenomics* **2020**, *12*, 1215–1237. [[CrossRef](#)]
68. Manganelli, M.; Grossi, I.; Corsi, J.; D’Agostino, V.G.; Jurikova, K.; Cusanelli, E.; Molfino, S.; Portolani, N.; Salvi, A.; De Petro, G. Expression of Cellular and Extracellular TERRA, TERC and TERT in Hepatocellular Carcinoma. *Int. J. Mol. Sci.* **2022**, *23*, 6183. [[CrossRef](#)]
69. Chen, X.; Xue, S.; Ren, G.; Li, Y.; Chao, W.; Chen, S.; Li, X. Level of lncRNA GAS5 and Hippocampal Volume are Associated with the Progression of Alzheimer’s Disease. *Clin. Interv. Aging* **2022**, *17*, 745–753. [[CrossRef](#)]
70. Peng, T.; Ji, D.; Jiang, Y. Long non-coding RNA GAS5 suppresses rheumatoid arthritis progression via miR-128-3p/HDAC4 axis. *Mol. Cell. Biochem.* **2021**, *476*, 2491–2501. [[CrossRef](#)]
71. Song, J.; Ahn, C.; Chun, C.H.; Jin, E.J. A long non-coding RNA, GAS5, plays a critical role in the regulation of miR-21 during osteoarthritis. *J. Orthop. Res.* **2014**, *32*, 1628–1635. [[CrossRef](#)] [[PubMed](#)]
72. Zheng, D.; Hou, Y.; Li, Y.; Bian, Y.; Khan, M.; Li, F.; Huang, L.; Qiao, C. Long Non-coding RNA Gas5 Is Associated With Preeclampsia and Regulates Biological Behaviors of Trophoblast via MicroRNA-21. *Front. Genet.* **2020**, *11*, 188. [[CrossRef](#)] [[PubMed](#)]
73. Winkle, M.; El-Daly, S.M.; Fabbri, M.; Calin, G.A. Noncoding RNA therapeutics—Challenges and potential solutions. *Nat. Rev. Drug Discov.* **2021**, *20*, 629–651. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.